Chromatin structure and 3C-like data

Davide Baù & François Serra

Genome Biology Group (CNAG) Structural Genomics Group (CRG)



The role of chromatin structure

It can give insights into how distant genomic elements interacts with each other

It helps to understand the compartmentalization of chromosomes within the nucleus

It is essential to understand the mechanisms that regulate the cell











Chromatin definition

Chromatin is composed of DNA complexed with histones and other proteins

Chromatin formation enables the genome to be hierarchically packaged or condensed so that it can fit inside the nuclear space

The compaction allows to modulate gene transcription, DNA repair, recombination, and replication

Chromatin structure is considered highly dynamic



Resolution gap





Chromatin structures





The nuclear organization of DNA



Adapted from Richard E. Ballermann, 2012



The nucleosome







Histone modification effects

Type of modification	H3K4	Н3К9	H3K14	H3K27	H3K79	H4K20	H2BK5
mono- methylation	activation	activation		activation	activation	activation	activation
di-methylation	activation	repression		repression	activation		
tri-methylation	activation	repression		repression	activation, repression		repression
acetylation		activation	activation				



The chromatin compaction levels

Several nucleosomes in a row form what is often referred to as a beads-on-a-string fiber (the 11 nm fiber)

When histones H1 or H5, referred to as linker histones, are added to the 11-nm fiber, the condensed 30 nm fiber is formed

The 30 nm fibers form the next level of compaction by forming loops







The chromatin compaction levels



Adapted from Annu. Rev. Genomics Hum. Genet. 2012, 13:59-82



Euchromatin and heterochromatin

Electron microscopy



Euchromatin:

chromatin that is located away from the nuclear lamina, is generally less densely packed, and contains actively transcribed genes

Heterochromatin:

chromatin that is near the nuclear lamina, tightly condensed, and transcriptionally silent



Complex genome organization

Takizawa, T., Meaburn, K. J. & Misteli, T. The meaning of gene positioning. Cell 135, 9–13 (2008).





Lamina-genome interactions



Most genes in Lamina Associated Domains are transcriptionally silent, suggesting that **lamina-genome interactions** are widely involved in the control of **gene expression**

Adapted from Molecular Cell 38, 603-613, 2010



Complex genome organization

Cavalli, G. & Misteli, T. Functional implications of genome topology. Nat Struct Mol Biol 20, 290–299 (2013).





Chromatin loops



Loops bring distal genomic regions in close proximity to one another

This in turn can have profound effects on gene transcription

Enhancers can be thousands of kilobases away from their target genes in any direction (or even on a separate chromosome)



Main approaches

Light microscopy (FISH)



Cell/molecular biology (3C-based methods)





Restrain based modeling (IMP)











Job Dekker



Dostie et al. Genome Res (2006) vol. 16 (10) pp. 1299-309



Hi-L data and genomic tracks data

Dekker, J., Marti-Renom, M. A. & Mirny, L. A. Exploring the three-dimensional organization of genomes: interpreting chromatin interaction data. Nat Rev Genet 14, 390–403 (2013).





Complex genome organization

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Topologically Associating Domains (TADs)



Topologically associating domains (TADs) can be made of up to hundreds of kb in size

Loci located within TADs tend to interact more frequently with each other than with loci located outside their domain

The human and mouse genomes are each composed of over 2,000 TADs, covering over 90% of the genome



Human a-globin domain

ENm008 genomic structure and environment

ENCODE Consortium. Nature (2007) vol. 447 (7146) pp. 799-816



Toy interaction matrix





Toy interaction matrix



cnag 🦓 🚆

Real interaction matrix





Real interaction matrix







Ter

Take home message

Chromatin = DNA + (histone) proteins

The genome is well organized and hierarchically packaged

Histone modifications affect chromatin structure and activity

3C-like data measure the frequency of interaction between distant loci



How DNA is packaged





The Integrative Modeling Platform

http://integrativemodeling.org



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IMP, the Integrative Modeling Platform

IMP's broad goal is to contribute to a comprehensive structural characterization of biomolecules ranging in size and complexity from small peptides to large macromolecular assemblies, by integrating data from diverse biochemical and biophysical experiments. IMP provides an open source C++ and Python toolbox for solving complex modeling problems, and a number of applications for tackling some common problems in a user-friendly way. IMP can also be used from the Chimera molecular modeling system, or via one of several web applications.

IMP is open source software, mostly available under the terms of the GNU Lesser General Public License (LGPL). (Some IMP modules are available under the GNU GPL instead.)

Get started with IMP by downloading it and checking out the documentation.





The IMP software is used as part of the National Center for Dynamic Interactome Research (NCDIR).

If you use IMP, please cite D. Russel, K. Lasker, B. Webb, D. Schneidman, J. Velázquez-Muriel, A. Sali, "Putting the pieces together: integrative structure determination of macromolecular assemblies", PLoS Biology, 2012. The main page of each IMP module in the documentation also lists publications relevant to that module.



Installing IMP

Install the required libraries:

sudo apt-get install cmake sudo apt-get install libboost1.49-all-dev sudo apt-get install libhdf5-dev sudo apt-get install swig sudo apt-get install libcgal-dev sudo apt-get install python-dev

Download the IMP tarball file from http://salilab.org/imp/ and uncompress it:

wget <u>http://salilab.org/imp/get.php?pkg=2.0.1/download/imp-2.0.1.tar.gz</u> -0 imp-2.0.1.tar.gz tar xzvf imp-2.0.1.tar.gz

Move to the IMP directory and compile the code



Compiling IMP

cd imp-2.0.1 cmake . -DCMAKE_BUILD_TYPE=Release -DIMP_MAX_CHECKS=NONE -DIMP_MAX_LOG=SILENT make -j4

Once the compilation has finished, open the file setup_environment.sh in your IMP directory and copy the first lines into your ~/.bashrc file (if this file in not present in your home directory, create it). These lines should look like:

LD_LIBRARY_PATH="/SOMETHING/imp-2.0.1/lib:/SOMETHING/imp-2.0.1/lib:/SOMETHING/imp-2.0.1/src/ dependency/RMF/:\$LD_LIBRARY_PATH" export LD_LIBRARY_PATH

PYTHONPATH="/SOMETHING/imp-2.0.1/lib:/SOMETHING/imp-2.0.1/lib:/SOMETHING/imp-2.0.1/src/dependency/ RMF/:\$PYTHONPATH" export PYTHONPATH

>> Do not copy the lines above, copy them from setup_environment.sh,

where SOMETHING is replaced by your real path to IMP <<



Installing Chimera

http://www.cgl.ucsf.edu/chimera/



Quick Links



UCSF CHIMERA

an Extensible Molecular Modeling System

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UCSF Chimera is a highly extensible program for interactive visualization and analysis of molecular structures and related data, including density maps, supramolecular assemblies, sequence alignments, docking results, trajectories, and conformational ensembles. High-quality images and animations can be generated. Chimera includes complete documentation and several tutorials, and can be downloaded free of charge for academic, government, non-profit, and personal use. Chimera is developed by the <u>Resource for Biocomputing</u>. Visualization, and Informatics. funded by the <u>National Institutes of Health</u> (NGMS P41-GM103311).

Feature Highlight

Multiscale Models

The <u>Multiscale Models</u> extension allows Chimera to display large complexes such as <u>virus capsids</u>, <u>ribosomes</u>, and <u>chromatin</u>. It displays the quatemary structure of PDB models and allows subunits to be selected and shown in atomic detail. Matrices are read from PDB files that specify the biological unit. Crystallographic packing can also be shown.

(More features...)



Chimera commands

Align

match.sh #1 #0

Select select #model:particles

Measure

distance #0:1-2 angle #0:1-2

Display

vdwdefine #radius shape tube #0 radius 1 bandLength 3 segmentSubdivisions 10 shape tube #0 rad 1 band 3 seg 10

Surface

molmap #all 80 color transparency

