### **Comparative Protein Structure Prediction**



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# Objective

# TO LEARN HOW-TO MODEL A 3D-STRUCTURE FROM A SEQUENCE AND A KNOWN STRUCTURE

### **DISCLAIMER!**

Name	Type	World Wide Web address
DATABASES	.,,,	
CATH	s	http://www.biochem.ucl.ac.uk/bsm/cath/
DBAII	s	http://www.salilab.org/DBAli/
GenBank	s	http://www.ncbi.nlm.nih.gov/Genbank/GenbankSearch.html
GeneCensus	s	http://bioinfo.mbb.yale.edu/genome
MODBASE	s	http://salilab.org/modbase/
MSD	s	http://www.ebi.ac.uk/msd/
NCBI	s	http://www.ncbi.nlm.nih.gov/
PDB	s	http://www.rcsb.org/pdb/
PSI	s	http://www.nigms.nih.gov/psi/
Sacch3D	s	http://genome-www.stanford.edu/Sacch3D/
SCOP	s	http://scop.mrc-lmb.cam.ac.uk/scop/
TIGR	s	http://www.tigr.org/tdb/mdb/mdbcomplete.html
TrEMBL	s	http://srs.ebi.ac.uk/
FOLD ASSIGNM	ENT	
123D	s	

http://sgu.bioinfo.cipf.es/home/?page=resources

### Programs, servers and databases

http://salilab.org

#### LS-SNP **PIBASE MODLOOP CCPR** Web Server **Database Web Server Center for Computational** http://salilab.org/LS-SNP http://salilab.org/pibase http://salilab.org/modloop **Proteomics Research** Predicts functional impact Contains structurally defined Models loops in protein http://www.ccpr.ucsf.edu of residue substitution protein interfaces structures **MODBASE DBALI MODWEB MODELLER Database Database Web Server Program** http://salilab.org/modbase http://salilab.org/dbali Fold assignments, alignments http://salilab.org/modweb http://salilab.org/modeller Contains a comprehensive models, model assessments Provides a web interface to Implements most operations set of pairwise and multiple for all sequences related to a **MODPIPE** in comparative modeling structure-based alignments known structure **EVA ICEDB MODPIPE LIGBASE Web Server** Database/LIMS **Database Program** http://salilab.org/eva http://nysgxrc.org Automatically calculates Ligand binding sites and Evaluates and ranks web Tracks targets for structural comparative models of many inheritance (accessible servers for protein structure through MODBASE) protein sequences genomics by NYSGXRC prediction

#### **External Resources**

PDB, Uniprot, GENBANK, NR, PIR, INTERPRO, Kinase Resource UCSC Genome Browser, CHIMERA, Pfam, SCOP, CATH

### Nomenclature

**Homology**: Sharing a common ancestor, may have similar or dissimilar functions

**Similarity**: Score that quantifies the degree of relationship between two sequences.

**Identity**: Fraction of identical aminoacids between two aligned sequences (case of similarity).

**Target**: Sequence corresponding to the protein to be modeled.

**Template**: 3D structure/s to be used during protein structure prediction.

**Model**: Predicted 3D structure of the target sequence.

### Nomenclature

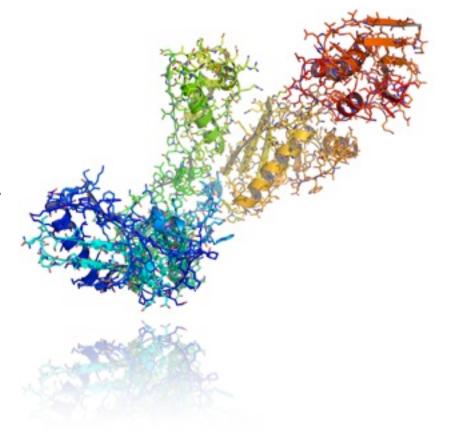
**Fold**: Three dimensional conformation of a protein sequence (usually at domain level).

**Domain**: Structurally globular part of a protein, which may independently fold.

**Secondary Structure**: Regular subdomain structures composed by alphahelices, beta-sheets and coils (or loops).

**Backbone**: Protein structure skeleton composed by the carbon, nitrogen and oxygen atoms.

**Side-Chain**: Specific atoms identifying each of the 20 residues types.



### protein prediction .vs. protein determination

X-Ray

**NMR** 

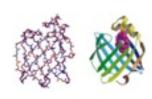
Experimental

data

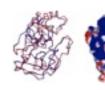
**Comparative Modeling** 

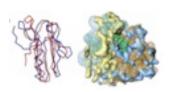
**Threading** 

**Ab-initio** 



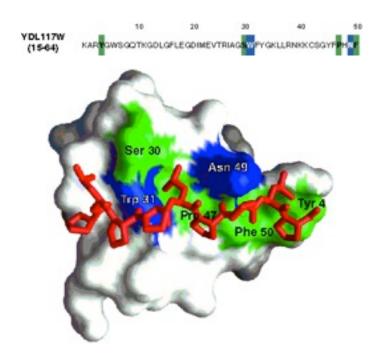
inferred data\_





# Why is it useful to know the structure of a protein, not only its sequence?

- The biochemical function (activity) of a protein is defined by its interactions with other molecules.
- The biological function is in large part a consequence of these interactions.
- The 3D structure is more informative than sequence because interactions are determined by residues that are close in space but are frequently distant in sequence.

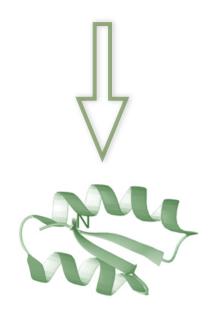


In addition, since evolution tends to conserve function and function depends more directly on structure than on sequence, **structure is more conserved in evolution than sequence**.

The net result is that patterns in space are frequently more recognizable than patterns in sequence.

## Principles of protein structure

GFCHIKAYTRLIMVG...



Anacystis nidulans

Anabaena 7120

Anabaena 7120

GFCHIKAYTRLIMVG...

Condrus crispus

Desulfovibrio vulgaris

Folding (physics)

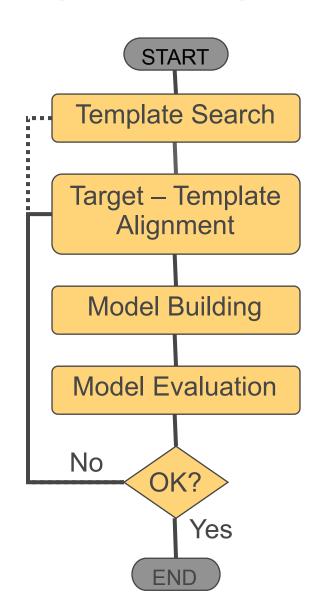
Ab initio prediction

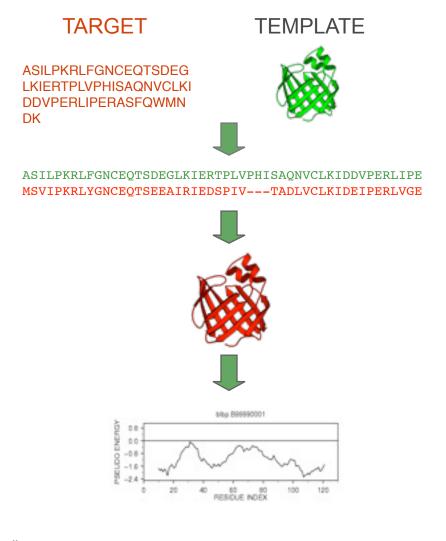
Evolution (rules)

Threading Comparative Modeling

D. Baker & A. Sali. Science 294, 93, 2001.

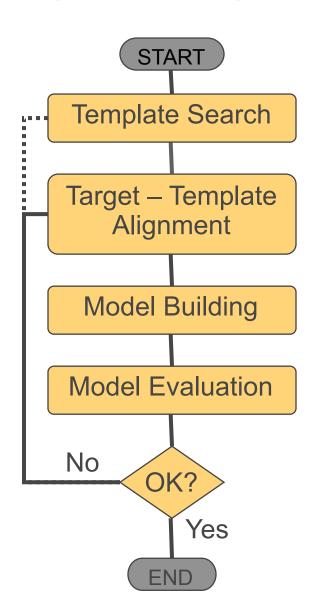
### **Steps in Comparative Protein Structure Modeling**

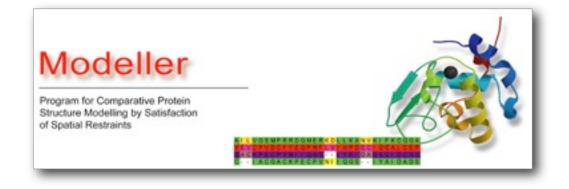




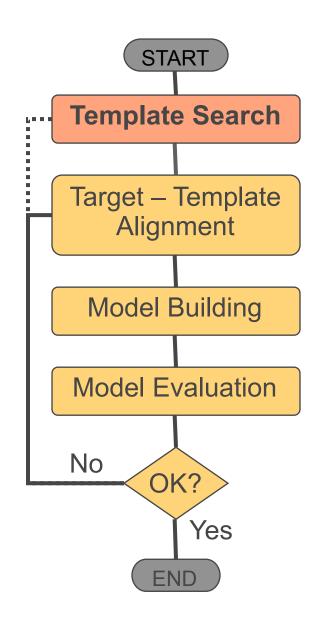
A. Šali, Curr. Opin. Biotech. 6, 437, 1995.
R. Sánchez & A. Šali, Curr. Opin. Str. Biol. 7, 206, 1997.
M. Marti et al. Ann. Rev. Biophys. Biomolec. Struct., 29, 291, 2000.

### **Steps in Comparative Protein Structure Modeling**





### **Template Search**



Sequence-Sequence search

**BLAST** 

http://www.ncbi.nlm.nih.gov/blast/

Profile-Sequence search

**PSI-BLAST** 

http://www.ncbi.nlm.nih.gov/blast/

Profile-Profile search

pp\_scan

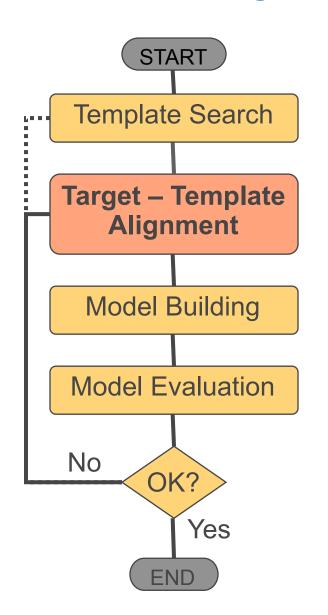
mod9v8

Sequence-Structure search

GenThreader

http://bioinf.cs.ucl.ac.uk/psipred/

### **Target-Template Alignment**



Sequence-Sequence search

**BLAST** 

http://www.ncbi.nlm.nih.gov/blast/

Profile-Sequence search

**PSI-BLAST** 

http://www.ncbi.nlm.nih.gov/blast/

**Profile-Profile search** 

pp\_scan

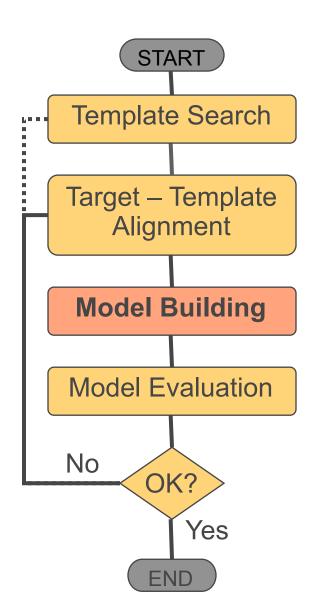
mod9v8

Sequence-Structure search

GenThreader

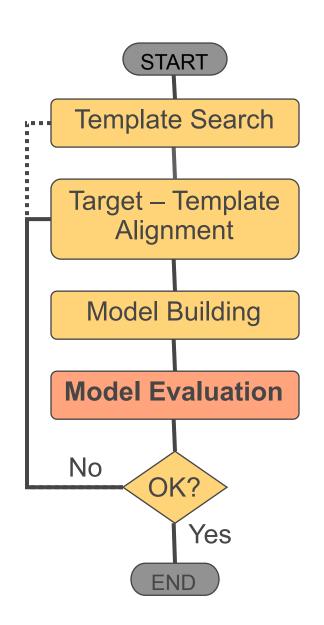
http://bioinf.cs.ucl.ac.uk/psipred/

### **Model Building**





### **Model Evaluation**



Classical potential of mean force

**PROSA-WEB** 

https://prosa.services.came.sbg.ac.at

Protein conformation free energies

**DFIRE** 

http://sparks.informatics.iupui.edu/vuevang/DFIRE/dDFIRE-service

Discrete optimized protein energy

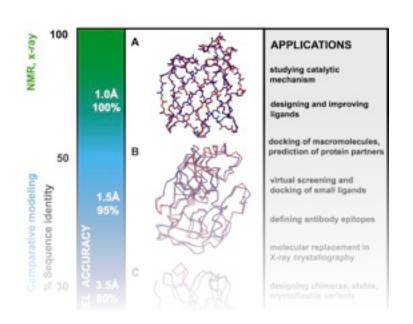
assess\_dope

mod9v8

Stereochemistry (<u>not necessary</u> but useful)

**PROCHECK** 

http://www.ebi.ac.uk/thornton-srv/software/PROCHECK/



# Accuracy and applicability of comparative models

# Comparative modeling by satisfaction of spatial restraints Types of errors and their impact



Wrong fold



Miss alignments



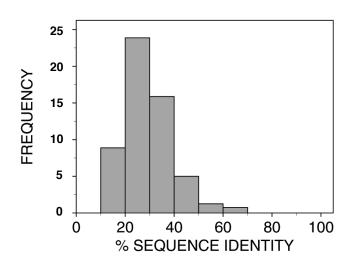
**Loop regions** 

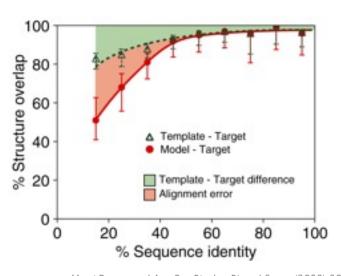


**Rigid body distortions** 



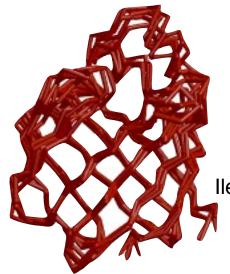
Side-chain packing





Marti-Renom etal. Ann Rev Biophys Biomol Struct (2000) 29, 291

### "Biological" significance of modeling errors



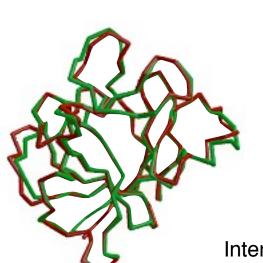
#### NMR - X-RAY

Erabutoxin 3ebx Erabutoxin 1era

#### **NMR**

Ileal lipid-binding protein

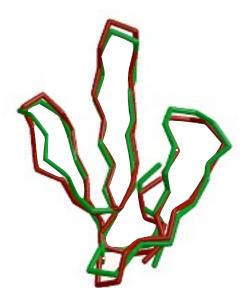
1eal

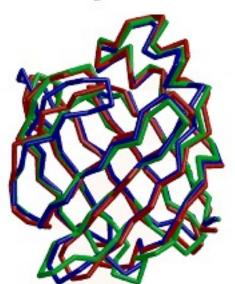


CRABPII 1opbB
FABP 1ftpA
ALBP 1lib
40% seq. id.



Interleukin 1β 41bi (2.9Å) Interleukin 1β 2mib (2.8Å)



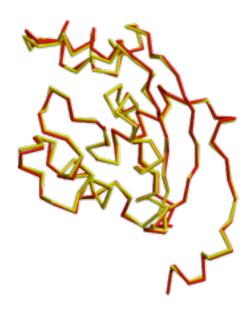


### **Model Accuracy**

#### HIGH ACCURACY

NM23 Seq id 77%

Cα equiv 147/148 RMSD 0.41Å

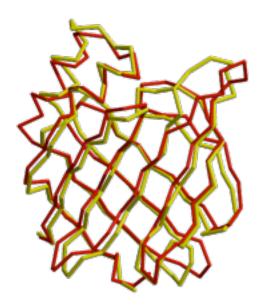


Sidechains Core backbone Loops

#### **MEDIUM ACCURACY**

CRABP Seq id 41%

Cα equiv 122/137 RMSD 1.34Å



Sidechains Core backbone Loops Alignment

#### **LOW ACCURACY**

EDN Seq id 33%

 $C\alpha$  equiv 90/134 RMSD 1.17Å

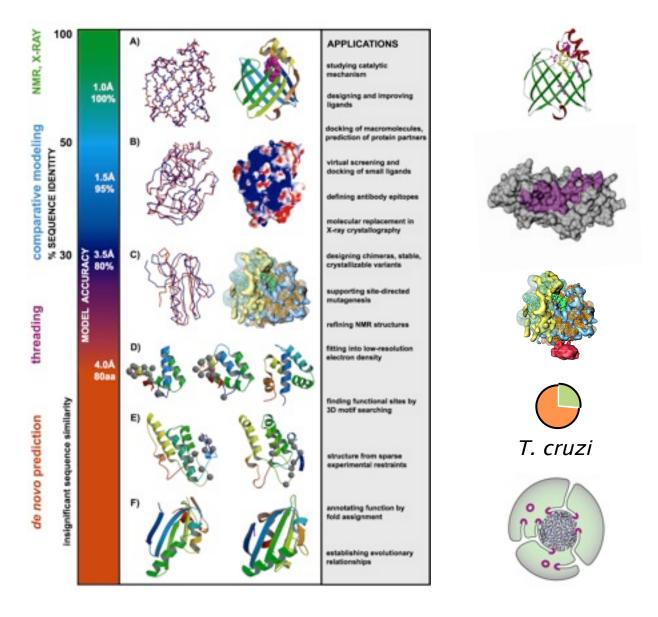


Sidechains
Core backbone
Loops
Alignment
Fold assignment

X-RAY / MODEL

Marti-Renom et al. Annu.Rev.Biophys.Biomol.Struct. 29, 291-325, 2000.

### Utility of protein structure models, despite errors



D. Baker & A. Sali. Science 294, 93, 2001.



# What is the physiological ligand of Brain Lipid-Binding Protein?

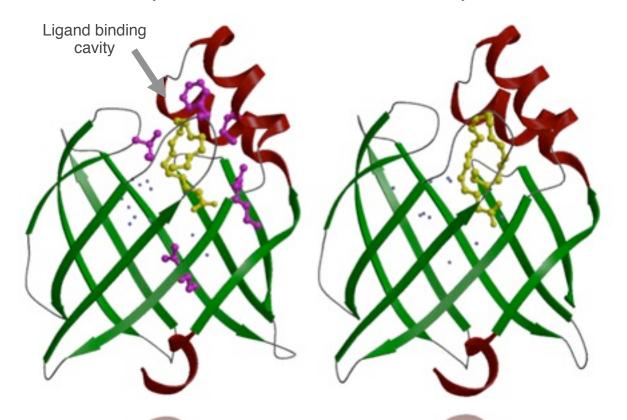
Predicting features of a model that are not present in the template

BLBP/oleic acid

BLBP/docosahexaenoic acid

Cavity is not filled

Cavity is filled



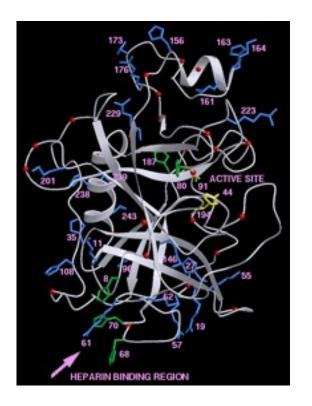
- 1. BLBP binds fatty acids.
  - 2. Build a 3D model.
- 3. Find the fatty acid that fits most snuggly into the ligand binding cavity.

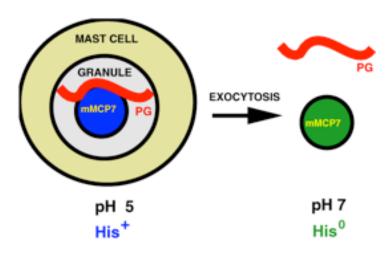
L. Xu, R. Sánchez, A. Šali, N. Heintz, J. Biol. Chem. 271, 24711, 1996.

#### Do mast cell proteases bind proteoglycans? Where? When?

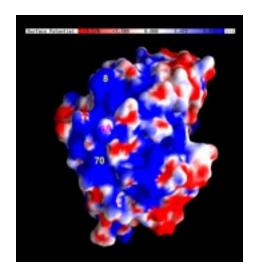
#### Predicting features of a model that are not present in the template

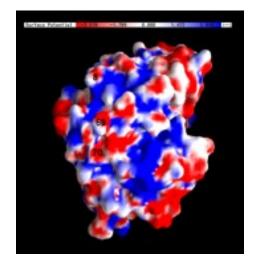
- 1. mMCPs bind negatively charged proteoglycans through electrostatic interactions
- 2. Comparative models used to find clusters of positively charged surface residues.
- 3. Tested by site-directed mutagenesis.



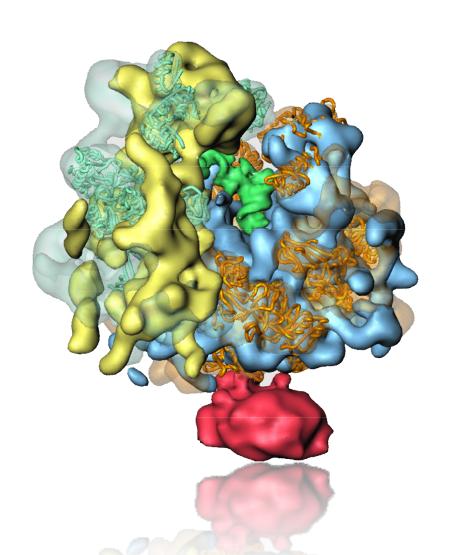


Huang et al. J. Clin. Immunol. **18**,169,1998. Matsumoto et al. J.Biol.Chem. **270**,19524,1995. Šali et al. J. Biol. Chem. **268**, 9023, 1993.





### S. cerevisiae ribosome



Fitting of comparative models into 15Å cryo-electron density map.

43 proteins could be modeled on 20-56% seq.id. to a known structure.

The modeled fraction of the proteins ranges from 34-99%.

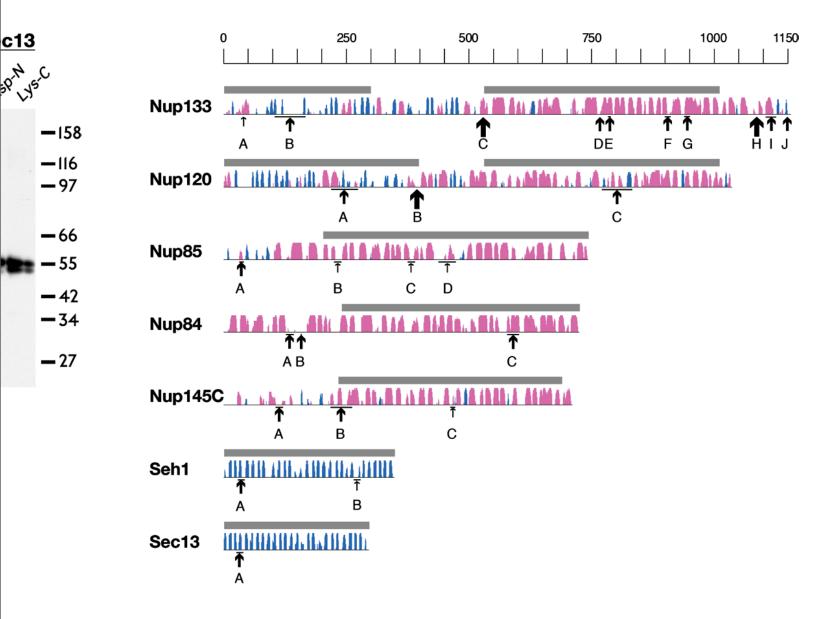
C. Spahn, R. Beckmann, N. Eswar, P. Penczek, A. Sali, G. Blobel, J. Frank. Cell 107, 361-372, 2001.

# **Common Evolutionary Origin of Coated Vesicles and Nuclear Pore Complexes**

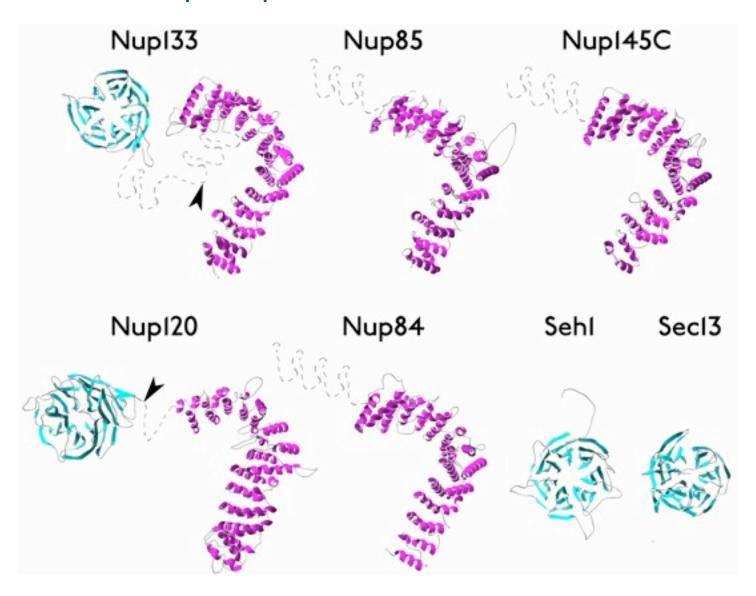
mGenThreader + SALIGN + MOULDER

D. Devos, S. Dokudovskaya, F. Alber, R. Williams, B.T. Chait, A. Sali, M.P. Rout. Components of Coated Vesicles and Nuclear Pore Complexes Share a Common Molecular Architecture. *PLOS Biology* **2(12)**:e380, 2004

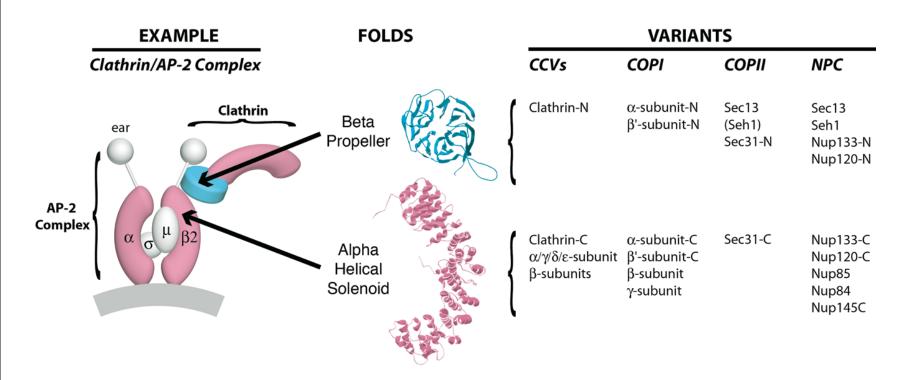
## yNup84 complex proteins



# All Nucleoporins in the Nup84 Complex are Predicted to Contain $\beta$ -Propeller and/or $\alpha$ -Solenoid Folds

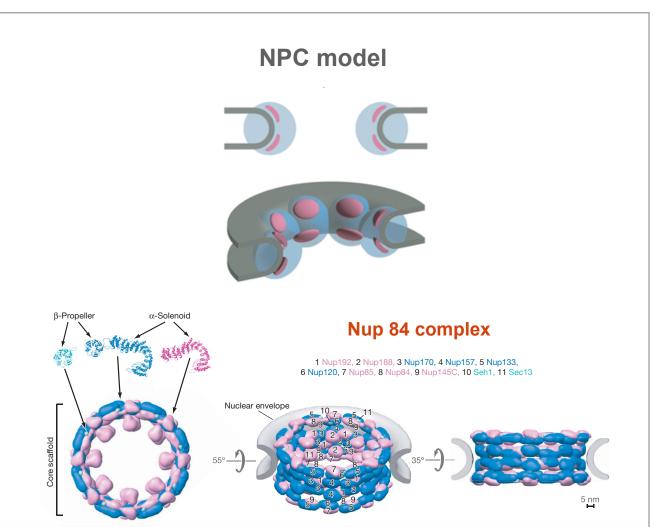


# NPC and Coated Vesicles Share the $\beta$ -Propeller and $\alpha$ -Solenoid Folds and Associate with Membranes



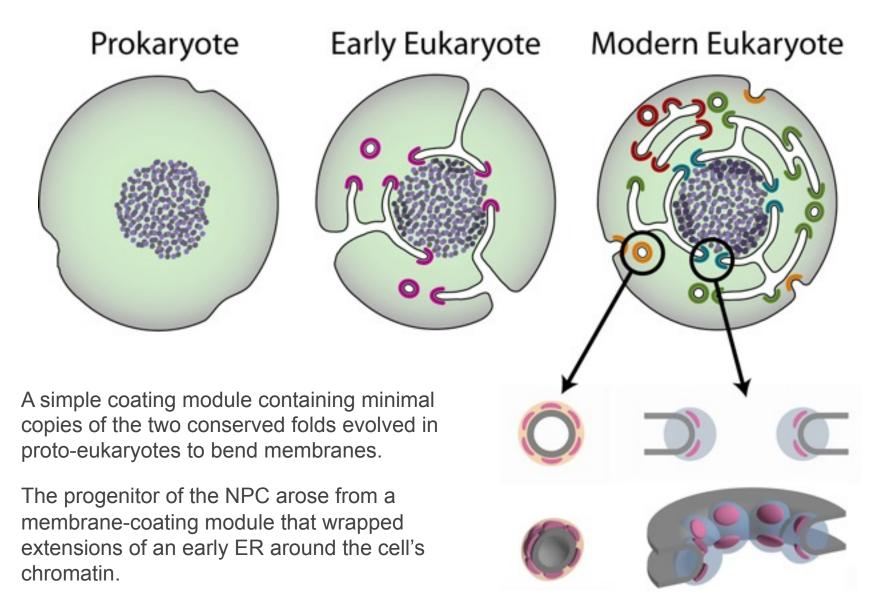
# NPC and Coated Vesicles Both Associate with Membranes





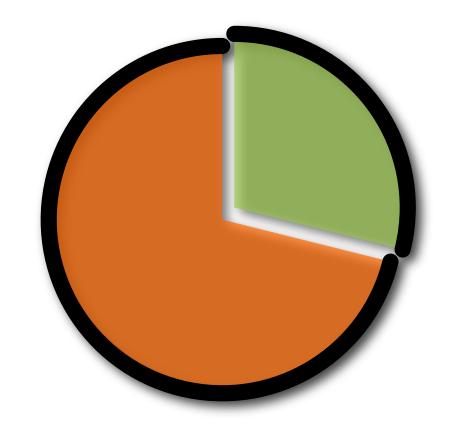
Alber et al. The molecular architecture of the nuclear pore complex. Nature (2007) vol. 450 (7170) pp. 695-701

# A Common Evolutionary Origin for Nuclear Pore Complexes and Coated Vesicles? The proto-coatomer hypothesis



### **Tropical Disease Initiative (TDI)**

Predicting binding sites in protein structure models.



http://www.tropicaldisease.org



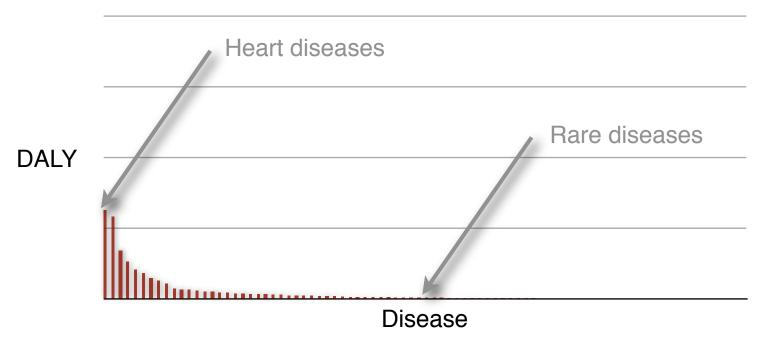






# Need is High in the Tail

- DALY Burden Per Disease in Developed Countries
- DALY Burden Per Disease in Developing Countries



Disease data taken from WHO, World Health Report 2004

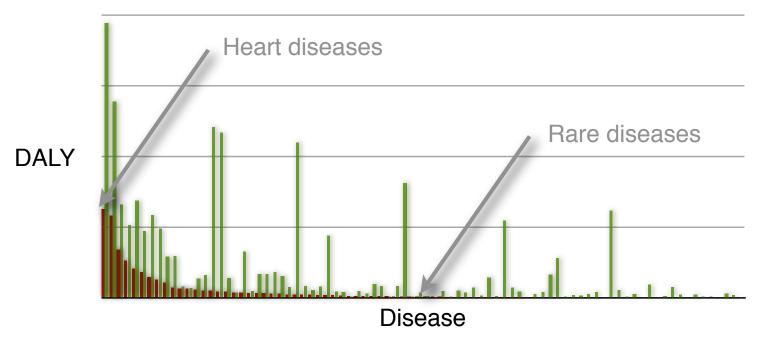
DALY - Disability adjusted life years

DALY is not a perfect measure of market size, but is certainly a good measure for importance.

DALYs for a disease are the sum of the years of life lost due to premature mortality (YLL) in the population and the years lost due to disability (YLD) for incident cases of the health condition. The DALY is a health gap measure that extends the concept of potential years of life lost due to premature death (PYLL) to include equivalent years of 'healthy' life lost in states of less than full health, broadly termed disability. One DALY represents the loss of one year of equivalent full health.

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# "Unprofitable" Diseases and Global DALY (in 1000's)

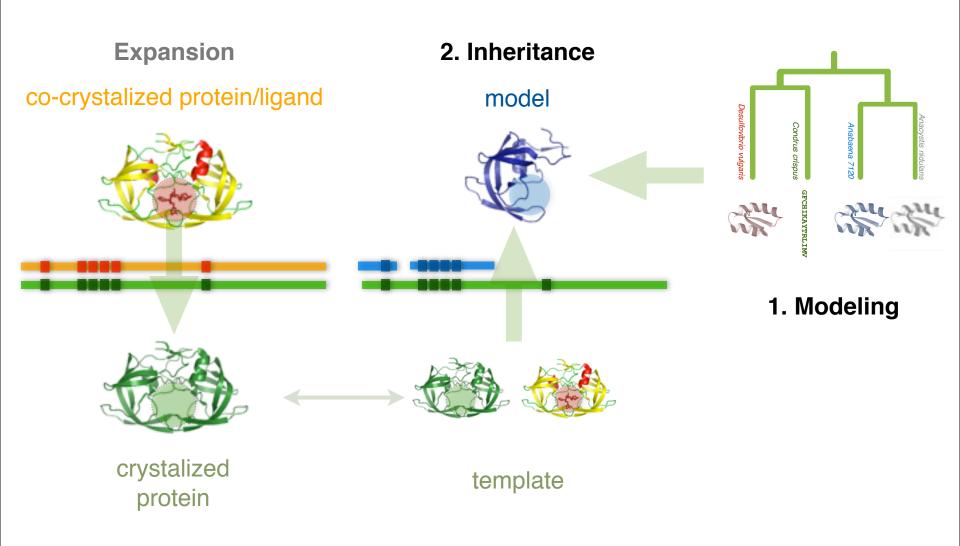
Malaria*	46,486
Tetanus	7,074
Lymphatic filariasis*	5,777
Syphilis	4,200
Trachoma	2,329
Leishmaniasis*	2,090
Ascariasis	1,817
Schistosomiasis*	1,702
Trypanosomiasis*	1,525

Trichuriasis	1,006
Japanese encephalitis	709
Chagas Disease*	667
Dengue*	616
Onchocerciasis*	484
Leprosy*	199
Diphtheria	185
Poliomyelitise	151
Hookworm disease	59

Disease data taken from WHO, <u>World Health Report 2004</u>
DALY - Disability adjusted life year in 1000's.

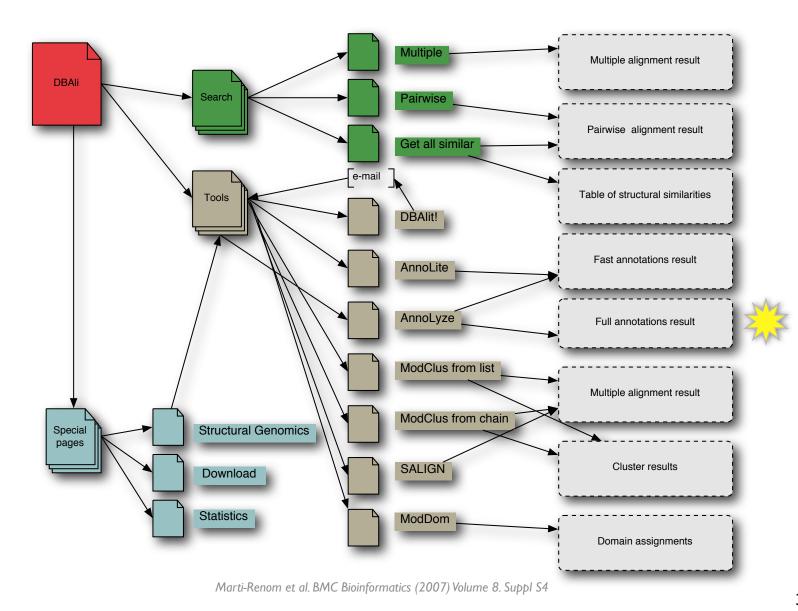
\* Officially listed in the WHO Tropical Disease Research <u>disease portfolio</u>.

# Comparative docking



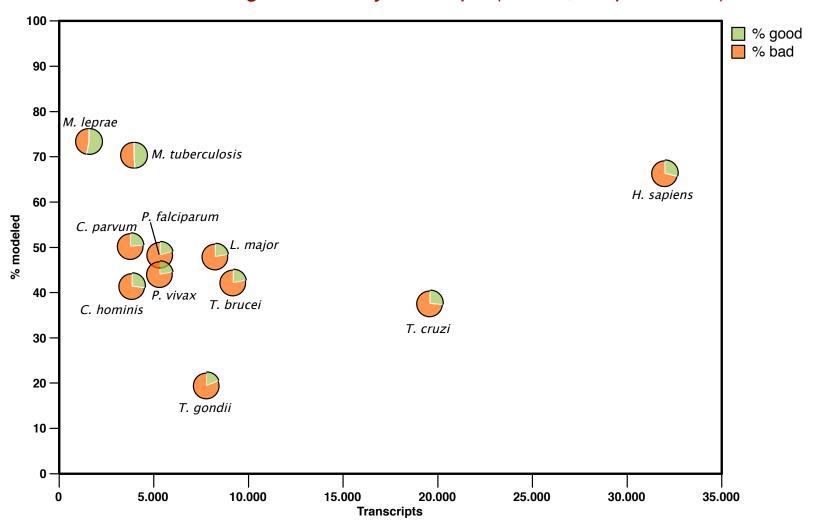
### DBAliv2.0 database

http://www.dbali.org



# **Modeling Genomes**

data from models generated by ModPipe (Eswar, Pieper & Sali)



# Summary table

#### models with inherited ligands

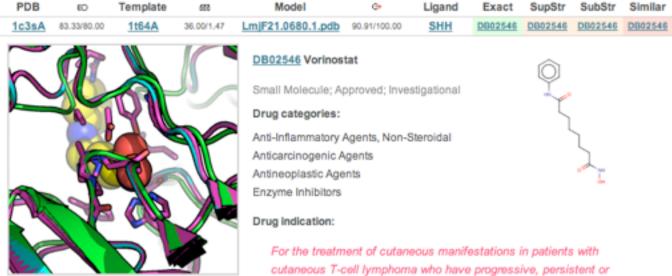
# 29,271 targets with good models, 297 inherited a ligand/substance similar to a known drug in DrugBank

	Transcripts	Modeled targets	Selected models	Inherited ligands	Similar to a drug	Drugs
C. hominis	3,886	1,614	666	197	20	13
C. parvum	3,806	1,918	742	232	24	13
L. major	8,274	3,975	1,409	478	43	20
М. Іергае	1,605	1,178	893	310	25	6
M. tuberculosis	3,991	2,808	1,608	365	30	10
P. falciparum	5,363	2,599	818	284	28	13
P. vivax	5,342	2,359	822	268	24	13
T. brucei	7,793	1,530	300	138	13	6
T. cruzi	19,607	7,390	3,070	769	51	28
T. gondii	9,210	3,900	1,386	458	39	21
TOTAL	68,877	29,271	11,714	3,499	297	143

## L. major Histone deacetylase 2 + Vorinostat

Template 1t64A a human HDAC8 protein.





recurrent disease on or following two systemic therapies.

## L. major Histone deacetylase 2 + Vorinostat

#### Literature

Proc. Natl. Acad. Sci. USA Vol. 93, pp. 13143–13147, November 1996 Medical Sciences

## Apicidin: A novel antiprotozoal agent that inhibits parasite histone deacetylase

(cyclic tetrapeptide/Apicomplexa/antiparasitic/malaria/coccidiosis)

Sandra J. Darkin-Rattray\*†, Anne M. Gurnett\*, Robert W. Myers\*, Paula M. Dulski\*, Tami M. Crumley\*, John J. Allocco\*, Christine Cannova\*, Peter T. Meinke‡, Steven L. Colletti‡, Maria A. Bednarek‡, Sheo B. Singh§, Michael A. Goetz§, Anne W. Dombrowski§, Jon D. Polishook§, and Dennis M. Schmatz\*

Departments of \*Parasite Biochemistry and Cell Biology, <sup>‡</sup>Medicinal Chemistry, and <sup>§</sup>Natural Products Drug Discovery, Merck Research Laboratories, P.O. Box 2000, Rahway, NJ 07065

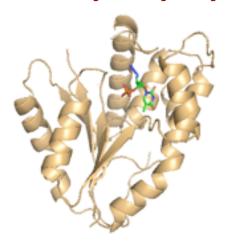
ANTIMICROBIAL AGENTS AND CHEMOTHERAPY, Apr. 2004, p. 1435–1436 0066-4804/04/\$08.00+0 DOI: 10.1128/AAC.48.4.1435–1436.2004 Copyright © 2004, American Society for Microbiology. All Rights Reserved.

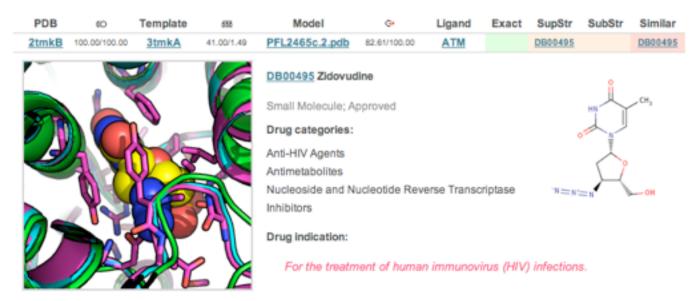
Vol. 48, No. 4

Antimalarial and Antileishmanial Activities of Aroyl-Pyrrolyl-Hydroxyamides, a New Class of Histone Deacetylase Inhibitors

### P. falciparum tymidylate kinase + zidovudine

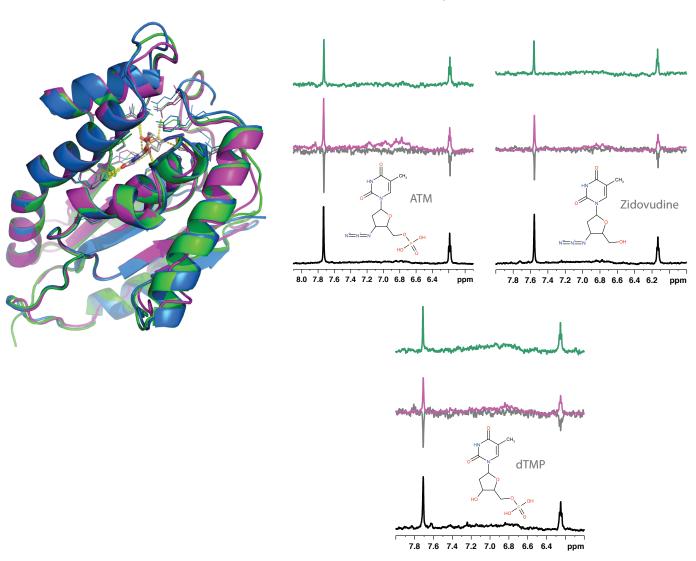
Template 3tmkA a yeast tymidylate kinase.





### P. falciparum tymydilate kinase + zidovudine

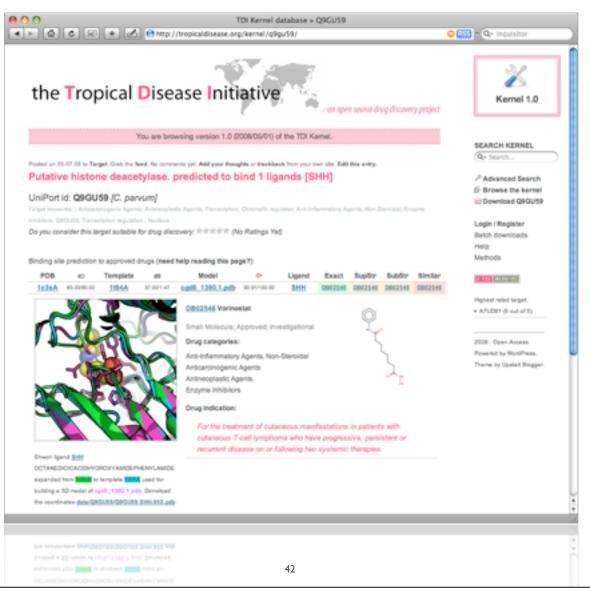
NMR Water-LOGSY and STD experiments



Leticia Ortí, Rodrigo J. Carbajo, and Antonio Pineda-Lucena

## TDI's kernel

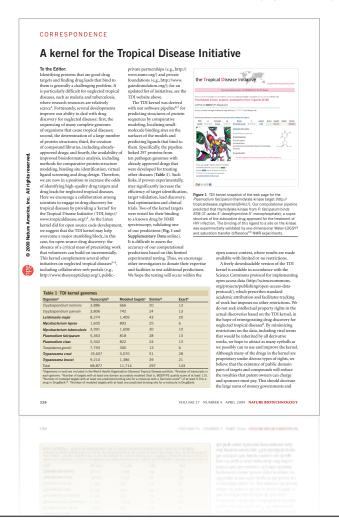
http://tropicaldisease.org/kernel



## TDI's kernel

#### http://tropicaldisease.org/kernel

L. Orti et al., Nat Biotechnol 27, 320 (Apr, 2009).



L. Orti et al., PLoS Negl Trop Dis 3, e418 (2009).



# **Acknowledgments**

http://sgu.bioinfo.cipf.es
http://tropicaldisease.org

http://integrativemodeling.org

## **COMPARATIVE MODELING Andrej Sali**

M. S. Madhusudhan Narayanan Eswar Min-Yi Shen

Ursula Pieper Ben Webb

Maya Topf (Birbeck College)

#### **MODEL ASSESSMENT**

David Eramian Min-Yi Shen Damien Devos

#### **FUNCTIONAL ANNOTATION**

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Prince Felipe Research Center

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STREP UE Grant

Marie Curie Reintegration Grant

#### **MODEL ASSESSMENT**

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Boris Turk (IJS)
Markus Gruetter (UE)
Matthias Wilmanns (EMBL)
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# **Comparative Protein Structure Prediction**MODELLER tutorial

\$>mod9v8 model.py

Marc A. Marti-Renom

http://bioinfo.cipf.es/sgu/

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# Obtaining MODELLER and related information

- MODELLER (9v8) web page
- http://www.salilab.org/modeller/
  - Download Software (Linux/Windows/Mac/Solaris)
  - ♦ HTML Manual
  - **♦ Join Mailing List**











# **Using MODELLER**

- ♦ No GUI! 😕
- Controlled by command file 88
- Script is written in PYTHON language
- You may know Python language is simple ©©

# **Using MODELLER**

- INPUT:
  - Target Sequence (FASTA/PIR format)
  - ♦ Template Structure (PDB format)
  - Python file
- OUTPUT:
  - Target-Template Alignment
  - Model in PDB format
  - Other data

# Modeling of BLBP Input

- Target: Brain lipid-binding protein (BLBP)
- ◆ BLBP sequence in PIR (Modeller) format:

```
>P1; blbp
sequence: blbp:::::::
VDAFCATWKLTDSQNFDEYMKALGVGFATRQVGNVTKPTVIISQEGGKVVIRTQCTFKNTEINFQLGEEFEETSIDDRNCKSVVRLDG
DKLIHVQKWDGKETNCTREIKDGKMVVTLTFGDIVAVRCYEKA*
```

## STEP 1: Align blbp and 1hms sequences

#### Python script for target-template alignment

```
# Example for: alignment.align()
# This will read two sequences, align them, and write the alignment
# to a file:
log.verbose()
env = environ()
aln = alignment(env)
mdl = model(env, file='1hms')
aln.append model(mdl, align codes='1hms')
aln.append(file='blbp.seg', align codes=('blbp'))
# The asl.sim.mat similarity matrix is used by default:
aln.align(gap penalties 1d=(-600, -400))
aln.write(file='blbp-1hms.ali', alignment format='PIR')
aln.write(file='blbp-1hms.pap', alignment format='PAP')
```

#### STEP 1: Align blbp and 1hms sequences

#### Python script for target-template alignment

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# Example for: alignment.align()
# This will read two sequences, align them, and write the alignment
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env = environ()
aln = alignment(env)
mdl = model(env, file='1hms')
aln.append model(mdl, align codes='1hms')
aln.append(file='blbp.seg', align codes=('blbp'))
# The asl.sim.mat similarity matrix is used by default:
aln.align(gap_penalties_1d=(-600, -400))
aln.write(file='blbp-1hms.ali', alignment format='PIR')
aln.write(file='blbp-1hms.pap', alignment format='PAP')
```

#### STEP 1: Align blbp and 1hms sequences

#### Python script for target-template alignment

```
# Example for: alignment.align()
# This will read two sequences, align them, and write the alignment
# to a file:
log.verbose()
env = environ()
aln = alignment(env)
mdl = model(env, file='1hms')
aln.append model(mdl, align codes='1hms')
aln.append(file='blbp.seq', align codes=('blbp'))
# The as1.sim.mat similarity matrix is used by default:
aln.align(gap_penalties_1d=(-600, -400))
aln.write(file='blbp-1hms.ali', alignment_format='PIR')
aln.write(file='blbp-1hms.pap', alignment_format='PAP')
```

#### STEP 1: Align blbp and 1hms sequences

#### Python script for target-template alignment

```
# Example for: alignment.align()
# This will read two sequences, align them, and write the alignment
# to a file:
log.verbose()
env = environ()
aln = alignment(env)
mdl = model(env, file='_hms')
aln.append model(mdl, align codes='lhms')
aln.append(file='blbp.seg', align codes=('blbp'))
# The asl.sim.mat similarity matrix is used by default:
aln.align(gap penalties 1d=(-600, -400))
aln.write(file='blbp-1hms.ali', alignment format='PIR')
aln.write(file='blbp-1hms.pap', alignment format='PAP')
```

# Modeling of BLBP STEP 1: Align blbp and 1hms sequences Output

```
>P1;1hms

structureX:1hms: 1 :: 131 :: undefined:undefined:-1.00:-1.00

VDAFLGTWKLVDSKNFDDYMKSLGVGFATRQVASMTKPTTIIEKNGDILTLKTHSTFKNTEISFKLGVEFDETTA

DDRKVKSIVTLDGGKLVHLQKWDGQETTLVRELIDGKLILTLTHGTAVCTRTYEKE*

>P1;blbp

sequence:blbp: :: :: : 0.00: 0.00

VDAFCATWKLTDSQNFDEYMKALGVGFATRQVGNVTKPTVIISQEGGKVVIRTQCTFKNTEINFQLGEEFEETSI

DDRNCKSVVRLDGDKLIHVQKWDGKETNCTREIKDGKMVVTLTFGDIVAVRCYEKA*
```

# Modeling of BLBP STEP 1: Align blbp and 1hms sequences Output

```
>P1;1hms

structureX:1hms: 1 :: 131 :: undefined:undefined:-1.00:-1.00

VDAFLGTWKLVDSKNFDDYMKSLGVGFATRQVASMTKPTTIIEKNGDILTLKTHSTFKNTEISFKLGVEFDETTA

DDRKVKSIVTLDGGKLVHLQKWDGQETTLVRELIDGKLILTLTHGTAVCTRTYEKE*

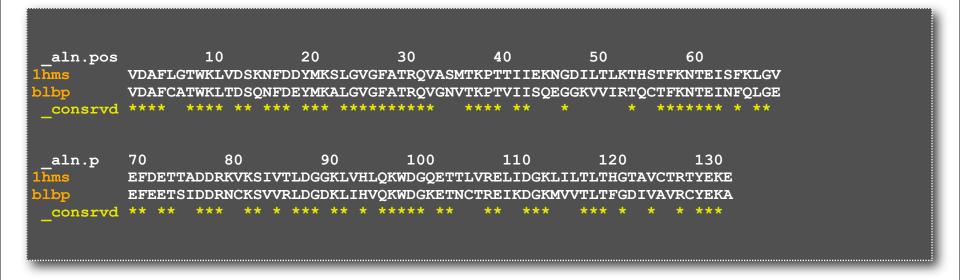
>P1;blbp

sequence:blbp: :: :: : 0.00: 0.00

VDAFCATWKLTDSQNFDEYMKALGVGFATRQVGNVTKPTVIISQEGGKVVIRTQCTFKNTEINFQLGEEFEETSI

DDRNCKSVVRLDGDKLIHVQKWDGKETNCTREIKDGKMVVTLTFGDIVAVRCYEKA*
```

# Modeling of BLBP STEP 1: Align blbp and 1hms sequences Output



# STEP 2: Model the blbp structure using the alignment from step 1.

#### Python script for model building

```
# Homology modelling by the automodel class
from modeller.automodel import * # Load the automodel class
log.verbose()
                                     # request verbose output
env = environ()
                                     # create a new MODELLER environment
# directories for input atom files
env.io.atom files directory = './:../atom files'
a = automodel(env,
             alnfile = 'blbp-1hms.ali', # alignment filename
             knowns = '1hms',
                                           # codes of the templates
             sequence = 'blbp')
                                            # code of the target
a.starting model= 1
                                 # index of the first model
a.ending model = 1
                                 # index of the last model
                                  # (determines how many models to calculate)
                                  # do the actual homology modelling
a.make()
```

# STEP 2: Model the blbp structure using the alignment from step 1.

### Python script for model building

```
# Homology modelling by the automodel class
from modeller.automodel import * # Load the automodel class
log.verbose()
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# STEP 2: Model the blbp structure using the alignment from step 1.

### Python script for model building

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                                     # request verbose output
env = environ()
                                     # create a new MODELLER environment
# directories for input atom files
env.io.atom files directory = './:../atom files'
a = automodel(env,
              lnfile = 'blbp-1hms.ali', # alignment filename
                                          # codes of the templates
               equence = 'blbp') # code of the target
                     # index of the first model
a.starting model= 1
                                 # index of the last model
                                  # (determines how many models to calculate)
                                  # do the actual homology modelling
a.make()
```

# STEP 2: Model the blbp structure using the alignment from step 1.

#### Python script for model building

#### PDB file

#### Can be viewed with Chimera

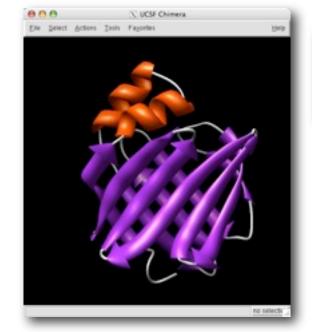
http://www.cql.ucsf.edu/chimera/

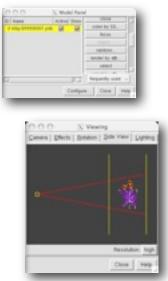
#### Rasmol

http://www.openrasmol.org

#### **PyMol**

http://pymol.sourceforge.net/

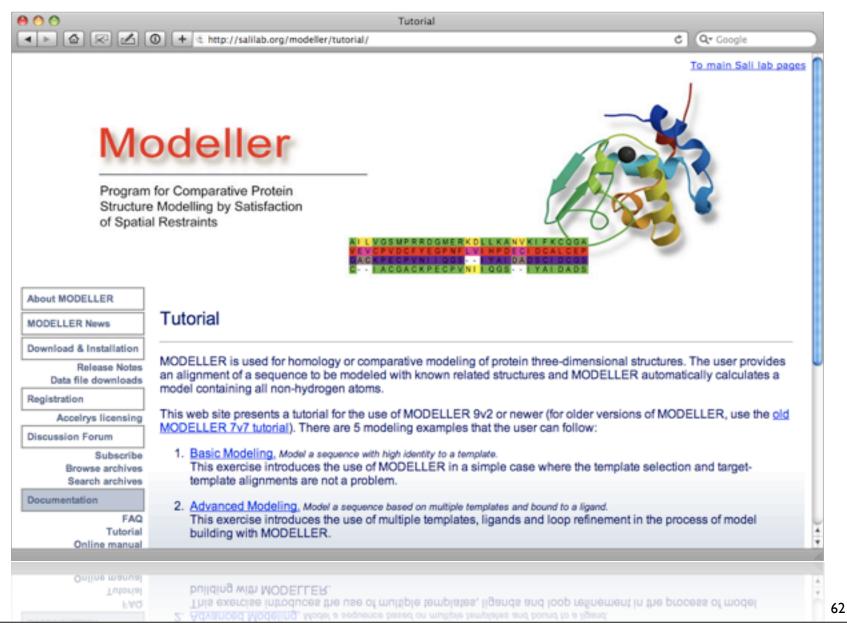




Model file →

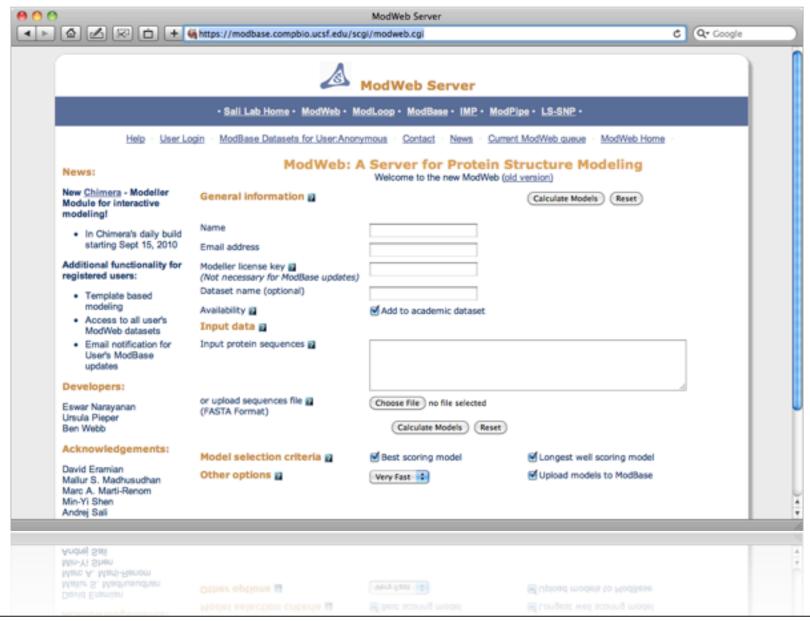
blbp.B9990001.pdb

#### http://www.salilab.org/modeller/tutorial/



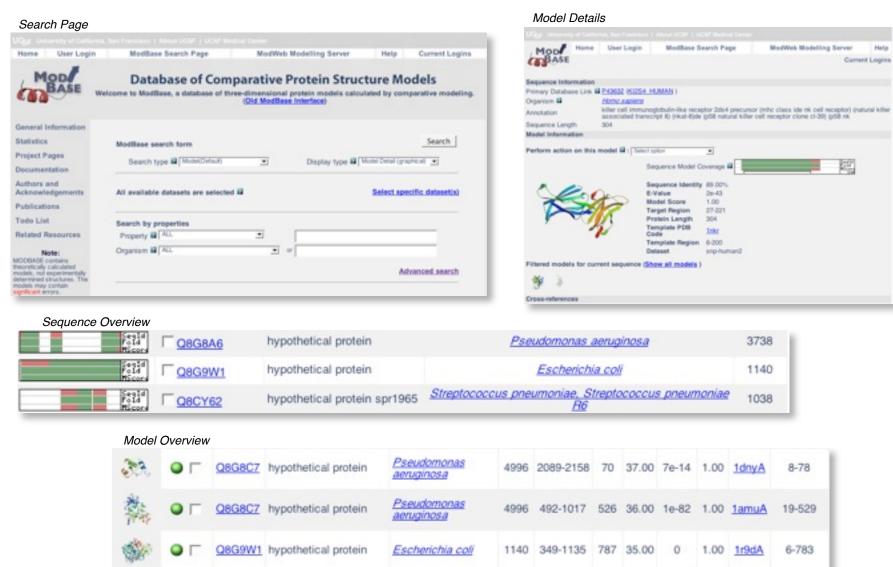
## **MODWEB**

http://salilab.org/modweb



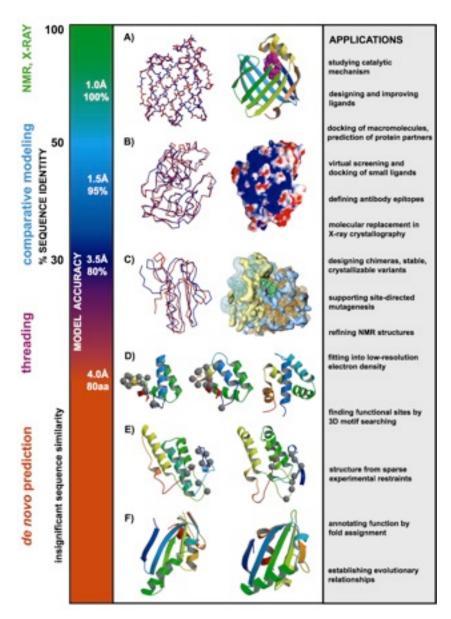
## **MODBASE**

http://salilab.org/modbase



Pieper et al. (2004) Nucleic Acids Research 32, D217-D222

## "take home" message



#### Download...



http://salilab.org/modeller/download installation.html

http://172.24.76.30/files

PLEASE... REGISTER TO GET A LICENSE!

MODELIRANJE