Comparative Protein Structure Prediction



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Program

Intro to comparative protein structure prediction

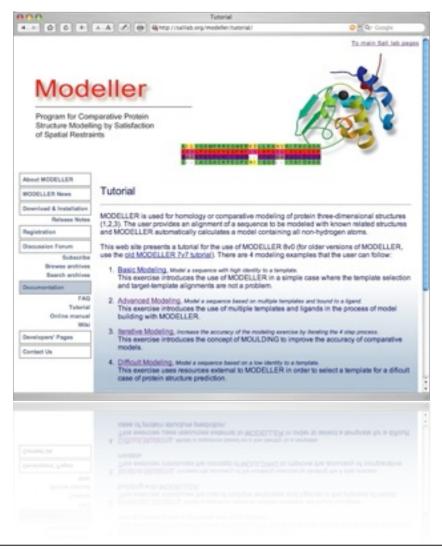
Template Search

Target – Template Alignment

Model Building

Model Evaluation

http://www.salilab.org/modeller/tutotial/



Objective

TO LEARN HOW-TO MODEL A 3D-STRUCTURE FROM A SEQUENCE AND A KNOWN STRUCTURE

DISCLAIMER!

Name	Type#	World Wide Web address	
DATABASES			
CATH	5	http://www.biochem.ucl.ac.uk/bsm/cath/	
DBAII	s	http://www.salilab.org/DBAII/	
GenBank	s	http://www.ncbi.nlm.nih.gov/Genbank/GenbankSearch.html	
GeneCensus	5	http://bioinfo.mbb.yale.edu/genome	
MODBASE	s	http://salilab.org/modbase/	
MSD	s	http://www.rcsb.org/databases.html	
NOSI	s	http://www.ncbi.nlm.nih.gov/	
PDB	8	http://www.rcsb.org/pdb/	
PSI	s	http://www.nigms.nih.gov/psi/	
Sacch3D	8	http://genome-www.stanford.edu/Sacch3D/	
SCOP	s	http://scop.mrc-lmb.cam.ac.uk/scop/	
TIGR	s	http://www.tigr.org/tdb/mdb/mdbcomplete.html	
TrEMBL	5	http://srs.ebi.ac.uk/	
FOLD ASSIGNM	ENT		
123D	s	http://123d.ncifcrf.gov/	
3D-PSSM	8	http://www.sbg.bio.ic.ac.uk/~3dpssm/	
BIOINBGU	s	http://www.cs.bgu.ac.il/~bioinbgu/	
BLAST	s	http://www.ncbl.nlm.nih.gov/BLAST/	
DALI	s	http://www2.ebi.ac.uk/dali/	
FASS	s	http://bioinformatics.burnham-inst.org/FFAS/index.html	
FastA	s	http://www.ebi.ac.uk/fasta3/	
FRSVR	8	http://fold.doe-mbi.ucla.edu/	
FUGUE	s	http://www-cryst.bloc.cam.ac.uk/~fugue/	

http://sgu.bioinfo.cipf.es/home/?page=resources

Programs, servers and databases

http://salilab.org

LS-SNP **PIBASE MODLOOP CCPR** Web Server **Database Web Server Center for Computational** http://salilab.org/LS-SNP http://salilab.org/pibase http://salilab.org/modloop **Proteomics Research** Predicts functional impact Contains structurally defined Models loops in protein http://www.ccpr.ucsf.edu of residue substitution protein interfaces structures **MODBASE DBALI MODWEB MODELLER Database Database Web Server Program** http://salilab.org/modbase http://salilab.org/dbali Fold assignments, alignments http://salilab.org/modweb http://salilab.org/modeller Contains a comprehensive models, model assessments Provides a web interface to Implements most operations set of pairwise and multiple for all sequences related to a **MODPIPE** in comparative modeling structure-based alignments known structure **EVA ICEDB MODPIPE LIGBASE Web Server** Database/LIMS **Database Program** http://salilab.org/eva http://nysgxrc.org Automatically calculates Ligand binding sites and Evaluates and ranks web Tracks targets for structural comparative models of many inheritance (accessible servers for protein structure through MODBASE) protein sequences genomics by NYSGXRC prediction

External Resources

PDB, Uniprot, GENBANK, NR, PIR, INTERPRO, Kinase Resource UCSC Genome Browser, CHIMERA, Pfam, SCOP, CATH

Nomenclature

Homology: Sharing a common ancestor, may have similar or dissimilar functions

Similarity: Score that quantifies the degree of relationship between two sequences.

Identity: Fraction of identical aminoacids between two aligned sequences (case of similarity).

Target: Sequence corresponding to the protein to be modeled.

Template: 3D structure/s to be used during protein structure prediction.

Model: Predicted 3D structure of the target sequence.

Nomenclature

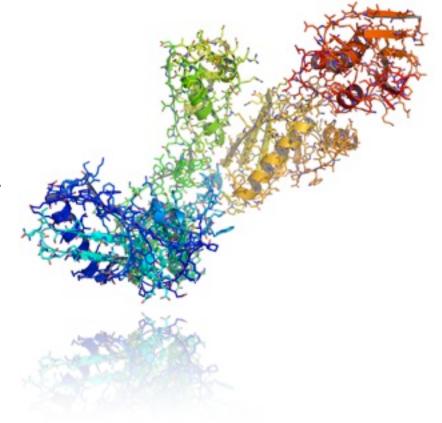
Fold: Three dimensional conformation of a protein sequence (usually at domain level).

Domain: Structurally globular part of a protein, which may independently fold.

Secondary Structure: Regular subdomain structures composed by alphahelices, beta-sheets and coils (or loops).

Backbone: Protein structure skeleton composed by the carbon, nitrogen and oxygen atoms.

Side-Chain: Specific atoms identifying each of the 20 residues types.



General References

Protein Structure Prediction:

Marti-Renom et al. Annu. Rev. Biophys. Biomol. Struct. 29, 291-325, 2000. Baker & Sali. Science 294, 93-96, 2001.

Comparative Modeling:

Madhusudhan et al. The Proteomics Protocols Handbook. Ed. Walker. Humana Press Inc., Totowa, NJ. 831-860, 2005.

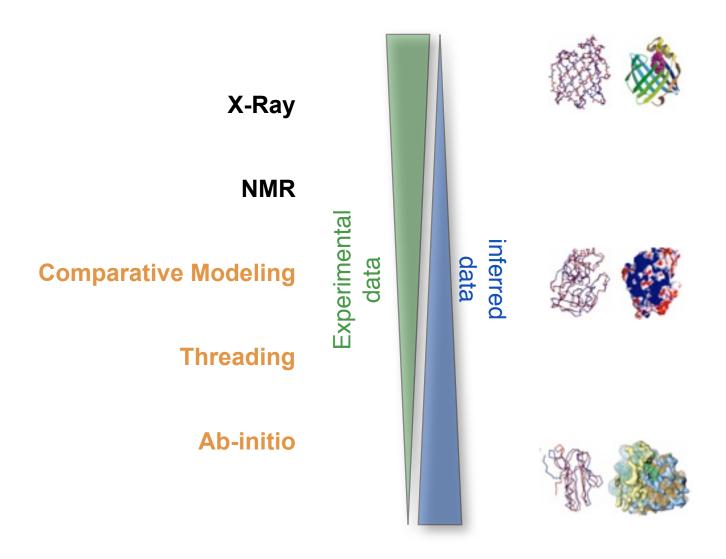
MODELLER:

Sali & Blundell. J. Mol. Biol. 234, 779-815, 1993.
Eswar, M. A. et al. Comparative Protein Structure Modeling With MODELLER.
Current Protocols in Bioinformatics, John Wiley & Sons, Inc.,
Supplement 15, 5.6.1-5.6.30, 2006.

Structural Genomics:

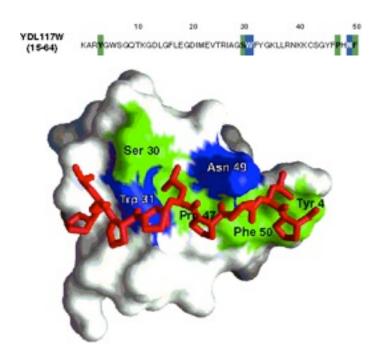
Sali. Nat. Struct. Biol. 5, 1029, 1998. Burley et al. Nat. Genet. 23, 151, 1999. Sali & Kuriyan. TIBS 22, M20, 1999. Sanchez et al. Nat. Str. Biol. 7, 986, 2000. Baker & Sali. Science 294, 93-96, 2001.

protein prediction .vs. protein determination



Why is it useful to know the structure of a protein, not only its sequence?

- The biochemical function (activity) of a protein is defined by its interactions with other molecules.
- The biological function is in large part a consequence of these interactions.
- The 3D structure is more informative than sequence because interactions are determined by residues that are close in space but are frequently distant in sequence.

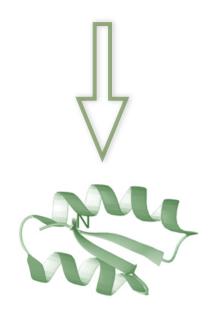


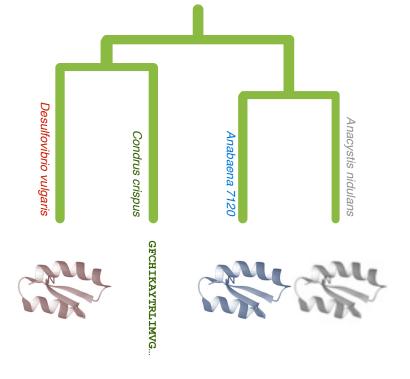
In addition, since evolution tends to conserve function and function depends more directly on structure than on sequence, **structure is more conserved in evolution than sequence**.

The net result is that patterns in space are frequently more recognizable than patterns in sequence.

Principles of protein structure

GFCHIKAYTRLIMVG...





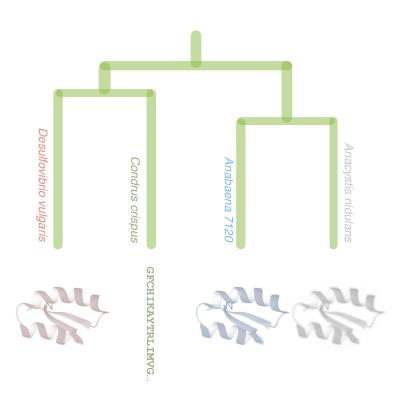
Folding (physics)

Ab initio prediction

Evolution (rules)

Threading Comparative Modeling

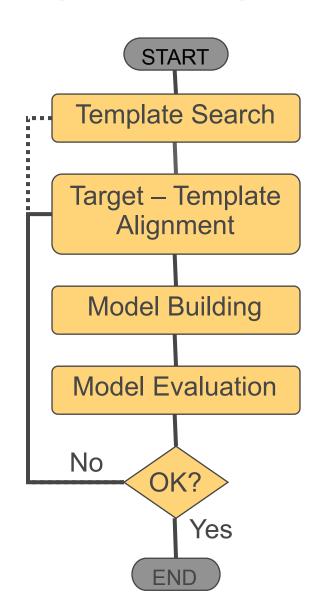
D. Baker & A. Sali. Science 294, 93, 2001.

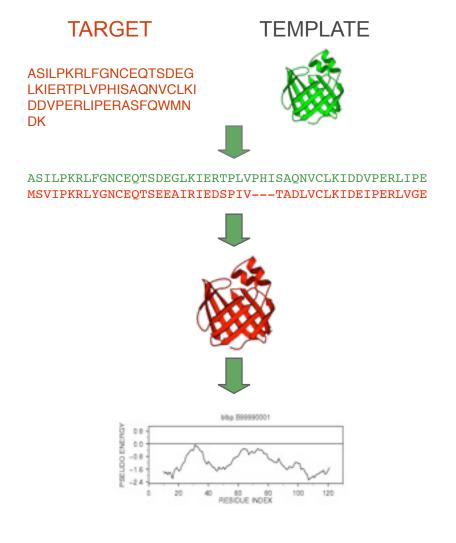


MODELLER

N. Eswar, et al. Comparative Protein Structure Modeling With MODELLER. Current Protocols in Bioinformatics, John Wiley & Sons, Inc., Supplement 15, 5.6.1-5.6.30, 2008.
 M.A. Marti-Renom, et al.. Comparative protein structure modeling of genes and genomes. Annu. Rev. Biophys. Biomol. Struct. 29, 291-325, 2000.
 A. Sali & T.L. Blundell. Comparative protein modelling by satisfaction of spatial restraints. J. Mol. Biol. 234, 779-815, 1993.
 A. Fiser, R.K. Do, & A. Sali. Modeling of loops in protein structures, Protein Science 9. 1753-1773, 2000.

Steps in Comparative Protein Structure Modeling





A. Šali, Curr. Opin. Biotech. 6, 437, 1995. R. Sánchez & A. Šali, Curr. Opin. Str. Biol. 7, 206, 1997. M. Marti et al. Ann. Rev. Biophys. Biomolec. Struct., 29, 291, 2000.

Template Selection

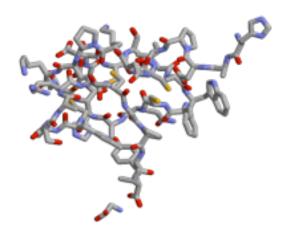
"Structural Space"

Structure-Structure alignments

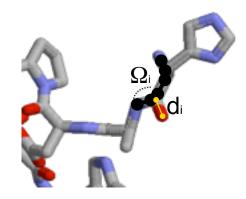
As any other bioinformatics problem...

- Representation
 - Scoring
 - Optimizer

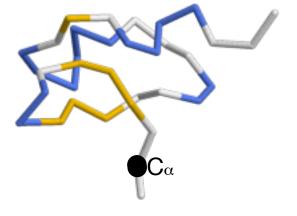
Structures



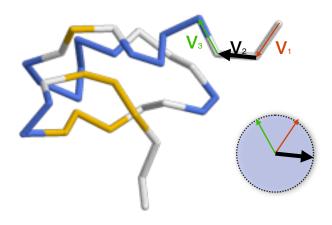
All atoms and coordinates



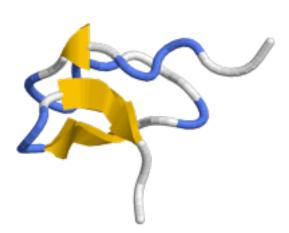
Dihedral space or distance space



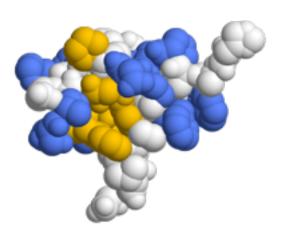
Reduced atom representation



Vector representation



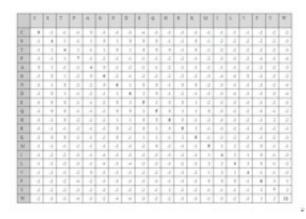
Secondary Structure



Accessible surface (and others)

Scoring

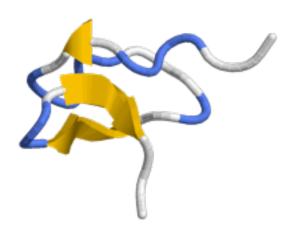
Raw scores

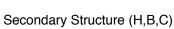


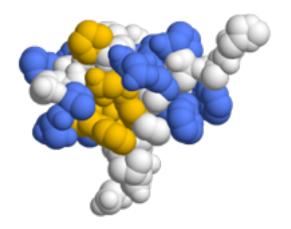
$$RMSD(x,y) = \sqrt{\left(\frac{1}{N}\right)} \sum_{i=1}^{N} (\left\|\mathbf{x}(i) - \mathbf{y}(i)\right\|^{2})$$

Aminoacid substitutions

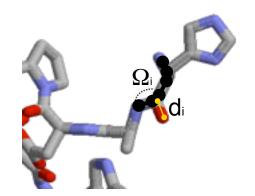
Root Mean Square Deviation







Accessible surface (B,A [%])

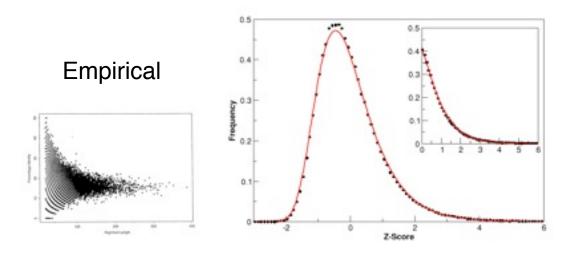


Angles or distances

Scoring

Significance of an alignment (score)

Probability that the optimal alignment of two random sequences/structures of the same length and composition as the aligned sequences/structures have at least as good a score as the evaluated alignment.



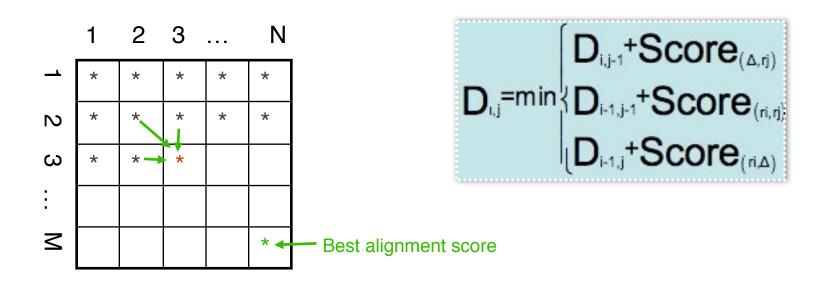
Sometimes approximated by Z-score (normal distribution).

$$P(s \ge x) = 1 - \exp(e^{-\lambda (s - \mu)})$$

Karlin and Altschul, 1990 PNAS 87, pp2264

Global dynamic programming alignment

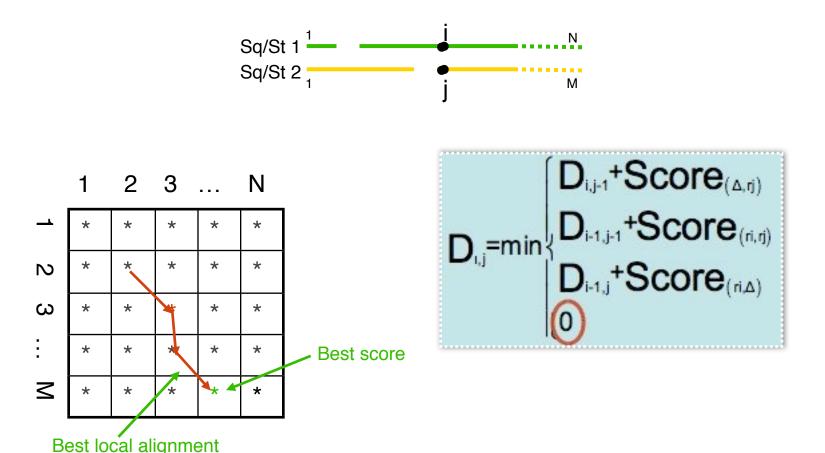




Backtracking to get the best alignment

Needleman and Wunsch (1970) J. Mol Biol, 3 pp443

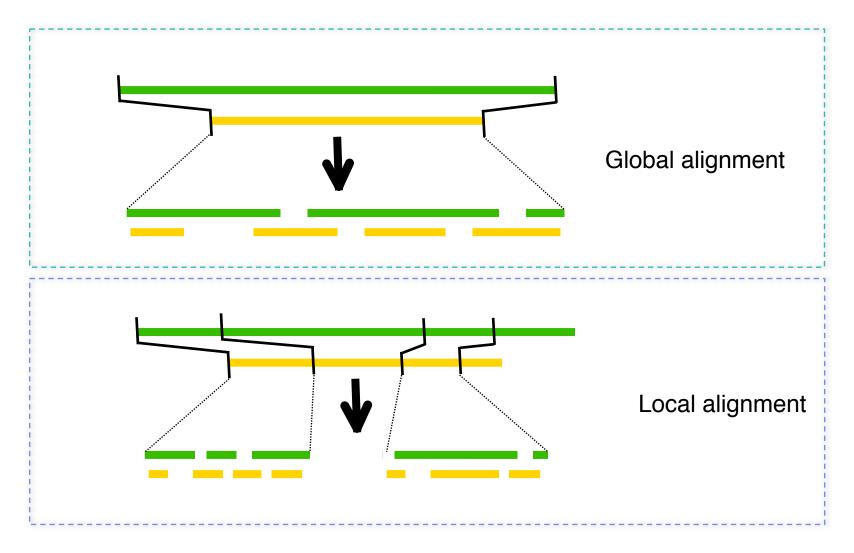
Local dynamic programming alignment



Backtracking to get the best alignment

Smith and Waterman (1981) J. Mol Biol, 147 pp195

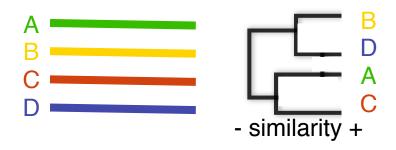
Global .vs. local alignment



Multiple alignment

Pairwise alignments

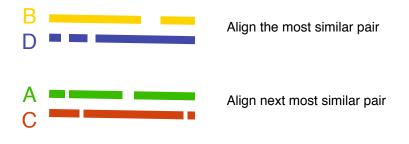
Example – 4 sequences A, B, C, D.



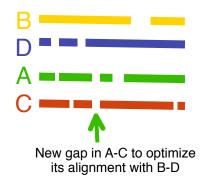
6 pairwise comparisons then cluster analysis

Multiple alignments

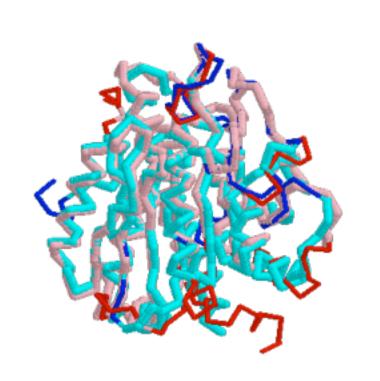
Following the tree from step 1

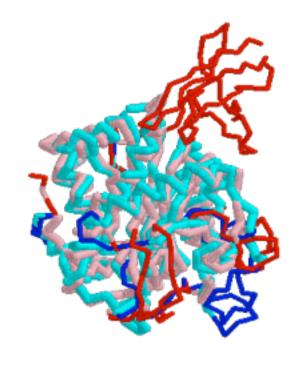


Align B-D with A-C



Coverage .vs. Accuracy



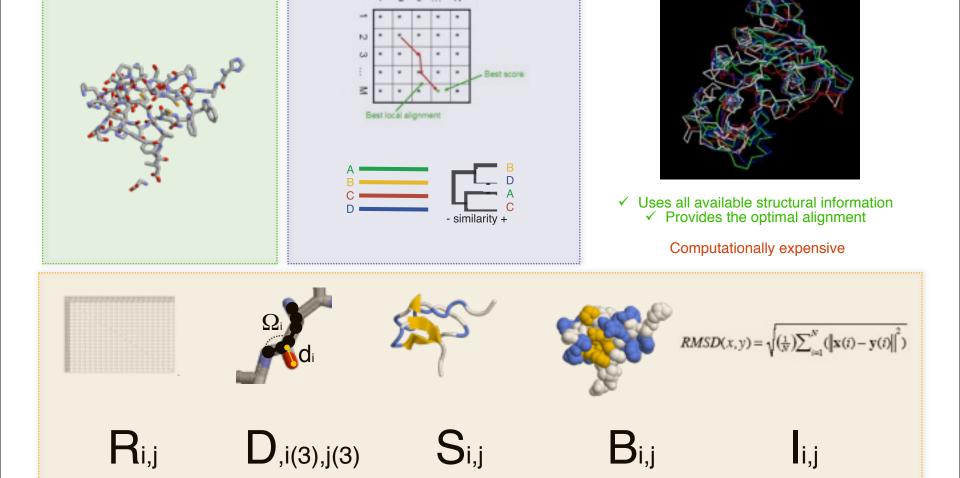


Same RMSD ~ 2.5Å

Coverage ~90% Cα

Coverage ~75% Cα

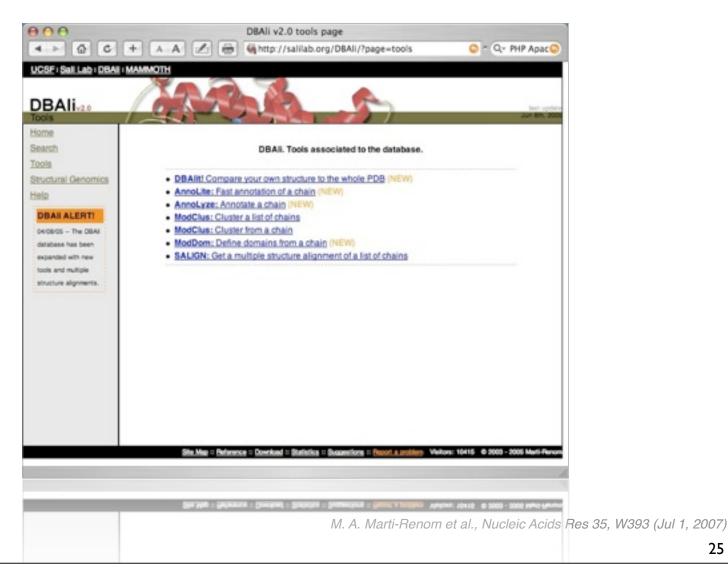
Structural alignment by properties conservation (SALIGN-MODELLER)



M. S. Madhusudhan, B. M. Webb, M. A. Marti-Renom, N. Eswar, A. Sali, Protein Eng Des Sel, (Jul 8, 2009).

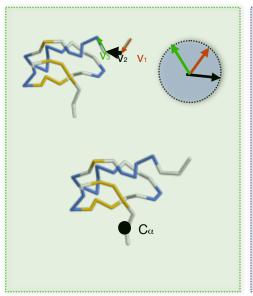
Structural alignment by properties conservation (SALIGN-MODELLER)

http://dbali.org

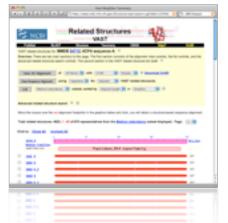


25

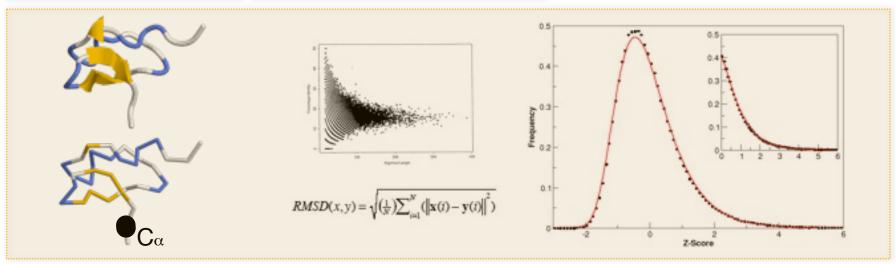
Vector Alignment Search Tool (VAST)



Graph theory search
of similar SSE
Refining by Monte Carlo
at all atom resolution



✓ Good scoring system with significance Reduces the protein representation



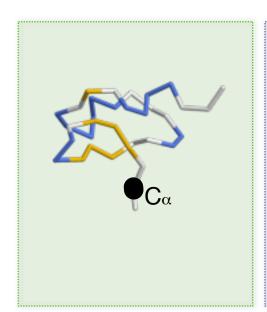
Gibrat JF et al. (1996) Curr Opin Struct Biol 3 pp377

Vector Alignment Search Tool (VAST)

http://www.ncbi.nlm.nih.gov/Structure/VAST/vast.shtml



Incremental combinatorial extension (CE)



Exhaustive combination of fragments

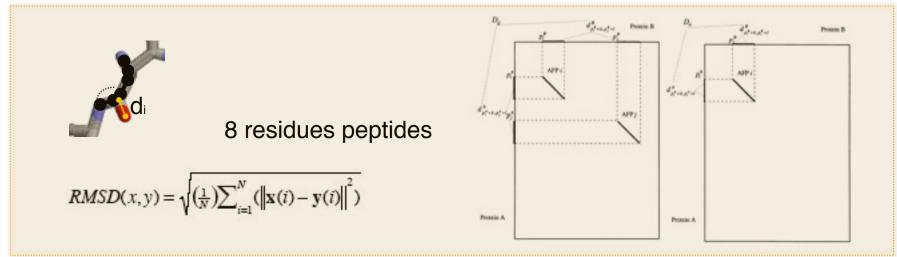
Longest combination of AFPs

Heuristic similar to PSI-BLAST



- ✓ FAST!
- ✓ Good quality of local alignments

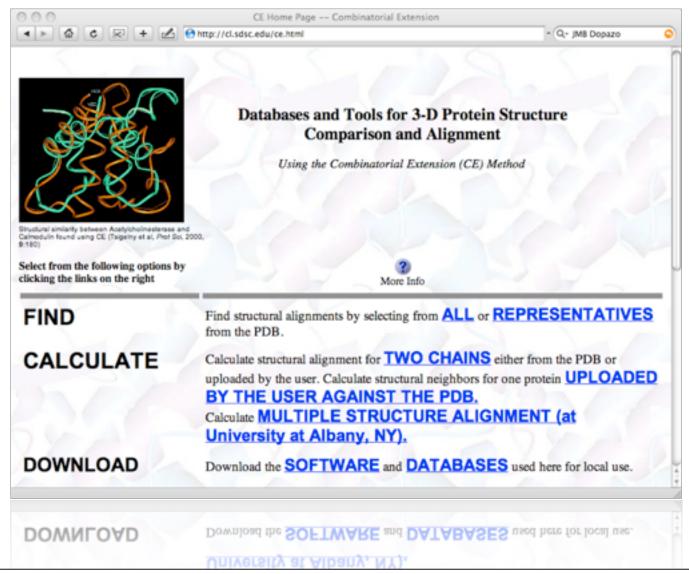
Complicated scoring and heuristics



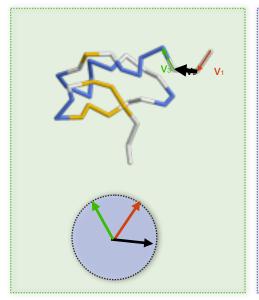
Shindyalov IN, amd Bourne PE. (1998) Protein Eng. 9 pp739

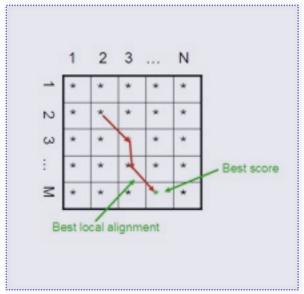
Incremental combinatorial extension (CE)

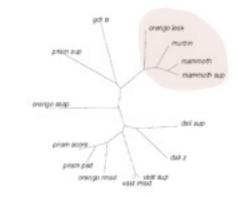
http://cl.sdsc.edu/ce.html



Matching molecular models obtained from theory (MAMMOTH)

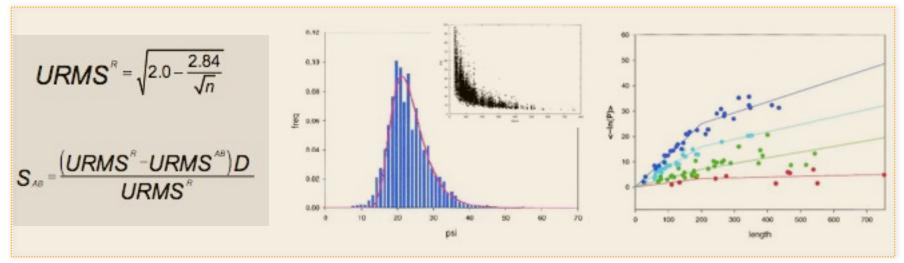






- √ VERY FAST!
- ✓ Good scoring system with significance

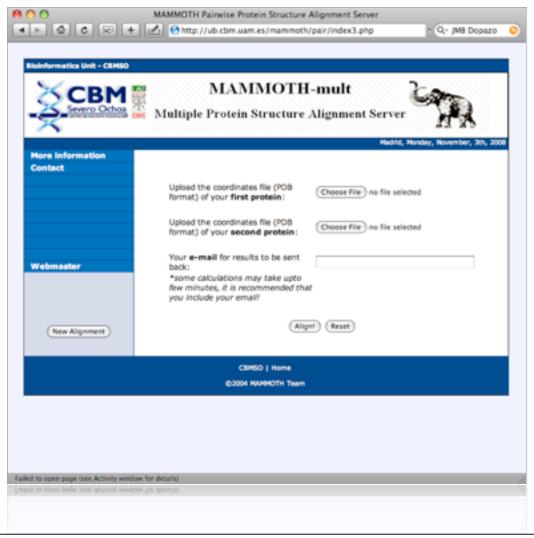
Reduces the protein representation



Ortiz AR, (2002) Protein Sci. 11 pp2606

Matching molecular models obtained from theory (MAMMOTH)

http://ub.cbm.uam.es/mammoth/pair/index3.php



Classification of the structural space

SCOP_{1.75} database

http://scop.mrc-lmb.cam.ac.uk/scop/



- ✓ Largely recognized as "standard of gold"
- ✓ Manually classification
- ✓ Clear classification of structures in:

CLASS FOLD SUPER-FAMILY FAMILY

√ Some large number of tools already available

Manually classification Not 100% up-to-date Domain boundaries definition

Class	Number of folds	Number of superfamilies	Number of families	
All alpha proteins	284	507	871	
All beta proteins	174	354	742	
Alpha and beta proteins (a/b)	147	244	803	
Alpha and beta proteins (a+b)	376	552	1055	
Multi-domain proteins	66	66	89	
Membrane and cell surface proteins	58	110	123	
Small proteins	90	129	219	
Total	1195	1962	3902	

Murzin A. G., el at. (1995). J. Mol. Biol. 247, 536-540.

CATH_{3,2} database

http://www.cathdb.info



Uses FSSP for superimposition

- √ Recognized as "standard of gold"
- √ Semi-automatic classification
- ✓ Clear classification of structures in:

CLASS
ARCHITECTURE
TOPOLOGY
HOMOLOGOUS SUPERFAMILIES

- √ Some large number of tools already available
- √ Easy to navigate

Semi-automatic classification Domain boundaries definition

rchitecture	Topology	Homologous Superfamily	S35 Family	S60 Family	S95 Family	S100 Family	Domains
	310	682	2078	2689	3540	6685	23491
)	196	438	2062	2902	4468	7656	29992
l .	512	956	4558	6473	8135	16346	58967
	92	102	173	217	301	445	1765
)	1110	2178	8871	12281	16444	31132	114215

Total 40

DBAliv2.0 database

http://salilab.org/DBAli/



Uses MAMMOTH for superimposition

- √ Fully-automatic
- ✓ Data is kept up-to-date with PDB releases
- √ Tools for "on the fly" classification of families
- ✓ Up-to-date multiple structure alignments
- √ Easy to navigate
- ✓ Provides some tools for structure comparison

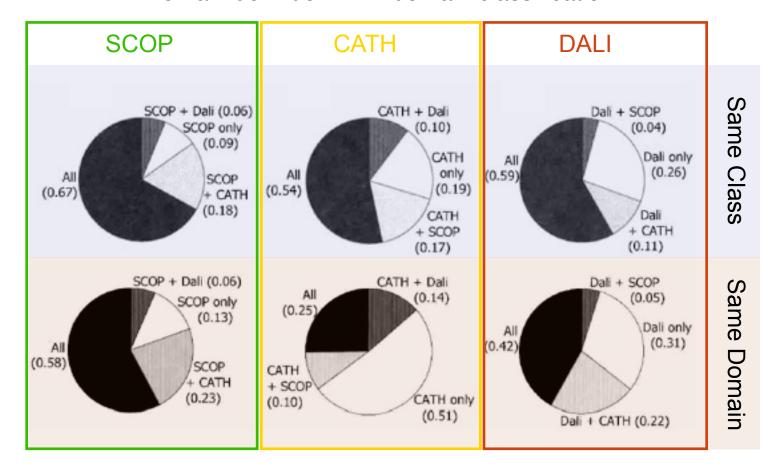
Does not provide a stable classification

Pairwise structure alignments	
Last update:	October 6th, 2007
Number of chains:	96,804
Number of structure-structure comparisons:*	1,748,371,897
Multiple structure alignments	
Last update:	August 1st, 2007
Number of representative chains:	34,637
Number of families:	12,732

Marti-Renom et al. 2001. Bioinformatics. 17, 746 Marti-Renom et al. 2007. BMC BMC Bioinformatics (2007) 8 (Suppl 4) S4 Marti-Renom et al. 2007. Nucleic Acid Research (2007) 35 W393-W397

Classification of the structural space Not an easy task!

Domain definition AND domain classification

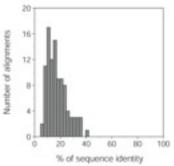


template search and template-target alignment (pp_scan)

Marti-Renom, et al. (2004) Prot. Sci. 13 pp1071 Narayanan, et al. in prepration

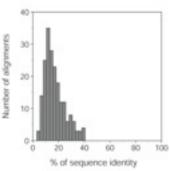
PP_SCAN or profile-profile alignments

A) Traning Set





Number of alignment



Seq.-Seq

ALIGN: DP pairwise method

BLAST2SEQ: Local heuristic method

Seq.-Str

SEA: Local structure prediction method

Prof.-Seq

SAM: HMM method

PSI-BLAST: Local search method that

uses multiple sequence information for one of the

sequences.

LOBSTER: HHM + Phylogeny Method

Number of alignments RSMD

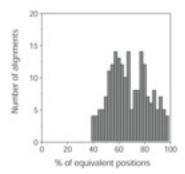
Prof.-Prof

CLUSTALW: DP multiple sequence

method.

COMPASS: DP profile-profile method

Number of alignments % of equivalent positions



RSMD

PP_SCAN: DP pairwise method that uses multiple sequence information for both sequences.

PP_SCAN protocols

Profile generation

- PSI-Blast (PBP)
- Henikoff & Henikoff (HH)
- Henikoff & Henikoff + Similarity (HS)
- Henikoff & Henikoff substitution matrix (MAT)

Profile comparison

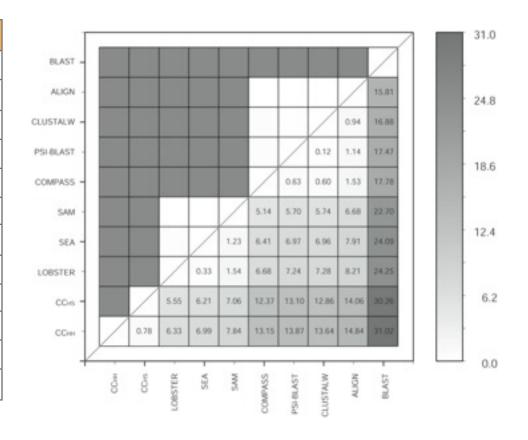
- Correlation coefficient (CC)
- Euclidean distance (ED)
- Dot product (DP)
- Jensen-Shannon distance (JS)
- Average value (Ave)

PP_SCAN protocols accuracy

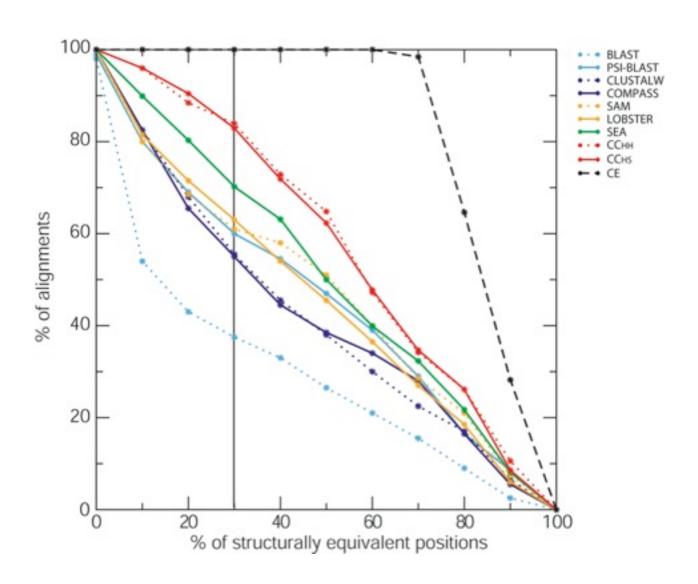
SALIGN protocol	CE overlap [%]	Shift score
ССРВР	55 ± 23	0.61 ± 0.24
ССнн	56 ± 23	0.61 ± 0.24
ССнѕ	56 ± 24	0.62 ± 0.23
ССмат	51 ± 25	0.55 ± 0.27
ЕДРВР	54 ± 24	0.60 ± 0.25
ЕДнн	54 ± 24	0.59 ± 0.26
EDHs	55 ± 24	0.59 ± 0.26
DР _{РВР}	55 ± 23	0.61 ± 0.24
DРнн	56 ± 23	0.60 ± 0.25
DPHS	55 ± 24	0.61 ± 0.24
JSнн	53 ± 24	0.60 ± 0.24
JS HS	54 ± 24	0.60 ± 0.24
Avemat	49 ± 26	0.52 ± 0.29
ТОР	62 ± 20	0.67 ± 0.20

PP_SCAN accuracy

Method	CE overlap	Shift score			
CE	100 ±0	1.00 ± 0.00			
BLAST	26 ± 29	0.32 ± 0.33			
PSI-BLAST	43 ± 31	0.48 ± 0.35			
SAM	48 ± 26	0.50 ± 0.34			
LOBSTER	50 ± 27	0.51 ± 0.32			
SEA	49 ± 27	0.53 ± 0.29			
ALIGN	42 ± 25	0.44 ± 0.28			
CLUSTALW	43 ± 27	0.44 ± 0.31			
COMPASS	43 ± 32	0.49 ± 0.35			
ССнн	56 ± 23	0.61 ± 0.24			
ССнѕ	56 ± 24	0.62 ± 0.24			
ТОР	62 ± 20	0.67 ± 0.20			



PP_SCAN success



Alignment accuracy (CE overlap)

200 pairwise DBAli alignments

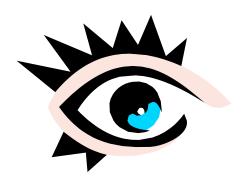
PSI-BLAST (sequence-profile alignment)	43%
SEA (local structure alignment)	49%
PP SCAN (profile-profile alignment)	56%



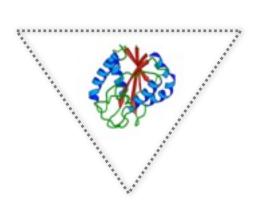
model building and model assessment

Information about a protein can come from three distinct sources

http://www.integrativemodeling.org/

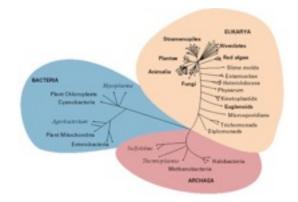


Experimental observations



STATE OF THE STATE

Laws of physics

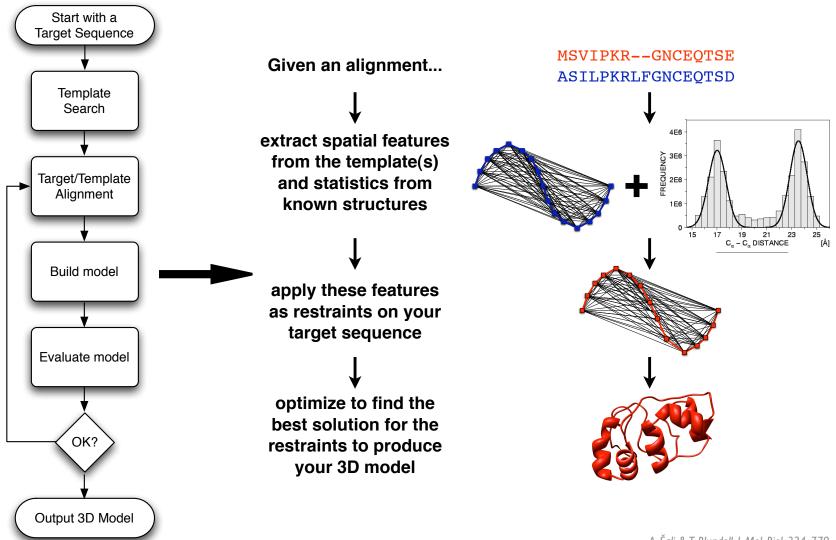


Statistical rules

Classes of methods for comparative protein structure modeling

- Model building by assembly of rigid bodies core, loops, side-chains.
- Model building by segment matching.
- Model building by satisfaction of spatial restraints.

Comparative modeling by satisfaction of spatial restraints MODELLER



A. Šali & T. Blundell. J. Mol. Biol. 234, 779, 1993. J.P. Overington & A. Šali. Prot. Sci. 3, 1582, 1994. A. Fiser, R. Do & A. Šali, Prot. Sci., 9, 1753, 2000.

Multiple Templates

Local similarity extracted from closest template



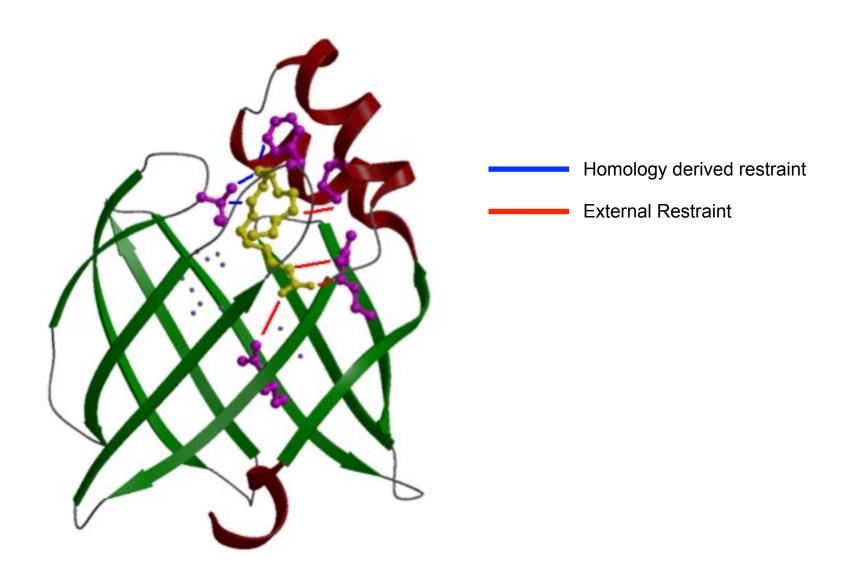
Templates

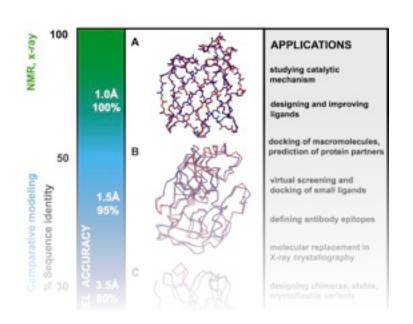
KSINPIHGDNCEOTSDEGLKIERTPL----OWLKSSICDMRGLIPE

Target

ASILPKRLFGNCEQTSDEGLKIERTPLVPHISAQNVCLKIDDVPERLIPE MSVIPKRLYGNCEOTSEEAIRIEDSPIVRWISAOLVCLKIDEIPERLVGE

Modeling ligands and using external restraints





Accuracy and applicability of comparative models

Comparative modeling by satisfaction of spatial restraints Types of errors and their impact



Wrong fold



Miss alignments



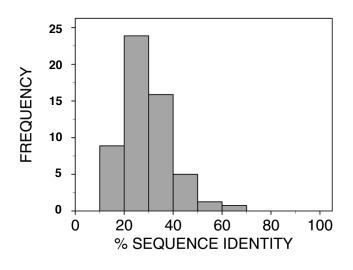
Loop regions

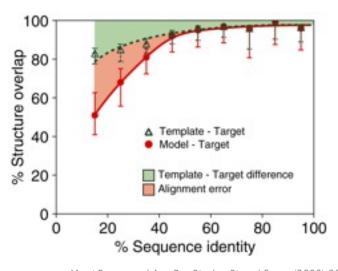


Rigid body distortions



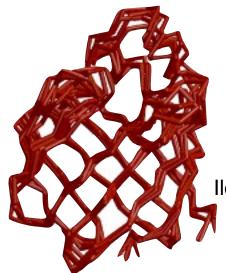
Side-chain packing





Marti-Renom etal. Ann Rev Biophys Biomol Struct (2000) 29, 291

"Biological" significance of modeling errors



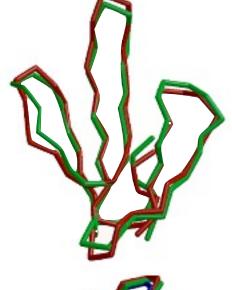
NMR - X-RAY

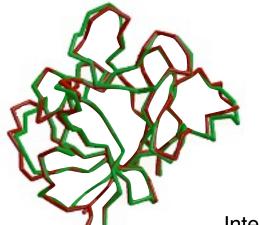
Erabutoxin 3ebx Erabutoxin 1era



Ileal lipid-binding protein

1eal

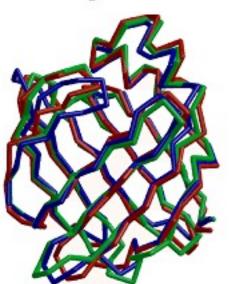




CRABPII 1opbB FABP 1ftpA ALBP 1lib 40% seq. id.



Interleukin 1β 41bi (2.9Å) Interleukin 1β 2mib (2.8Å)

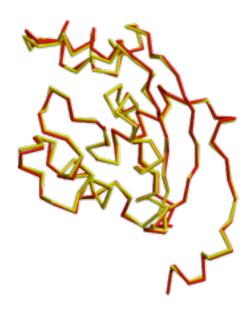


Model Accuracy

HIGH ACCURACY

NM23 Seq id 77%

Cα equiv 147/148 RMSD 0.41Å

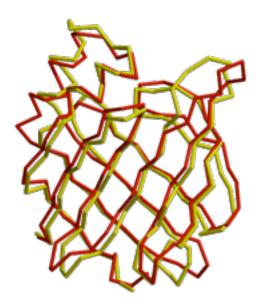


Sidechains Core backbone Loops

MEDIUM ACCURACY

CRABP Seq id 41%

Cα equiv 122/137 RMSD 1.34Å



Sidechains Core backbone Loops Alignment

LOW ACCURACY

EDN Seq id 33%

 $C\alpha$ equiv 90/134 RMSD 1.17Å

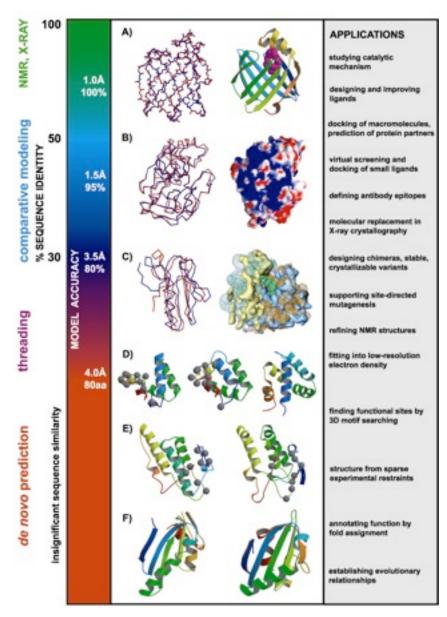


Sidechains
Core backbone
Loops
Alignment
Fold assignment

X-RAY / MODEL

Marti-Renom et al. Annu. Rev. Biophys. Biomol. Struct. 29, 291-325, 2000.

Utility of protein structure models, despite errors



Model Assessment (PMF)

Scoring

Statistical Potential (inspiration)

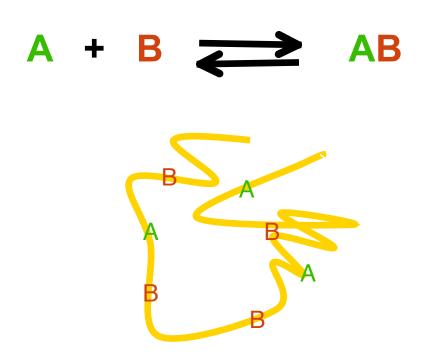
$$K = \frac{\begin{bmatrix} AB \end{bmatrix}}{\begin{bmatrix} A \end{bmatrix} \cdot \begin{bmatrix} B \end{bmatrix}}$$

$$\Delta G = -RT \ln(K) = -RT \ln \frac{\begin{bmatrix} AB \end{bmatrix}}{\begin{bmatrix} A \end{bmatrix} \cdot \begin{bmatrix} B \end{bmatrix}}$$

From statistical physics, we know that energy difference between two states (ΔE) and the ratio of their occupancies ($N_1:N_2$) are related [9]:

$$\Delta E = -kT \ln \left(\frac{N_1}{N_2} \right)$$
 (1)

in which T is the absolute temperature and k is the Boltzmann's constant. As we are interested in an interaction energy between two amino acid side chains, it would seem natural to define N₁ as the number of interactions between these two residues types in a group of real protein structures, a number which is readily available from simple database analysis. But this number must be compared with the number of interactions in some other system, N₂, to obtain the energy difference between them.



Tanaka and Sheraga (1975) PNAS, **72** pp3802 Sippl, (1990) J.Mo.Biol. **213** pp859 Godzik, (1996) Structure **15** pp363

Scoring

Significance of an alignment (score)

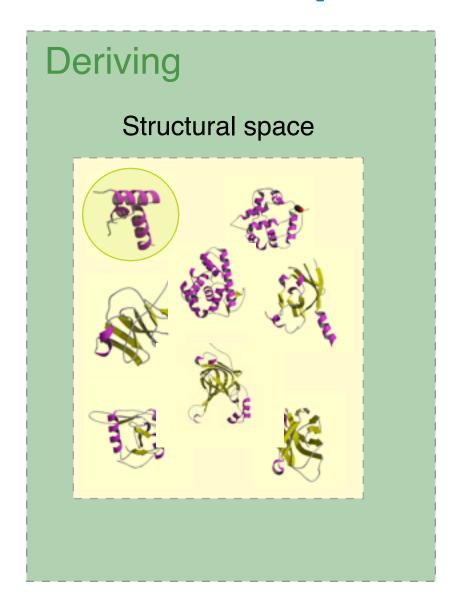
Energy Z-score the model with respect the energy of random models (or rest of decoys).

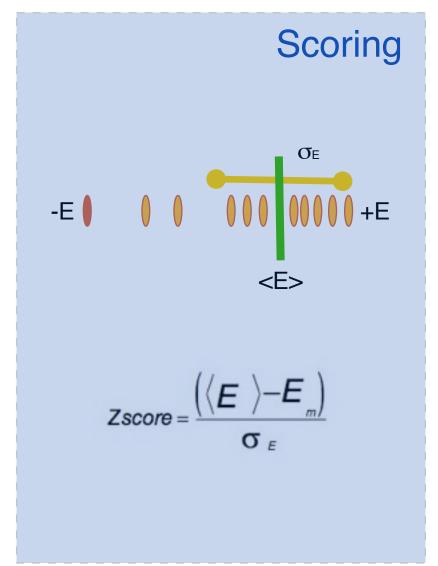
-E |
$$Zscore = \frac{\langle E \rangle - E_{m}}{\sigma_{E}}$$

Zscore =
$$\frac{\left\langle\!\left\langle E\right\rangle\!-\!E_{_{m}}\right\rangle}{\sigma_{_{E}}}$$

Prosall

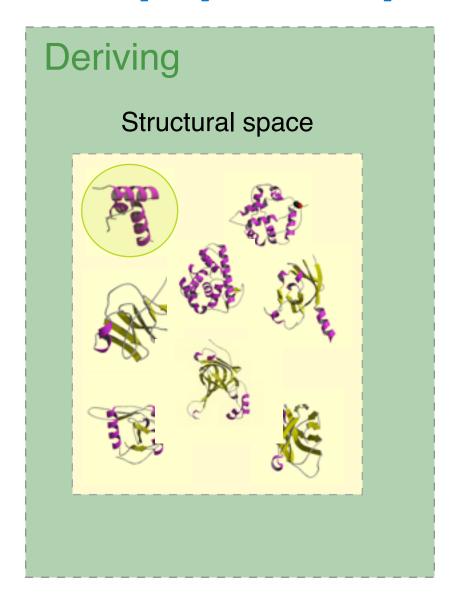
http://www.came.sbg.ac.at

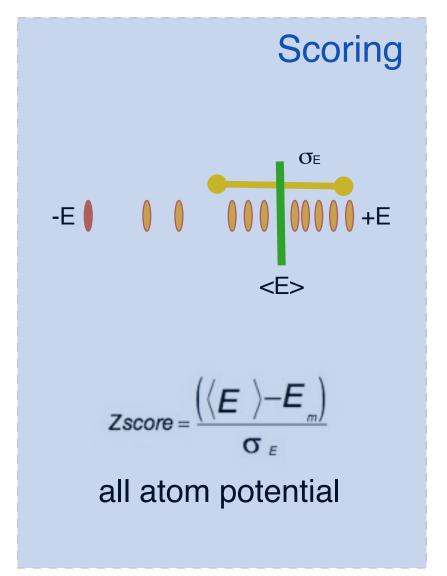




ANOLEA

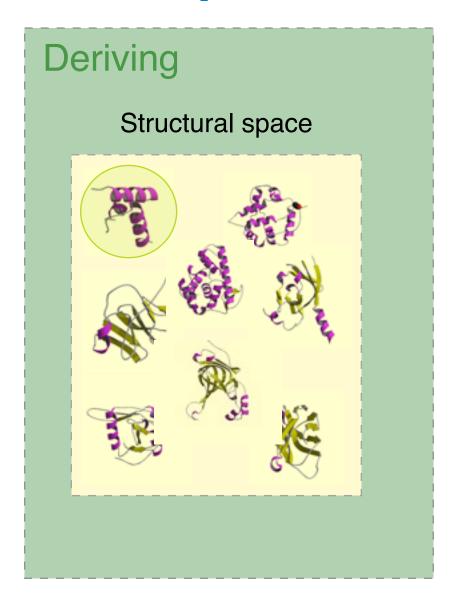
http://protein.bio.puc.cl/cardex/servers/anolea/

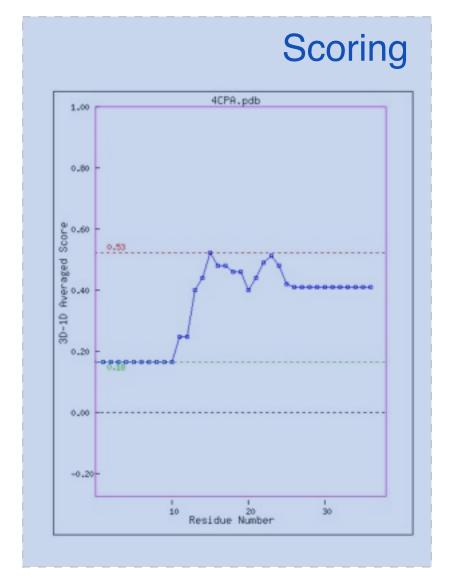




Verify3D

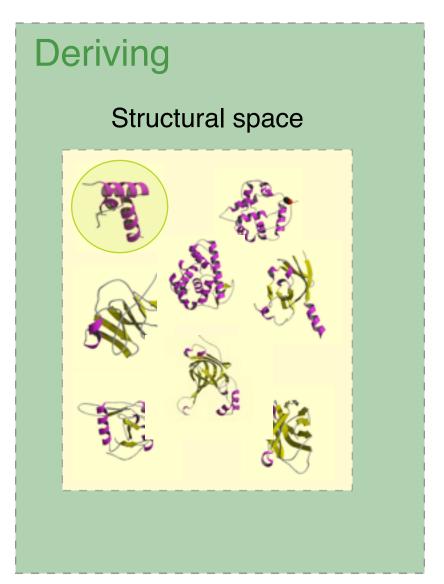
http://nihserver.mbi.ucla.edu/Verify_3D/





DFIRE

http://sparks.informatics.iupui.edu/

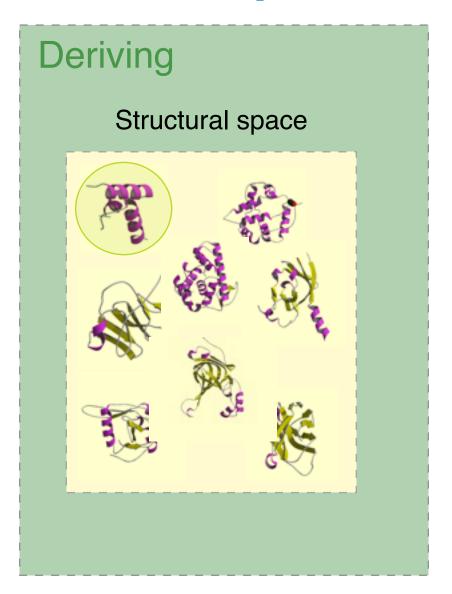


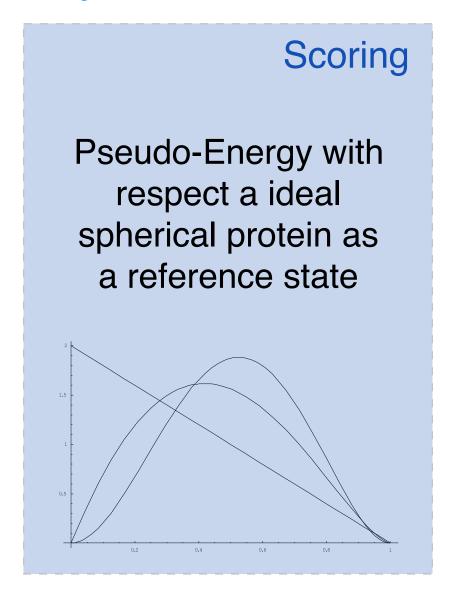
Scoring

Pseudo-Energy with respect a ideal gas-phase reference state

DOPE (MODELLER)

http://www.salilab.org/modeller/

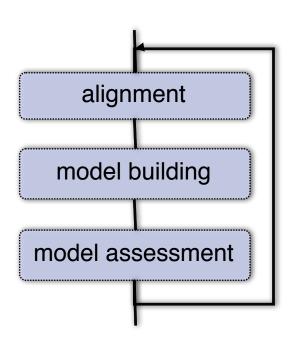


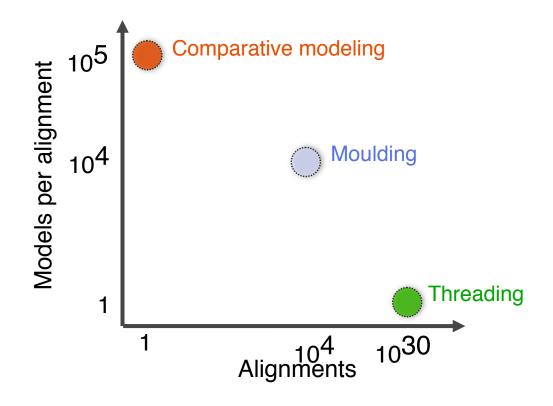




John, Sali (2003). NAR pp31 3982

Moulding: iterative alignment, model building, model assessment





Genetic algorithm operators

Single point cross-over ...TSSQ-NMK-LGVFWGY... ...TSSQ-NMKLGVFWGY-... ...V-SSCN-GDLHMKVGV... ...V-SSCNGDLHMKV-GV... ...TSSONMKLGVFWGY---... ...TSSQNMK——LGVFWGY... ...VSSCN—GDLHMKVGV... ...VSSCNGDLHMKV——GV... Gap insertion ...TSSQN—MKLGVFWGY... ...VSSCNGDLHMKVG—V... ...TSSONMKLGVFWGY... ...VSSCNGDLHMKVGV... ...-T-SSQNMKLGVFWGY... ...VSSCNGDLHMKVGV---... Gap shift ...T—S—SONMKLGVFWGY... ...VSSCNGDLHMKVGV—... ...T—SSONMKLGVFWGY... ...VSSCNGDLHMKVGV—... ...—TSSONMKLGVFWGY... ...VSSCNGDLHMKVGV—... ...TS-SQNMKLGVFWGY... ...VSSCNGDLHMKVGV-Also, "two point crossover" and "gap deletion".

Composite model assessment score

Weighted linear combination of several scores:

- Pair (P_D) and surface (P_S) statistical potentials;
- Structural compactness (S_C);
- Harmonic average distance score (H_a);
- Alignment score (A_S).

$$Z = 0.17 Z(P_P) + 0.02 Z(P_S) + 0.10 Z(S_C) + 0.26 Z(H_a) + 0.45 (A_S)$$

```
Z(score) = (score-\mu)/σ

\mu ... average score of all models

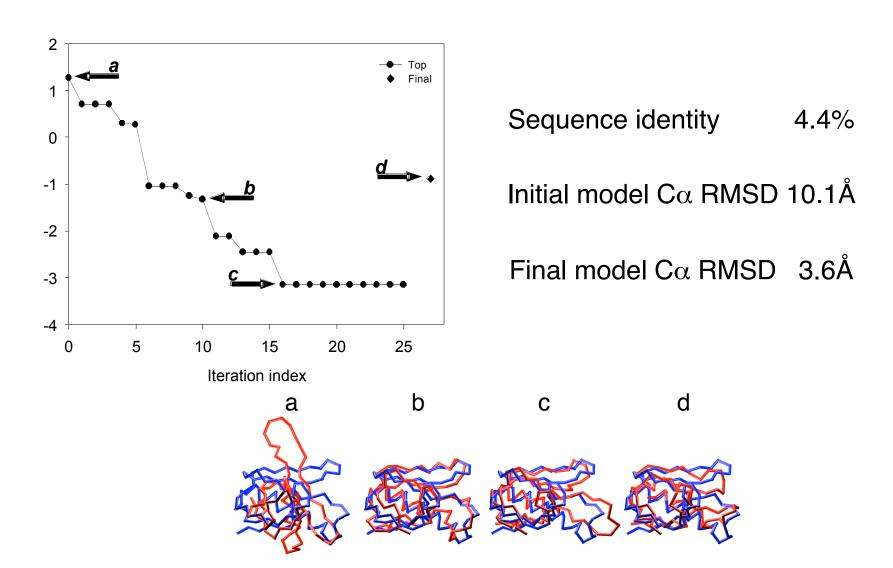
\sigma ... standard deviation of the scores
```

Benchmark with the "very difficult" test set

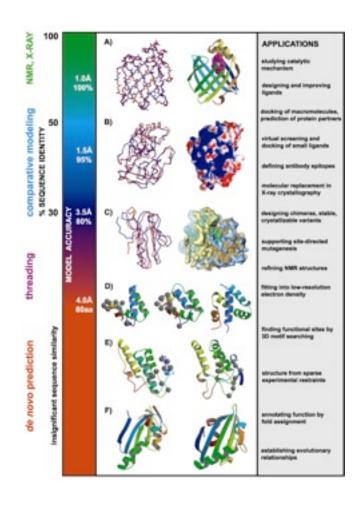
D. Fischer threading test set of 68 structural pairs (a subset of 19)

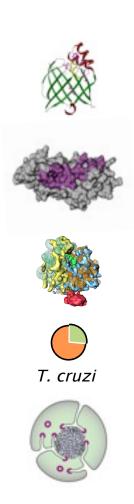
Target -template Sequence identity [%]			Initial prediction		Final prediction		Best prediction	
	Sequence identity [%]	uence entity Coverage [% aa]	Cα RMSD [Å]	CE overlap [%]	Cα RMSD [Å]	CE overlap [%]	Cα RMSD [Å]	CE overlap [%]
1ATR-1ATN	13.8	94.3	19.2	20.2	18.8	20.2	17.1	24.6
1BOV-1LTS	4.4	83.5	10.1	29.4	3.6	79.4	3.1	92.6
1CAU-1CAU	18.8	96.7	11.7	15.6	10.0	27.4	7.6	47.4
1COL-1CPC	11.2	81.4	8.6	44.0	5.6	58.6	4.8	59.3
1LFB-1HOM	17.6	75.0	1.2	100.0	1.2	100.0	1.1	100.0
1NSB-2SIM	10.1	89.2	13.2	20.2	13.2	20.1	12.3	26.8
1RNH-1HRH	26.6	91.2	13.0	21.2	4.8	35.4	3.5	57.5
1YCC-2MTA	14.5	55.1	3.4	72.4	5.3	58.4	3.1	75.0
2AYH-1SAC	8.8	78.4	5.8	33.8	5.5	48.0	4.8	64.9
2CCY-1BBH	21.3	97.0	4.1	52.4	3.1	73.0	2.6	77.0
2PLV-1BBT	20.2	91.4	7.3	58.9	7.3	58.9	6.2	60.7
2POR-2OMF	13.2	97.3	18.3	11.3	11.4	14.7	10.5	25.9
2RHE-1CID	21.2	61.6	9.2	33.7	7.5	51.1	4.4	71.1
2RHE-3HLA	2.4	96.0	8.1	16.5	7.6	9.4	6.7	43.5
3ADK-1GKY	19.5	100.0	13.8	26.6	11.5	37.7	7.7	48.1
3HHR-1TEN	18.4	98.9	7.3	60.9	6.0	66.7	4.9	79.3
4FGF-81IB	14.1	98.6	11.3	24.0	9.3	30.6	5.4	41.2
6XIA-3RUB	8.7	44.1	10.5	14.5	10.1	11.0	9.0	34.3
9RNT-2SAR	13.1	88.5	5.8	41.7	5.1	51.2	4.8	69.0
AVERAGE	14.2	85.2	9.6	36.7	7.7	44.8	6.3	57.8

Application to a difficult modeling case 1BOV-1LTS



Can we use models to infer function?







What is the physiological ligand of Brain Lipid-Binding Protein?

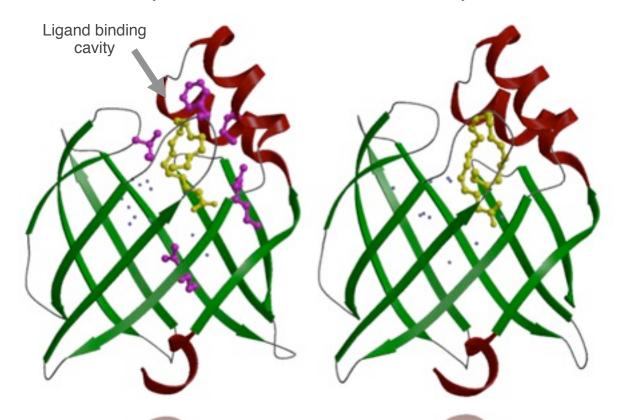
Predicting features of a model that are not present in the template

BLBP/oleic acid

BLBP/docosahexaenoic acid

Cavity is not filled

Cavity is filled



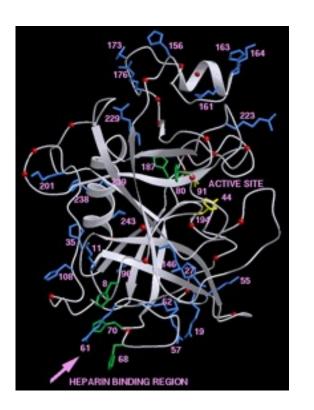
- 1. BLBP binds fatty acids.
 - 2. Build a 3D model.
- 3. Find the fatty acid that fits most snuggly into the ligand binding cavity.

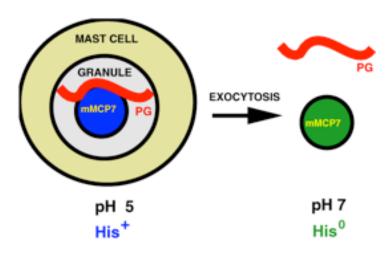
L. Xu, R. Sánchez, A. Šali, N. Heintz, J. Biol. Chem. 271, 24711, 1996.

Do mast cell proteases bind proteoglycans? Where? When?

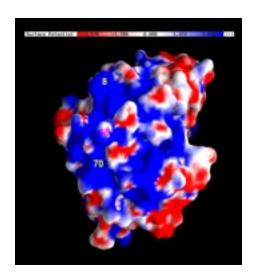
Predicting features of a model that are not present in the template

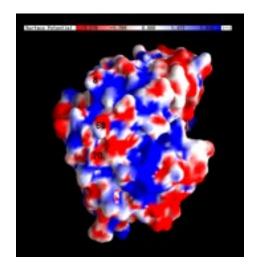
- 1. mMCPs bind negatively charged proteoglycans through electrostatic interactions
- 2. Comparative models used to find clusters of positively charged surface residues.
- 3. Tested by site-directed mutagenesis.



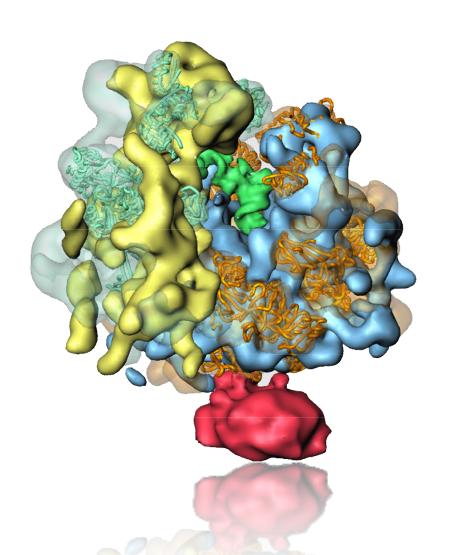


Huang et al. J. Clin. Immunol. **18**,169,1998. Matsumoto et al. J.Biol.Chem. **270**,19524,1995. Šali et al. J. Biol. Chem. **268**, 9023, 1993.





S. cerevisiae ribosome



Fitting of comparative models into 15Å cryo-electron density map.

43 proteins could be modeled on 20-56% seq.id. to a known structure.

The modeled fraction of the proteins ranges from 34-99%.

C. Spahn, R. Beckmann, N. Eswar, P. Penczek, A. Sali, G. Blobel, J. Frank. Cell 107, 361-372, 2001.

Common Evolutionary Origin of Coated Vesicles and Nuclear Pore Complexes

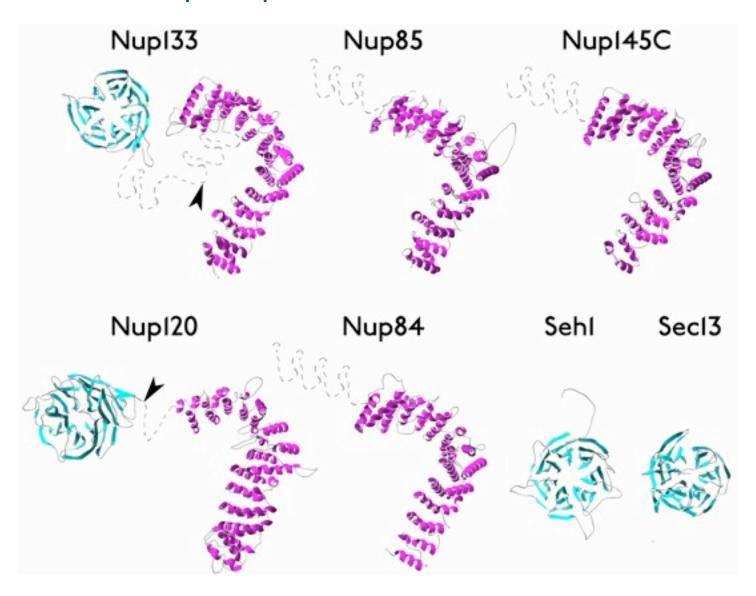
mGenThreader + SALIGN + MOULDER

D. Devos, S. Dokudovskaya, F. Alber, R. Williams, B.T. Chait, A. Sali, M.P. Rout. Components of Coated Vesicles and Nuclear Pore Complexes Share a Common Molecular Architecture. *PLOS Biology* **2(12)**:e380, 2004

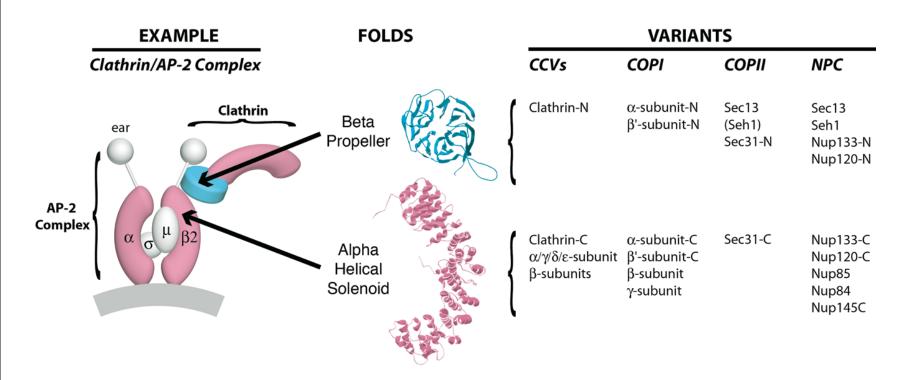
yNup84 complex proteins



All Nucleoporins in the Nup84 Complex are Predicted to Contain β -Propeller and/or α -Solenoid Folds

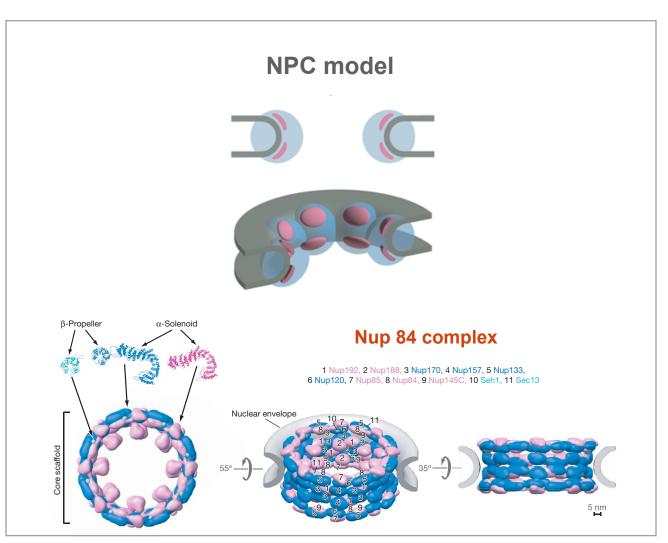


NPC and Coated Vesicles Share the β -Propeller and α -Solenoid Folds and Associate with Membranes



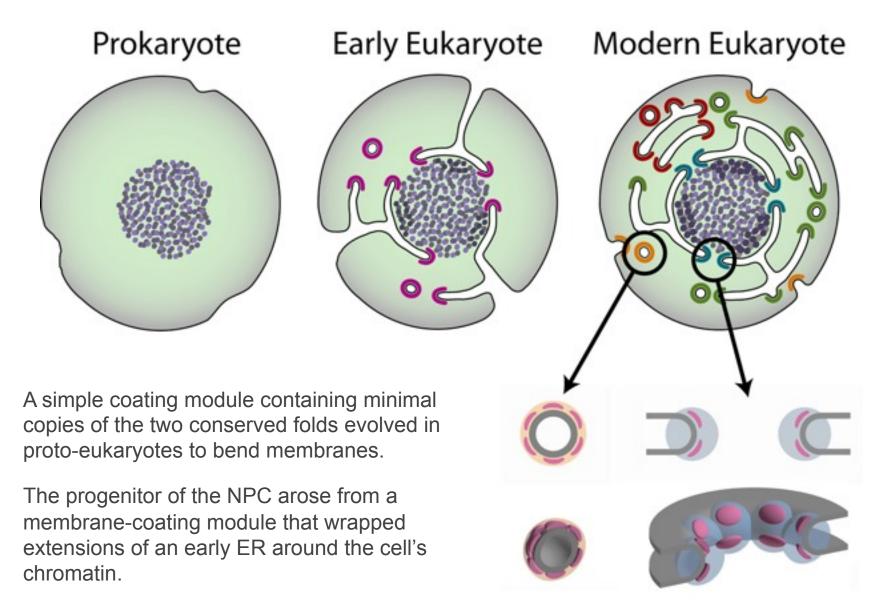
NPC and Coated Vesicles Both Associate with Membranes





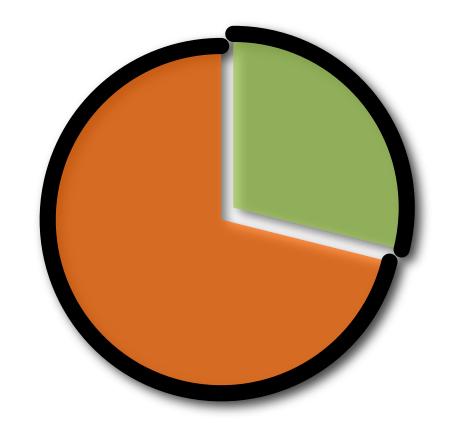
Alber et al. The molecular architecture of the nuclear pore complex. Nature (2007) vol. 450 (7170) pp. 695-701

A Common Evolutionary Origin for Nuclear Pore Complexes and Coated Vesicles? The proto-coatomer hypothesis



Tropical Disease Initiative (TDI)

Predicting binding sites in protein structure models.



http://www.tropicaldisease.org



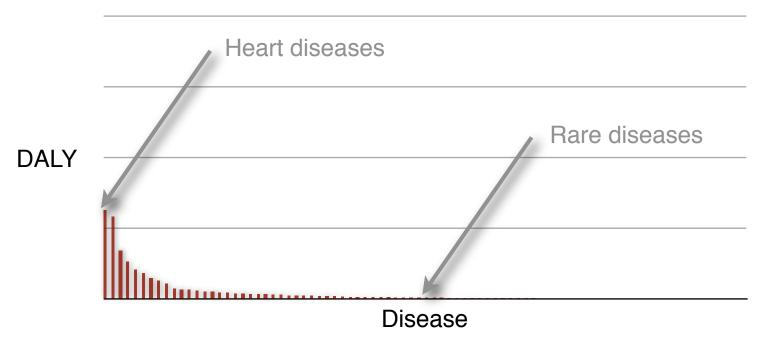






Need is High in the Tail

- DALY Burden Per Disease in Developed Countries
- DALY Burden Per Disease in Developing Countries



Disease data taken from WHO, World Health Report 2004

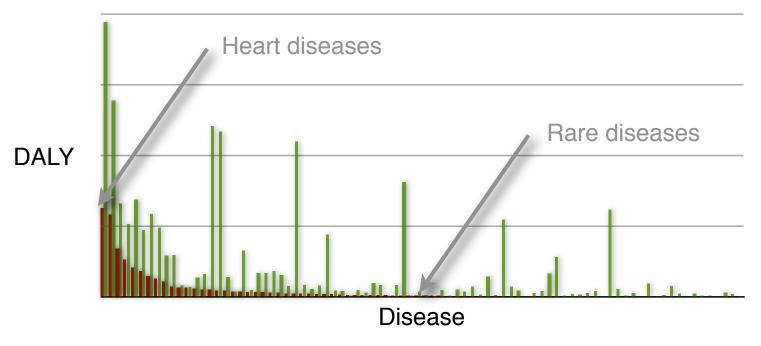
DALY - Disability adjusted life years

DALY is not a perfect measure of market size, but is certainly a good measure for importance.

DALYs for a disease are the sum of the years of life lost due to premature mortality (YLL) in the population and the years lost due to disability (YLD) for incident cases of the health condition. The DALY is a health gap measure that extends the concept of potential years of life lost due to premature death (PYLL) to include equivalent years of 'healthy' life lost in states of less than full health, broadly termed disability. One DALY represents the loss of one year of equivalent full health.

Need is High in the Tail

- DALY Burden Per Disease in Developed Countries
- DALY Burden Per Disease in Developing Countries



Disease data taken from WHO, World Health Report 2004

DALY - Disability adjusted life years

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"Unprofitable" Diseases and Global DALY (in 1000's)

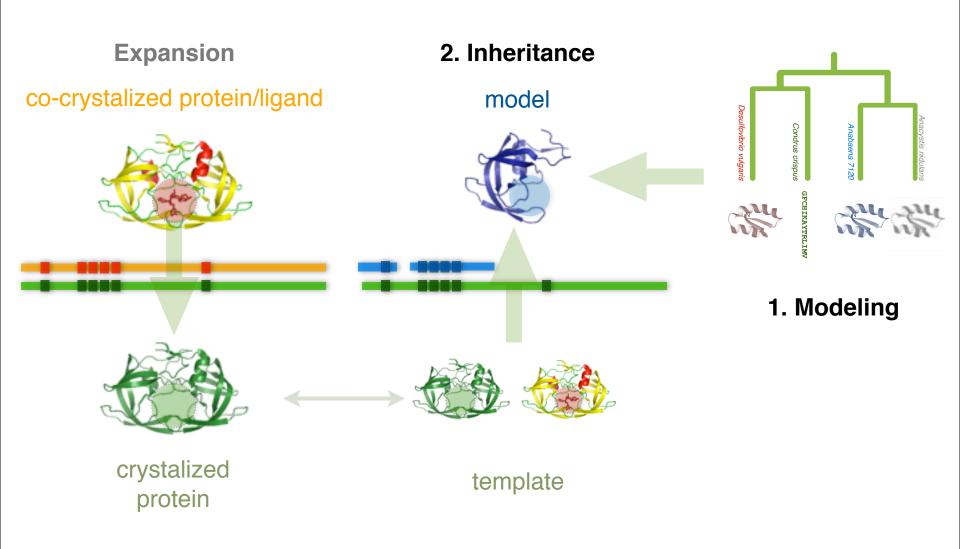
Malaria*	46,486
Tetanus	7,074
Lymphatic filariasis*	5,777
Syphilis	4,200
Trachoma	2,329
Leishmaniasis*	2,090
Ascariasis	1,817
Schistosomiasis*	1,702
Trypanosomiasis*	1,525

Trichuriasis	1,006
Japanese encephalitis	709
Chagas Disease*	667
Dengue*	616
Onchocerciasis*	484
Leprosy*	199
Diphtheria	185
Poliomyelitise	151
Hookworm disease	59

Disease data taken from WHO, <u>World Health Report 2004</u>
DALY - Disability adjusted life year in 1000's.

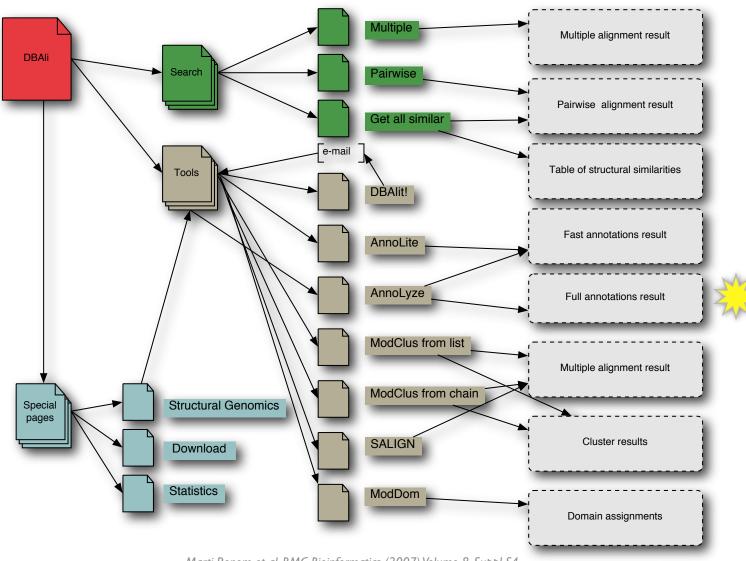
* Officially listed in the WHO Tropical Disease Research <u>disease portfolio</u>.

Comparative docking



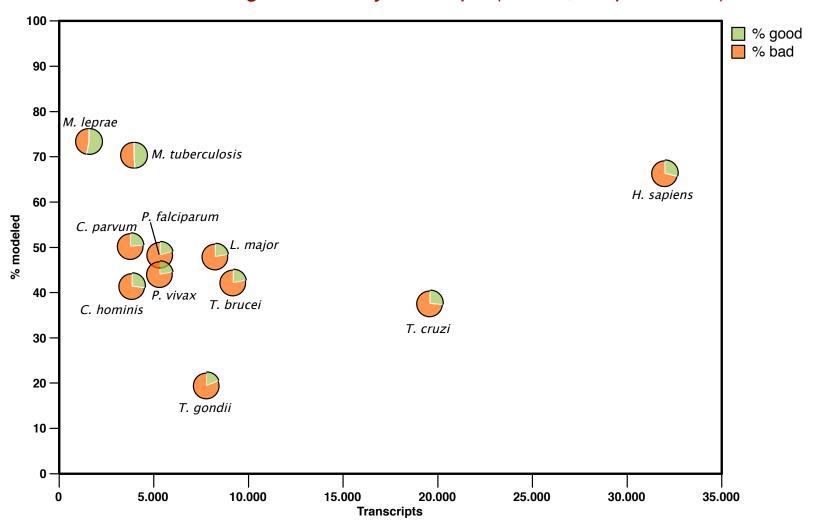
DBAliv2.0 database

http://www.dbali.org



Modeling Genomes

data from models generated by ModPipe (Eswar, Pieper & Sali)



Summary table

models with inherited ligands

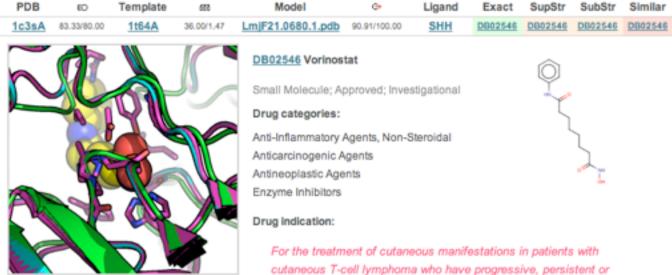
29,271 targets with good models, 297 inherited a ligand/substance similar to a known drug in DrugBank

	Transcripts	Modeled targets	Selected models	Inherited ligands	Similar to a drug	Drugs
C. hominis	3,886	1,614	666	197	20	13
C. parvum	3,806	1,918	742	232	24	13
L. major	8,274	3,975	1,409	478	43	20
М. Іергае	1,605	1,178	893	310	25	6
M. tuberculosis	3,991	2,808	1,608	365	30	10
P. falciparum	5,363	2,599	818	284	28	13
P. vivax	5,342	2,359	822	268	24	13
T. brucei	7,793	1,530	300	138	13	6
T. cruzi	19,607	7,390	3,070	769	51	28
T. gondii	9,210	3,900	1,386	458	39	21
TOTAL	68,877	29,271	11,714	3,499	297	143

L. major Histone deacetylase 2 + Vorinostat

Template 1t64A a human HDAC8 protein.





recurrent disease on or following two systemic therapies.

L. major Histone deacetylase 2 + Vorinostat

Literature

Proc. Natl. Acad. Sci. USA Vol. 93, pp. 13143–13147, November 1996 Medical Sciences

Apicidin: A novel antiprotozoal agent that inhibits parasite histone deacetylase

(cyclic tetrapeptide/Apicomplexa/antiparasitic/malaria/coccidiosis)

Sandra J. Darkin-Rattray*†, Anne M. Gurnett*, Robert W. Myers*, Paula M. Dulski*, Tami M. Crumley*, John J. Allocco*, Christine Cannova*, Peter T. Meinke‡, Steven L. Colletti‡, Maria A. Bednarek‡, Sheo B. Singh§, Michael A. Goetz§, Anne W. Dombrowski§, Jon D. Polishook§, and Dennis M. Schmatz*

Departments of *Parasite Biochemistry and Cell Biology, [‡]Medicinal Chemistry, and [§]Natural Products Drug Discovery, Merck Research Laboratories, P.O. Box 2000, Rahway, NJ 07065

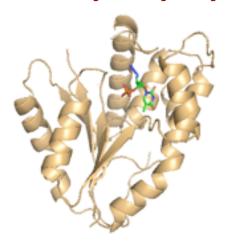
ANTIMICROBIAL AGENTS AND CHEMOTHERAPY, Apr. 2004, p. 1435–1436 0066-4804/04/\$08.00+0 DOI: 10.1128/AAC.48.4.1435–1436.2004 Copyright © 2004, American Society for Microbiology. All Rights Reserved.

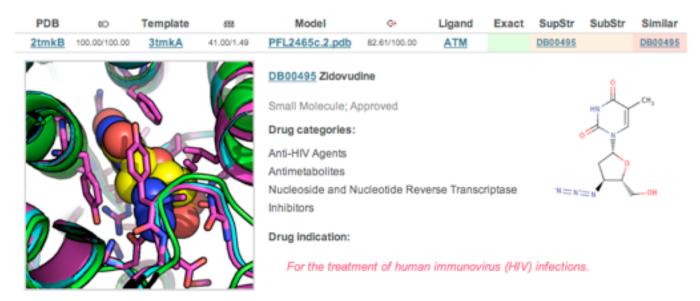
Vol. 48, No. 4

Antimalarial and Antileishmanial Activities of Aroyl-Pyrrolyl-Hydroxyamides, a New Class of Histone Deacetylase Inhibitors

P. falciparum tymidylate kinase + zidovudine

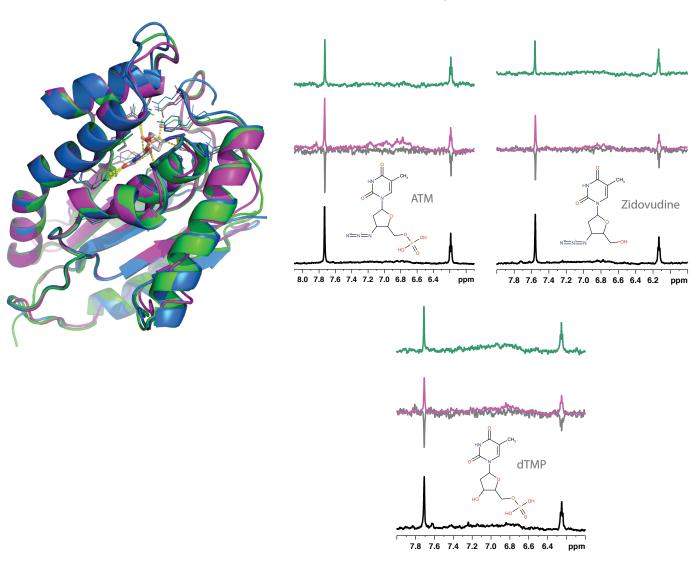
Template 3tmkA a yeast tymidylate kinase.





P. falciparum tymydilate kinase + zidovudine

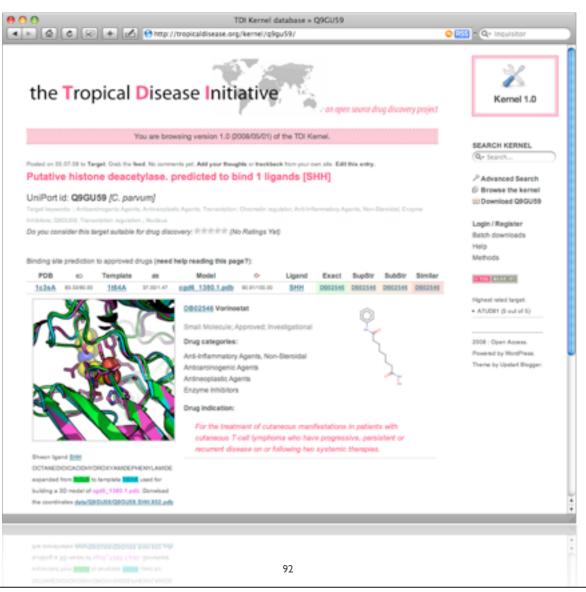
NMR Water-LOGSY and STD experiments



Leticia Ortí, Rodrigo J. Carbajo, and Antonio Pineda-Lucena

TDI's kernel

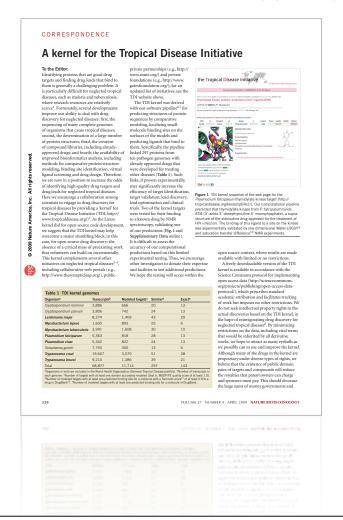
http://tropicaldisease.org/kernel



TDI's kernel

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http://integrativemodeling.org

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