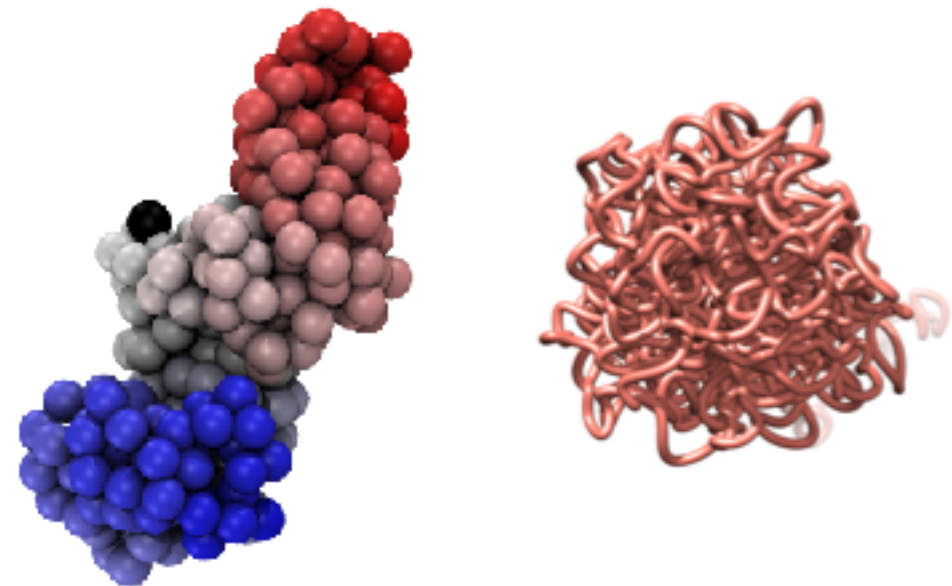


Genome structure dynamics using sparse interaction datasets

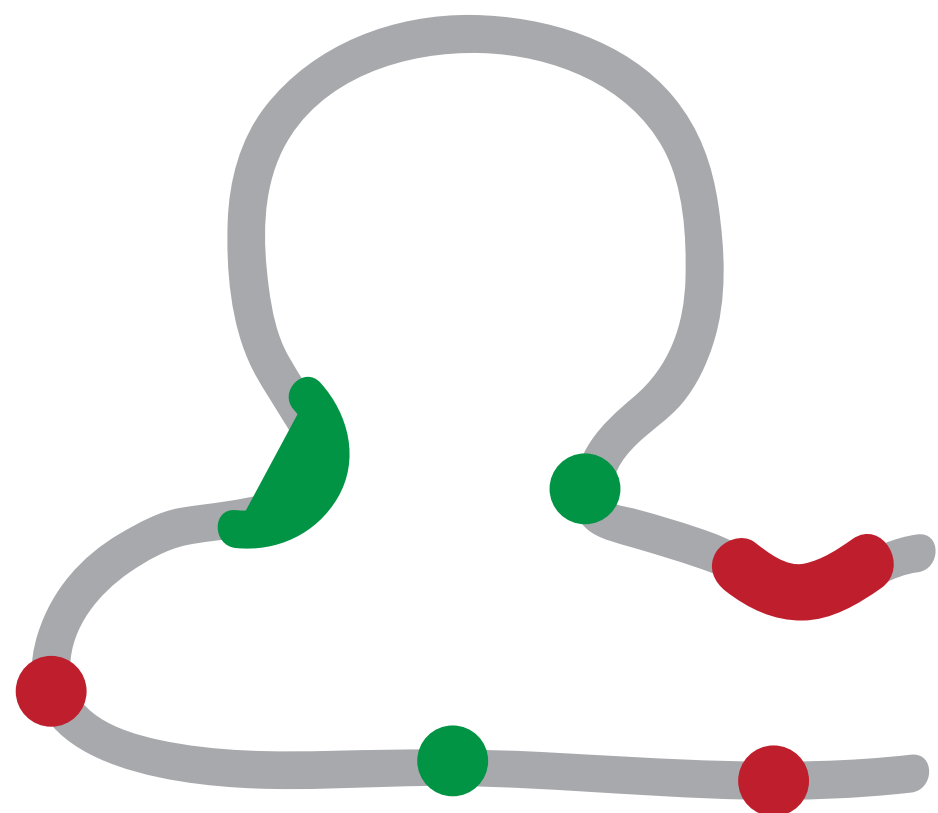
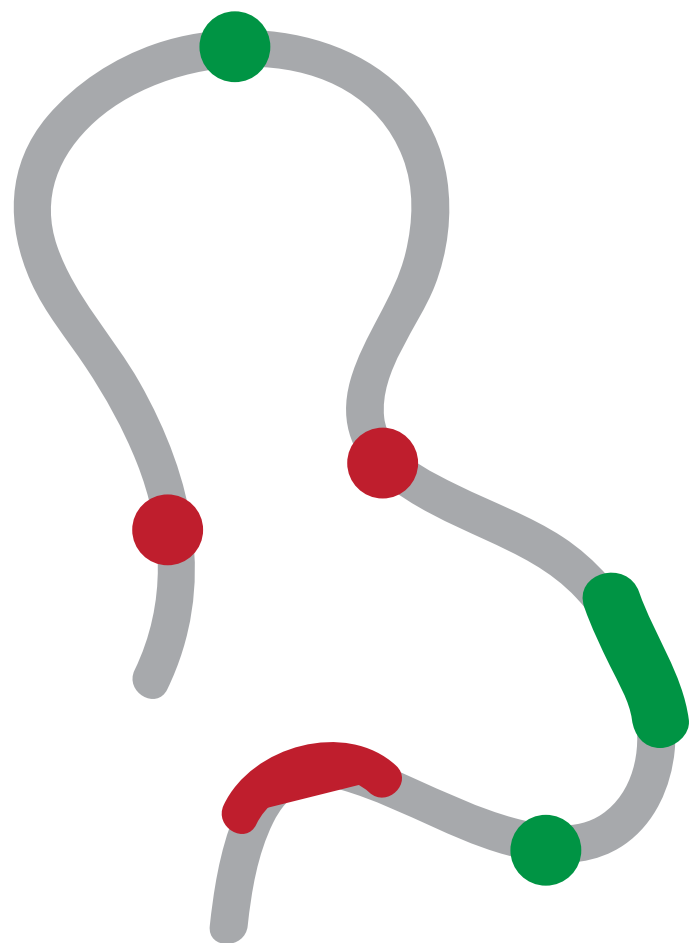
Marc A. Marti-Renom

Structural Genomics Group (ICREA, CNAG-CRG)



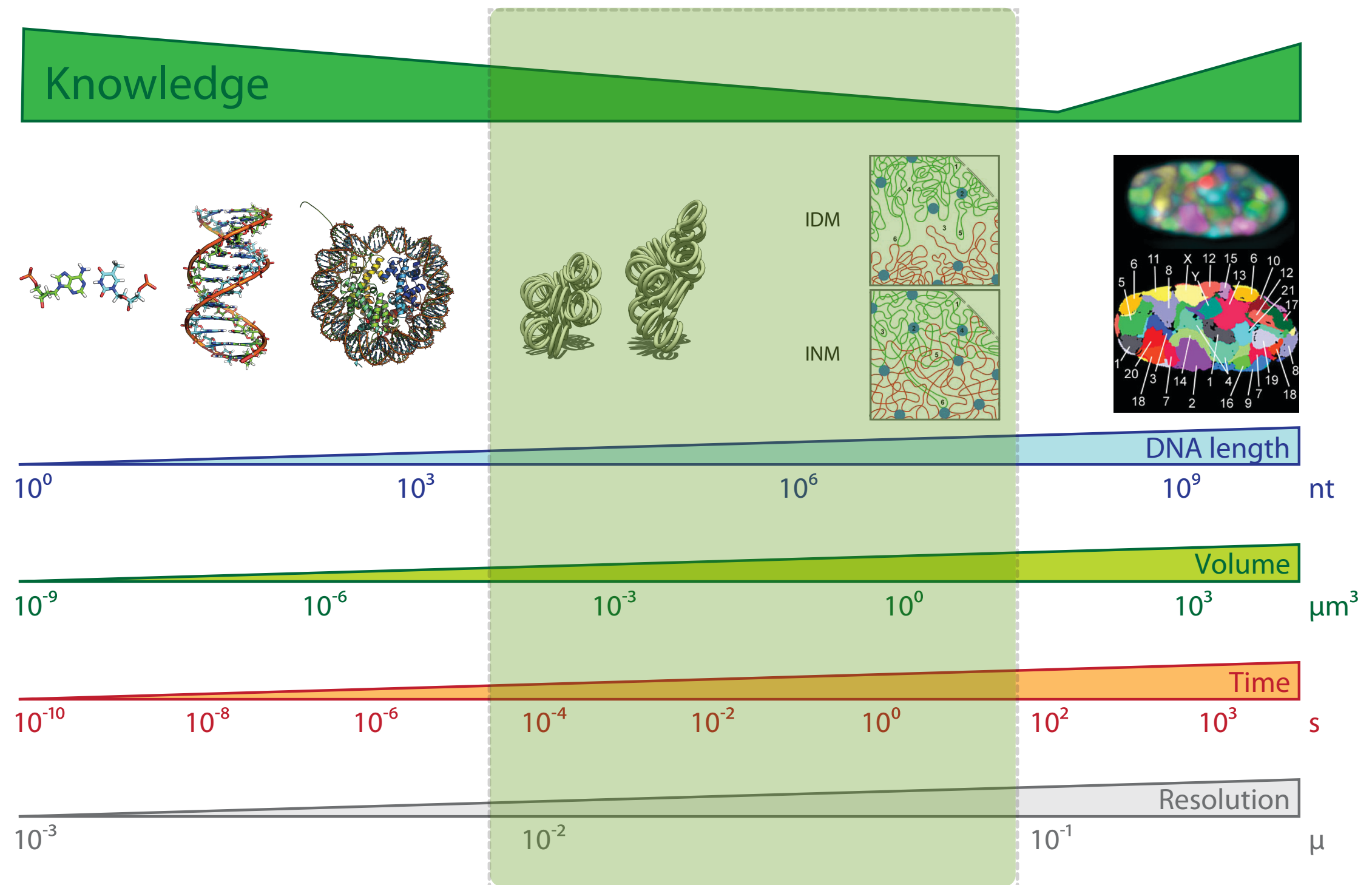
<http://marciuslab.org>
<http://3DGenomes.org>
<http://cnag.crg.eu>

cnag **CRG**  **ICREA**



Resolution Gap

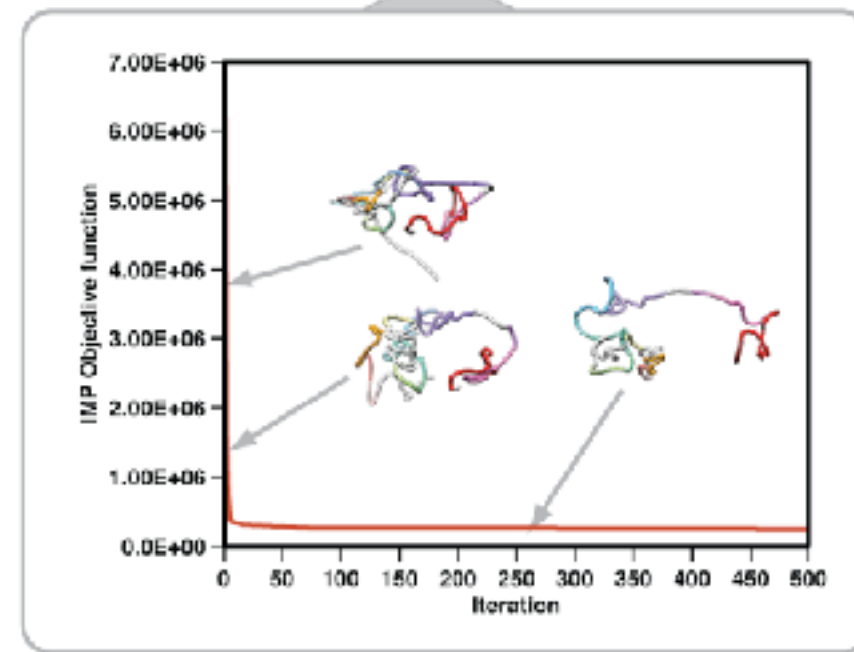
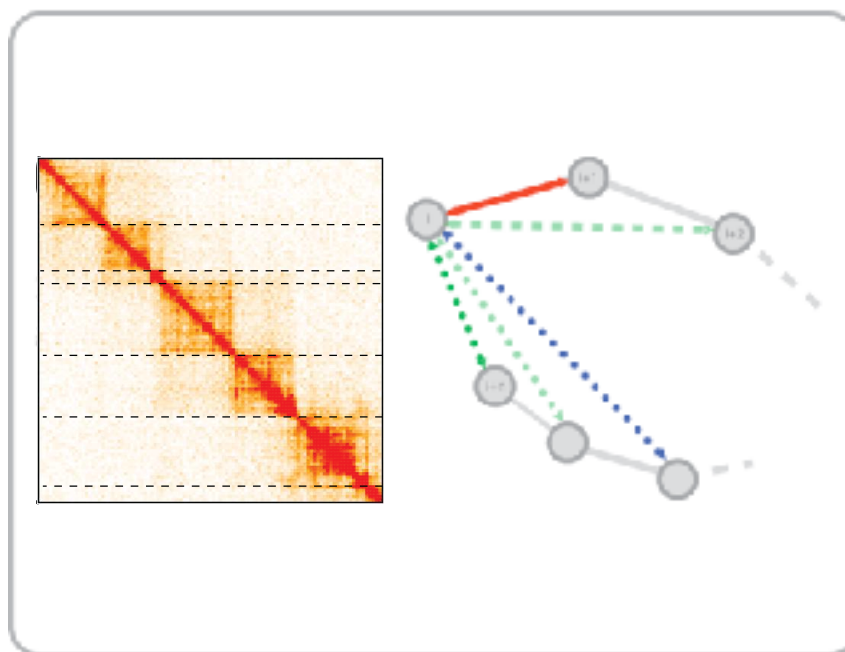
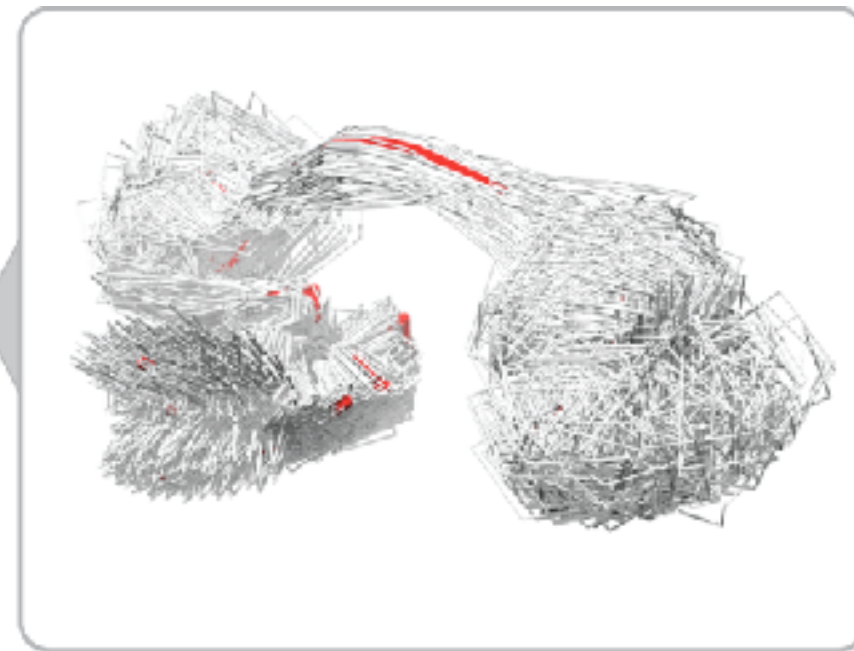
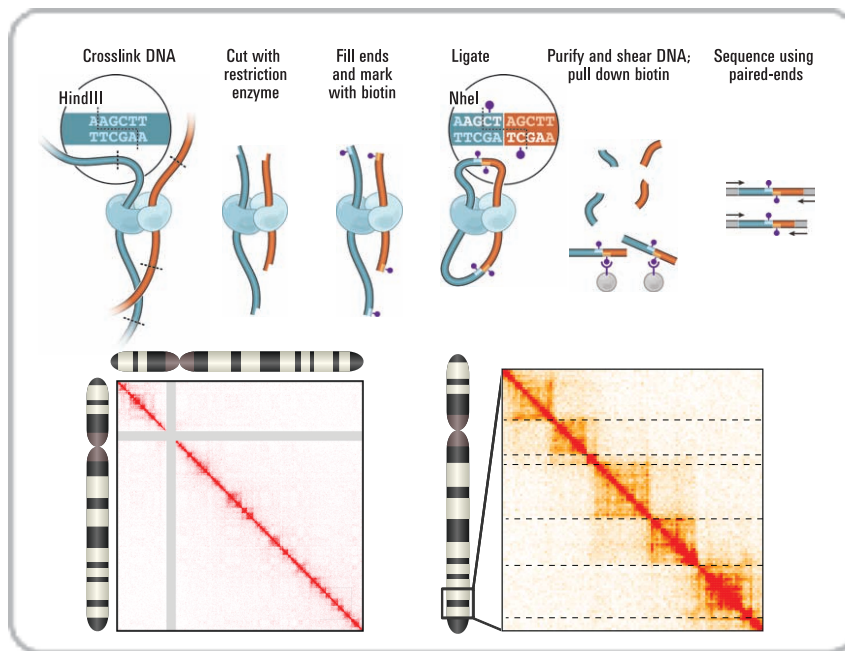
Marti-Renom, M. A. & Mirny, L. A. PLoS Comput Biol 7, e1002125 (2011)



Hybrid Method

Baù, D. & Marti-Renom, M. A. Methods 58, 300–306 (2012).

Experiments

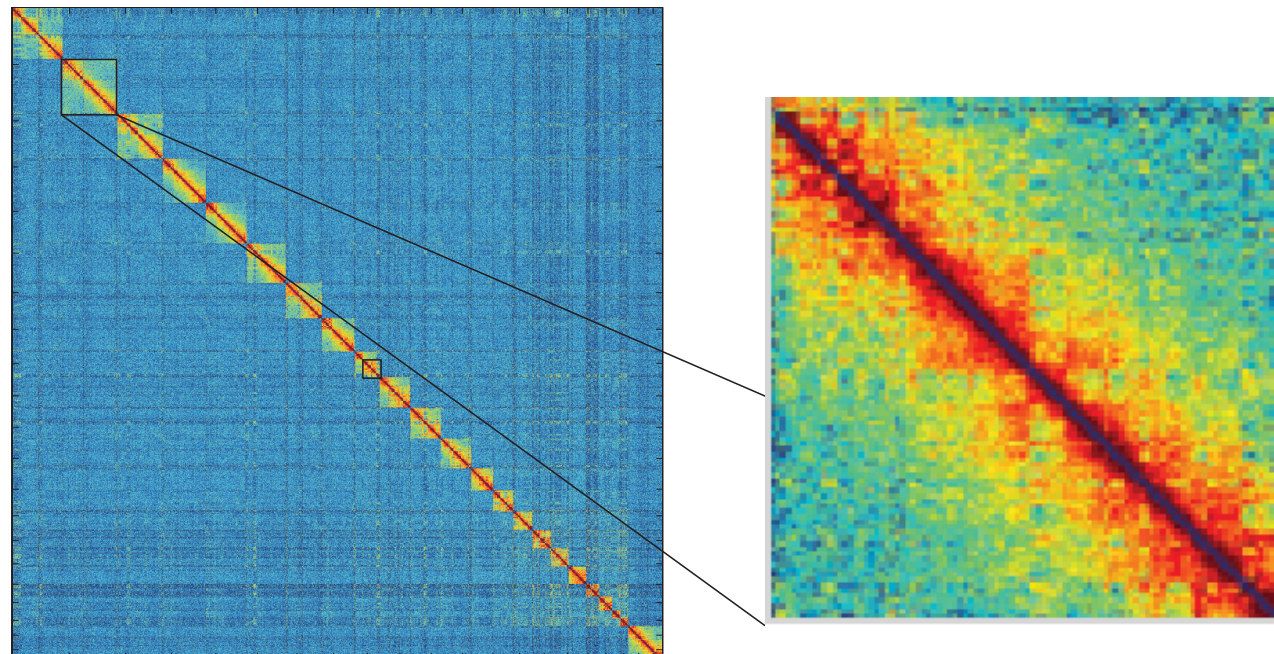
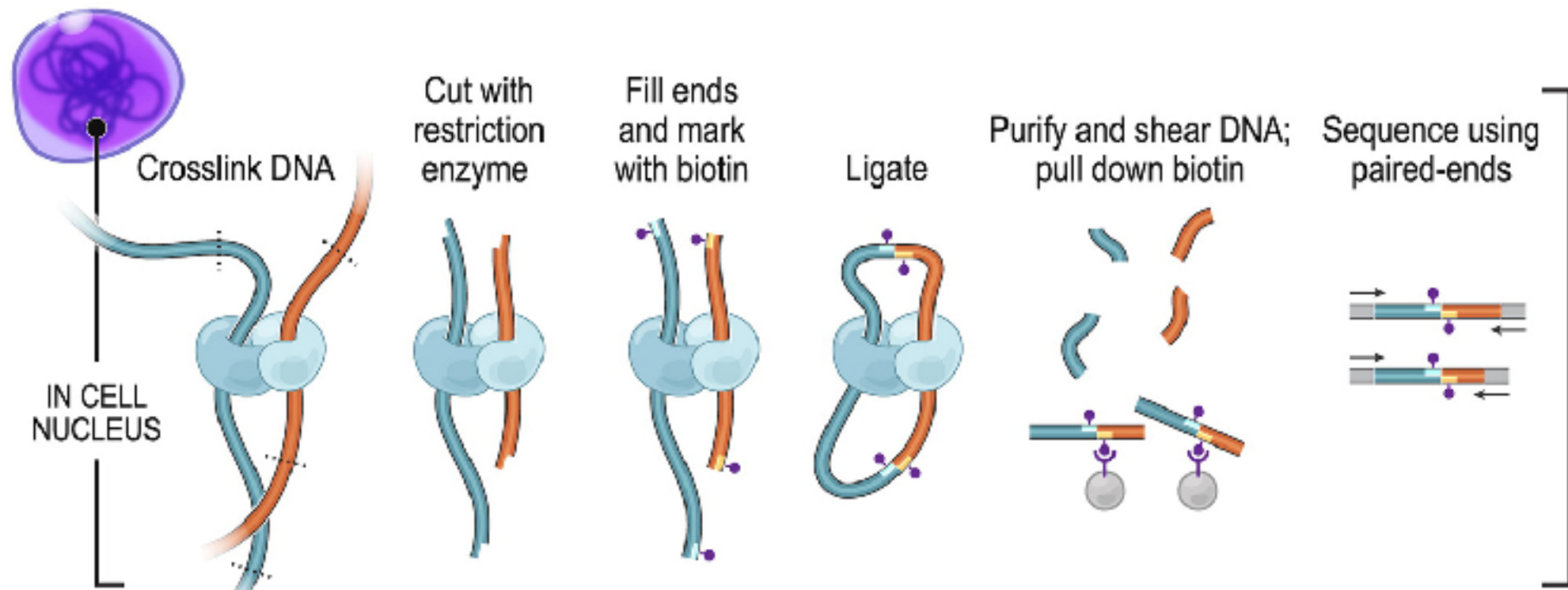


Computation

Chromosome Conformation Capture

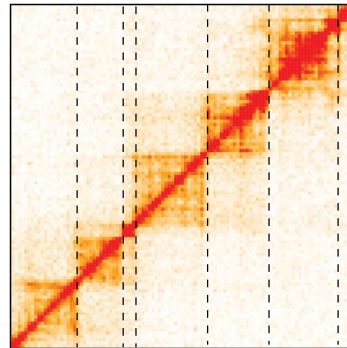
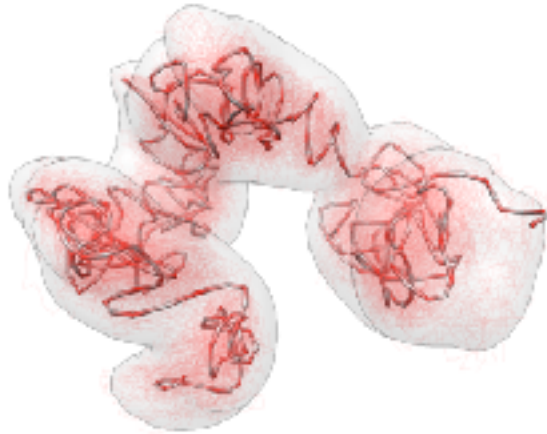
Dekker, J., Rippe, K., Dekker, M., & Kleckner, N. (2002). *Science*, 295(5558), 1306–1311.

Lieberman-Aiden, E., et al. (2009). *Science*, 326(5950), 289–293.

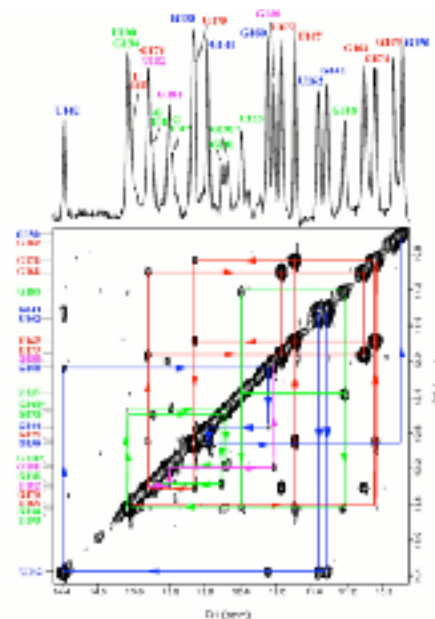
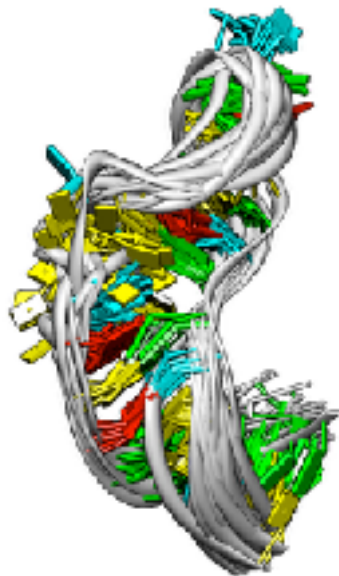


Restraint-based Modeling

Baù, D. & Marti-Renom, M. A. Methods 58, 300–306 (2012).



Chromosome structure determination
3C-based data

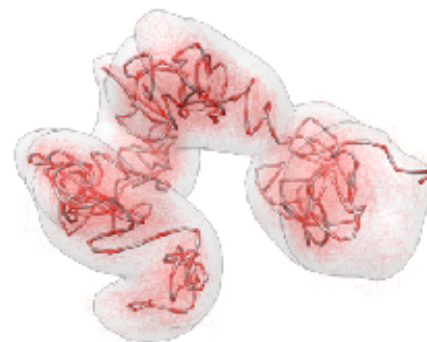
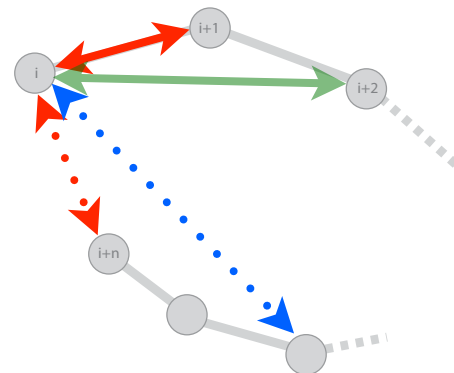
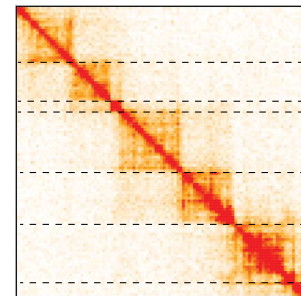


Biomolecular structure determination
2D-NOESY data



<http://3DGenomes.org>

Serra, F., Baù, D. et al. PLOS CB (2017)



FastQ files to Maps

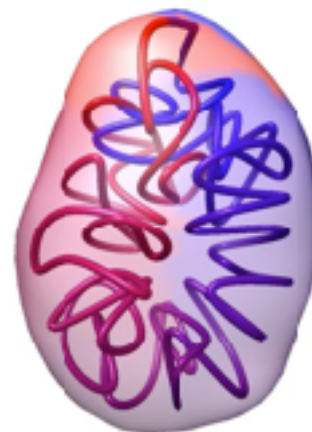
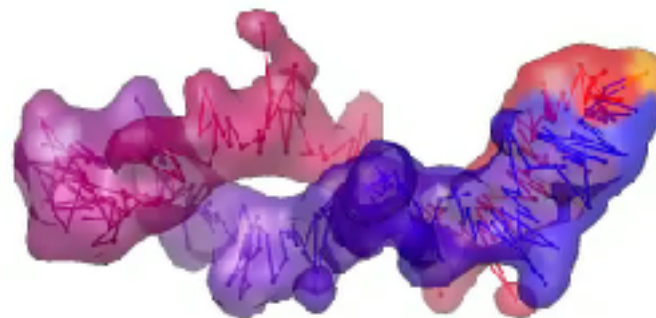
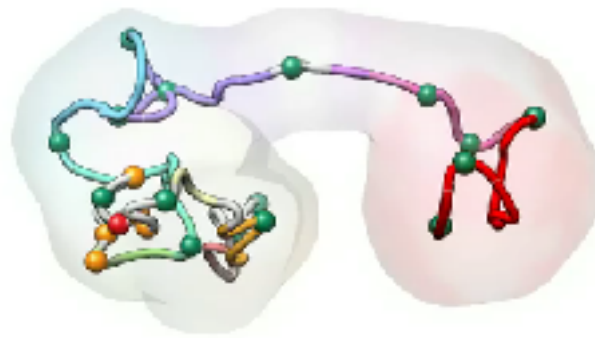
Map analysis

Model building

Model analysis

TADbit previous applications...

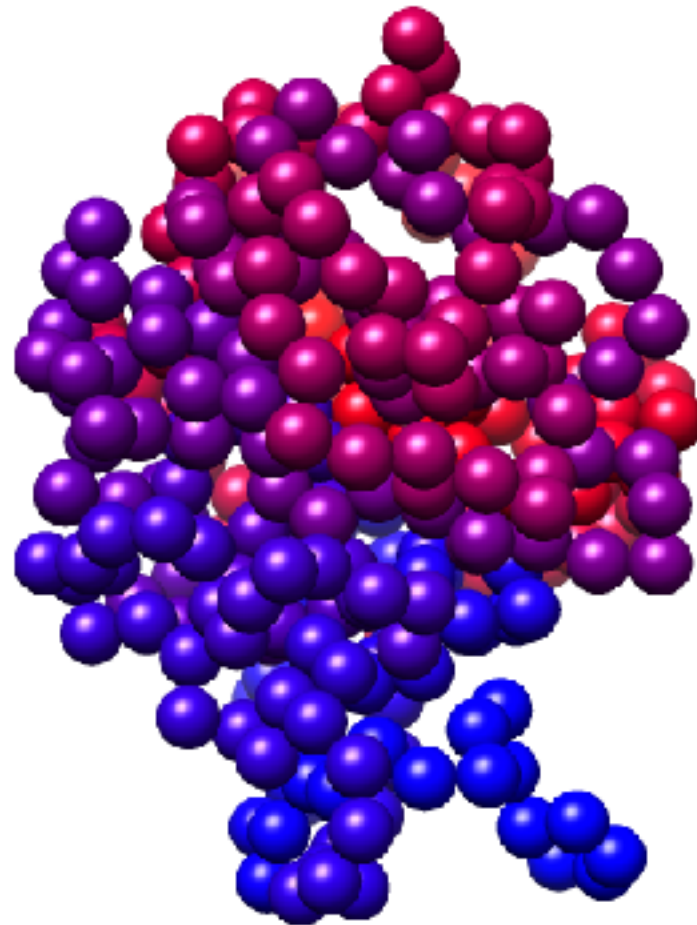
Baù, D. et al. Nat Struct Mol Biol (2011)
Umbarger, M. A. et al. Mol Cell (2011)
Le Dily, F. et al. Genes & Dev (2014)
Trussart M. et al. Nature Communication (2017)
Cattoni et al. Nature Communication (2017)
Stadhouders, R., Vidal, E. et al. Nature Genetics (2018)



TADdyn. Dynamics of chromatin



Marco Di Stefano



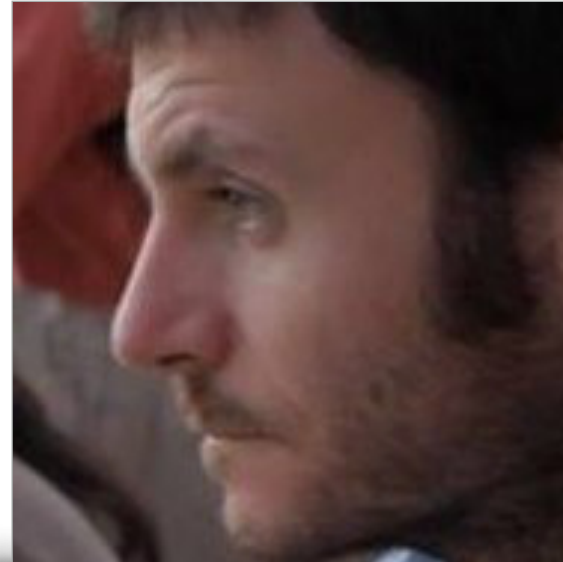
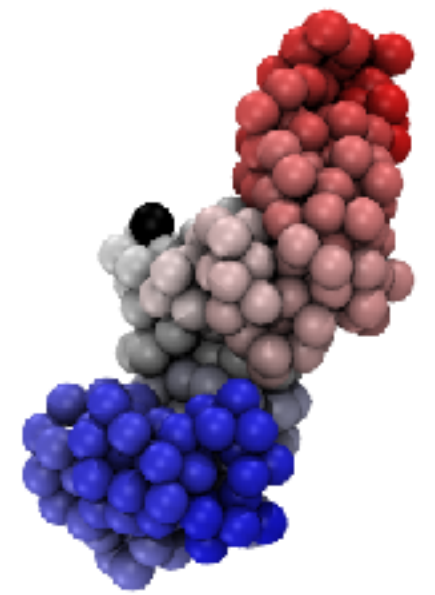
$$\mathcal{H}_{intra} = \sum_{i=1}^N U_{FENE}(i, i+1) + U_{br}(i, i+1, i+2) + \sum_{j=i+1}^N U_{LJ}(i, j)$$

Chain-connectivity interaction

Bending

Lennard-Jones Potential

Exploring the time dependent structural rearrangements of SOX2 locus during transdifferentiation



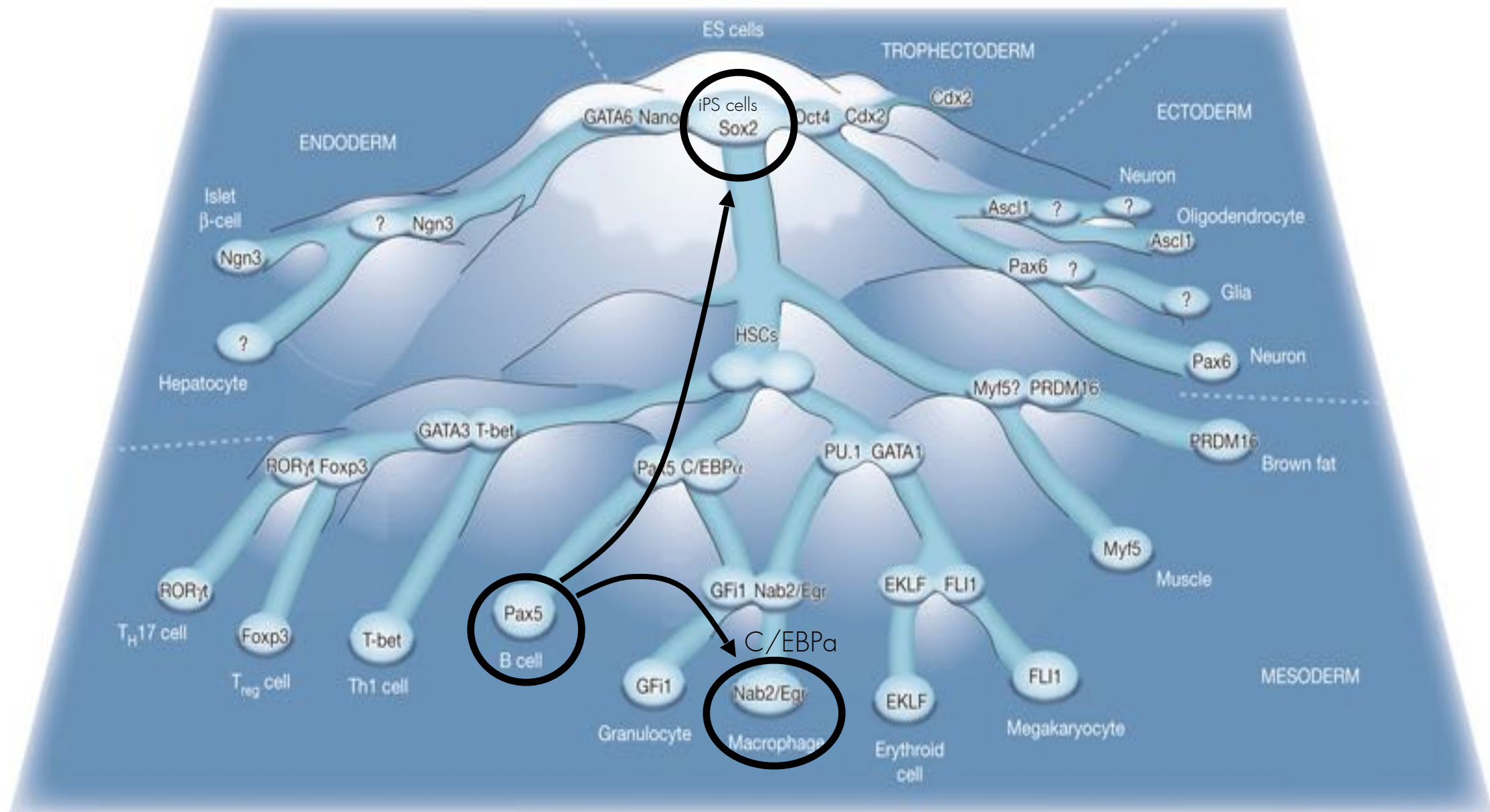
Stadhouders, R., Vidal, E. et al. (2018) Nature Genetics



Marco di Stefano

Transcription factors dictate cell fate

Graf & Enver (2009) Nature



Transcription factors (TFs) determine cell identity through gene regulation

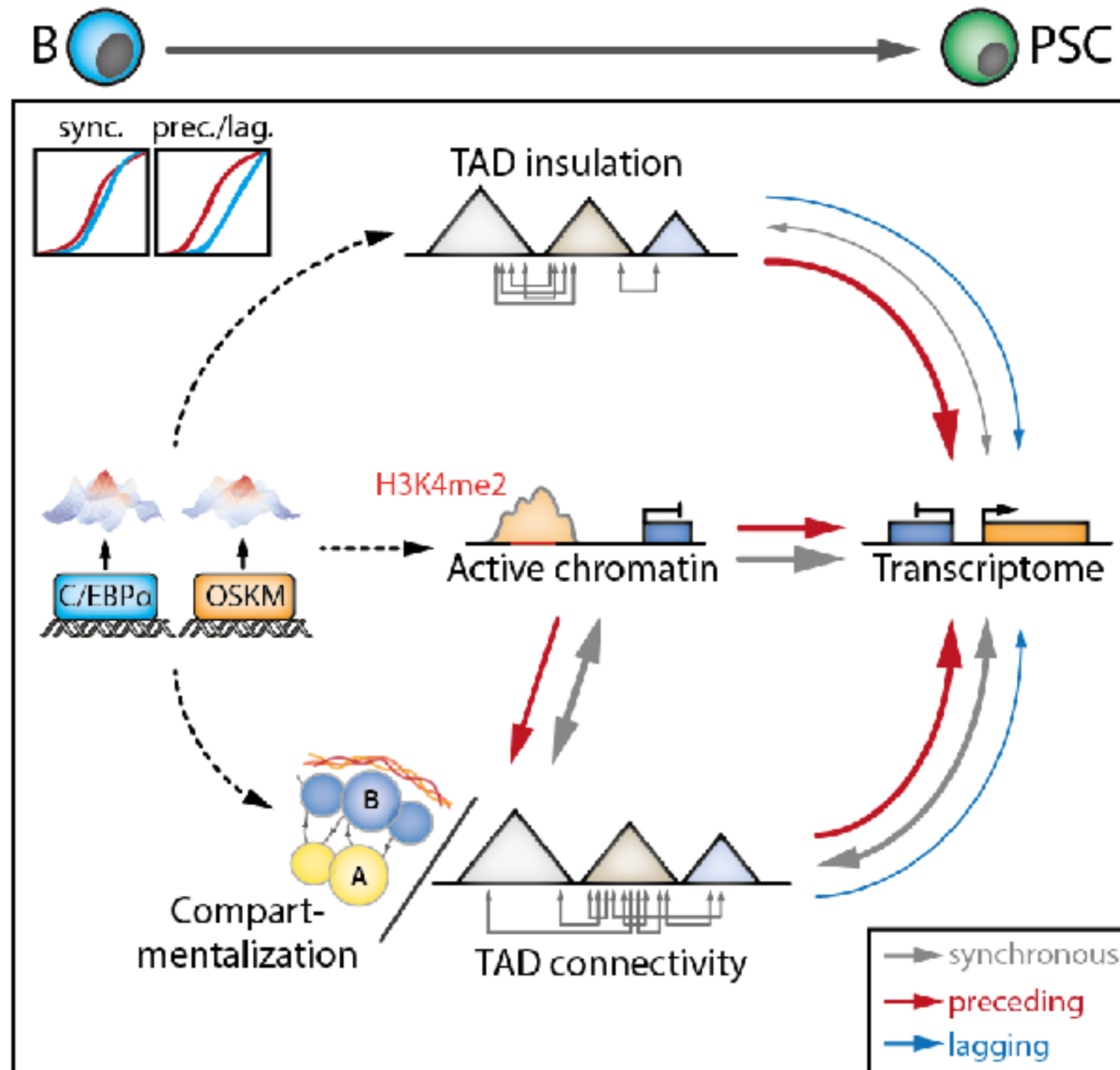
Normal 'forward' differentiation

Cell fates can be converted by enforced TF expression

Transdifferentiation or reprogramming

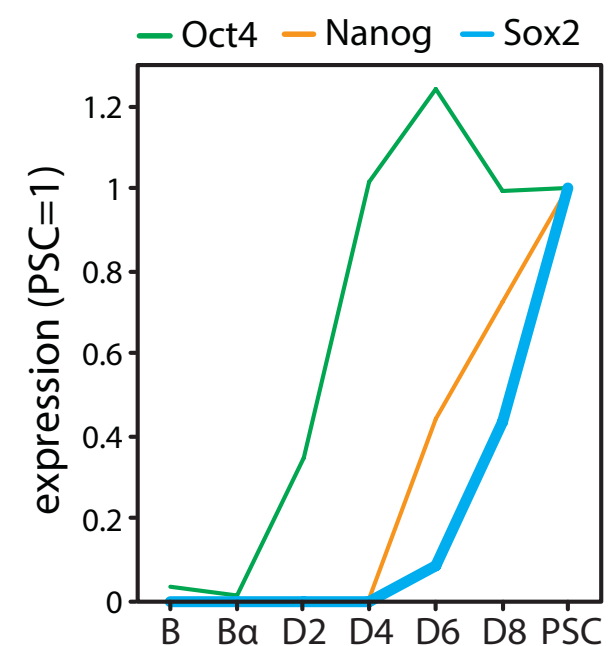
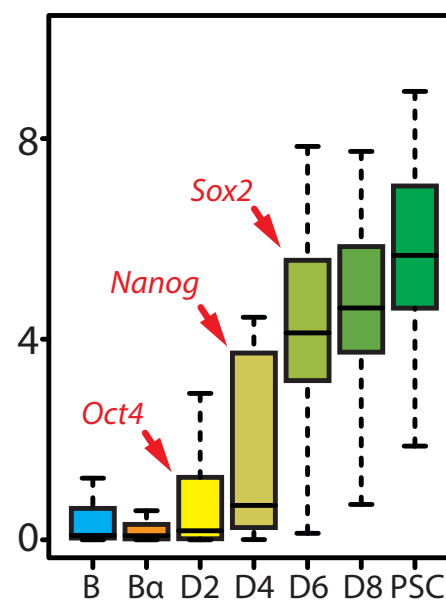
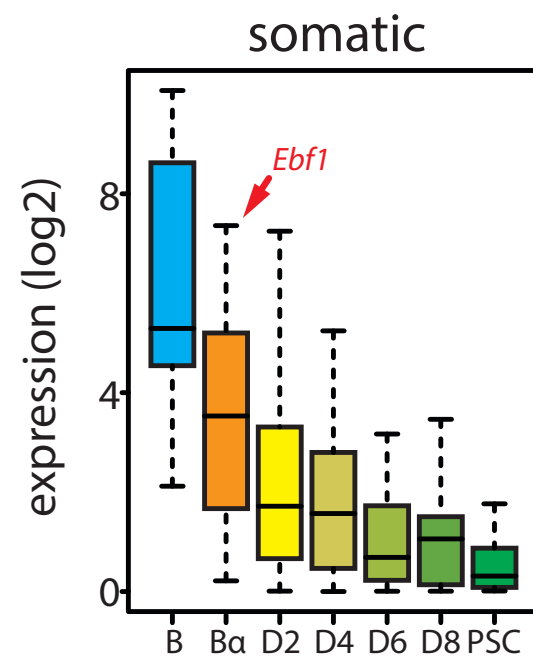
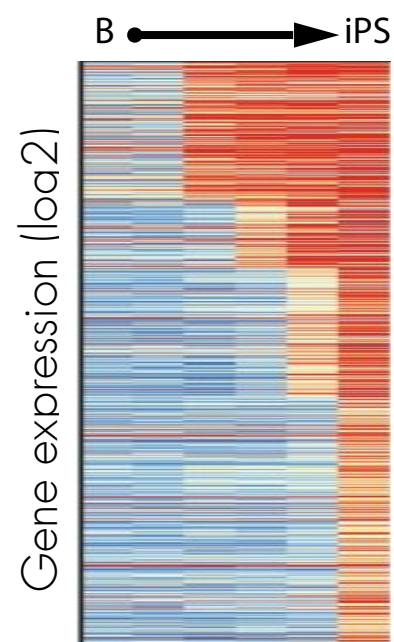
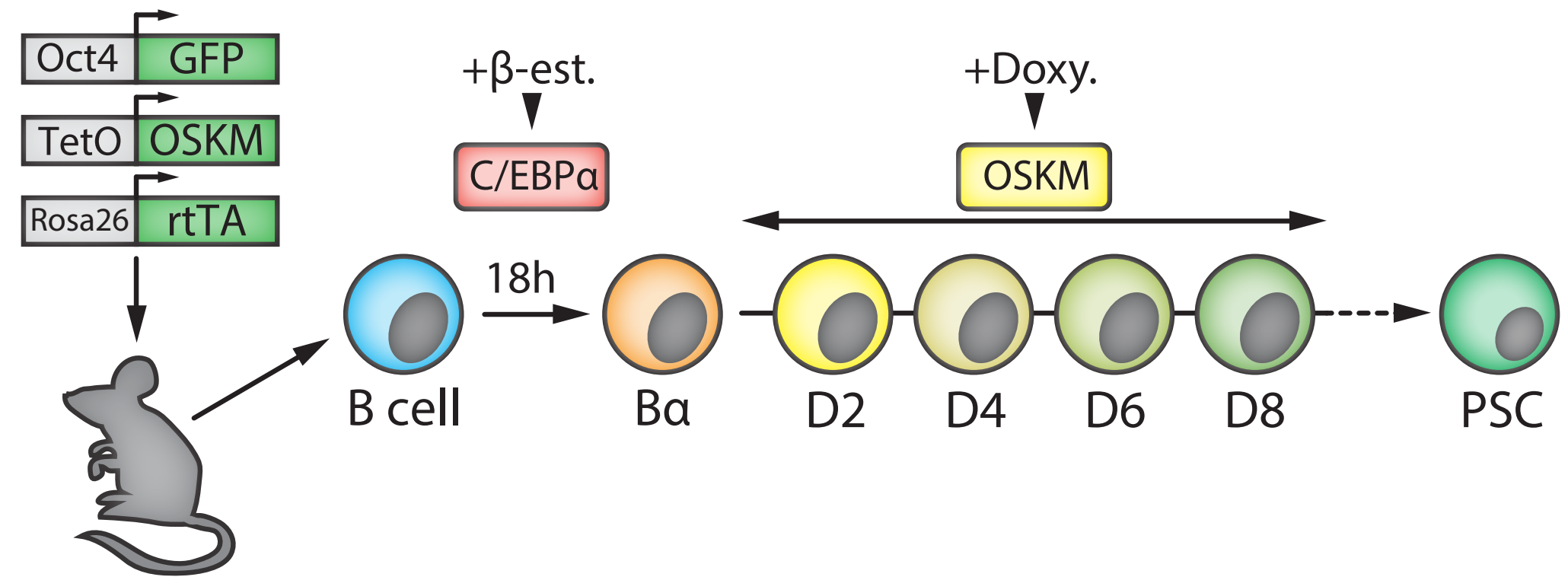
Interplay: topology, gene expression & chromatin

Stadhouders, R., Vidal, E. et al. (2018) Nature Genetics



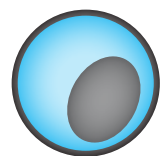
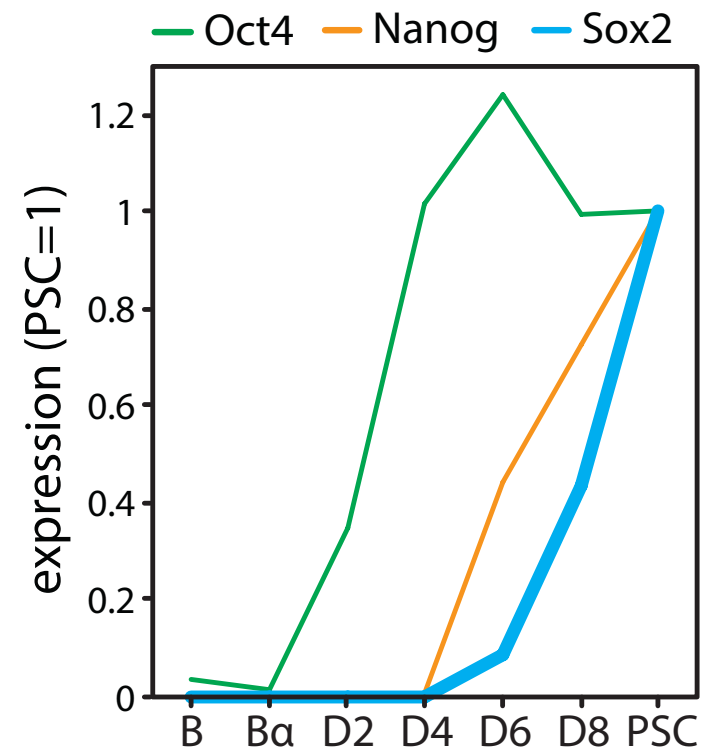
Reprogramming from B to PSC

Stadhouders, R., Vidal, E. et al. (2018) Nature Genetics

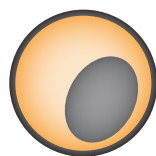


Hi-C maps of reprogramming from B to PSC

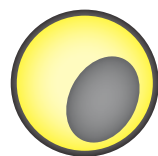
The SOX2 locus



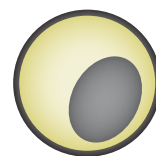
B cell



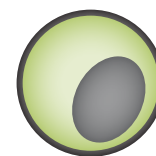
Bα



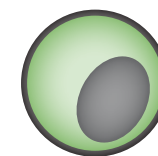
D2



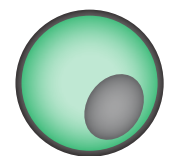
D4



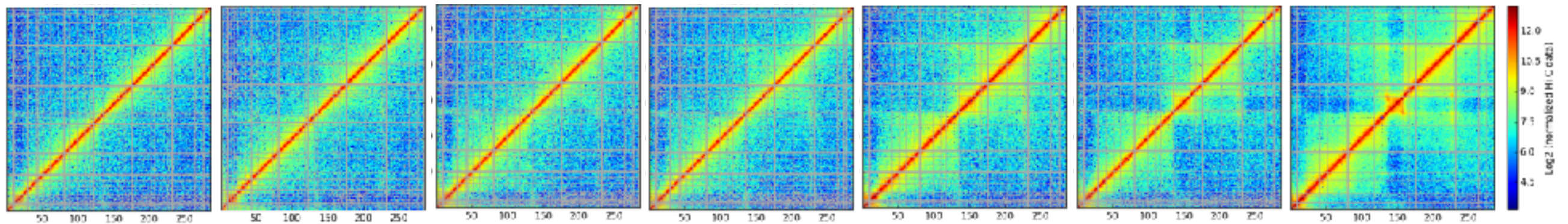
D6



D8

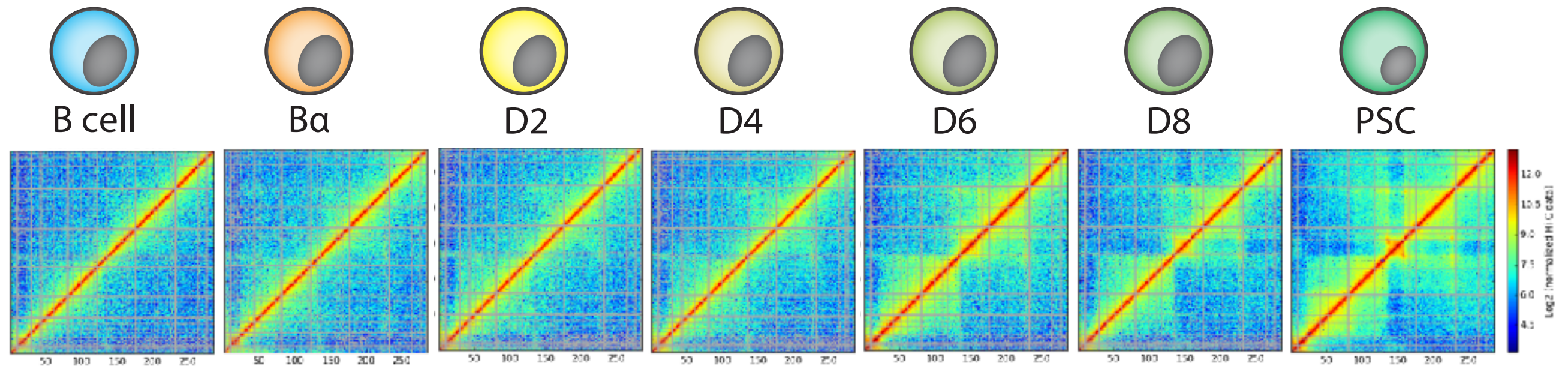


PSC



Hi-C maps of reprogramming from B to PSC

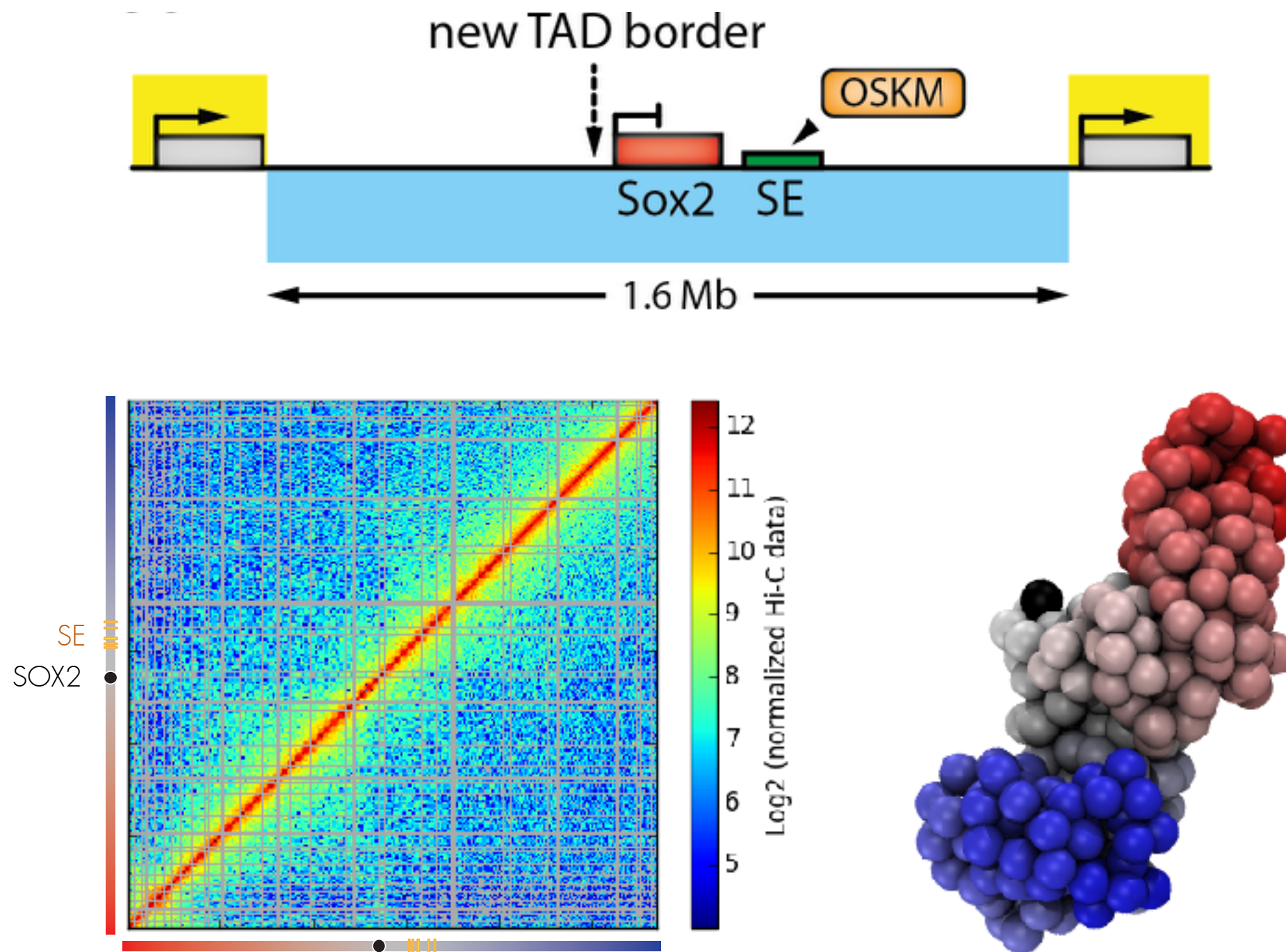
The SOX2 locus



How does these structural rearrangements interplay with the transcription activity?

What are the main drivers of structural transitions?

TADbit modeling of SOX2 from B cells Hi-C

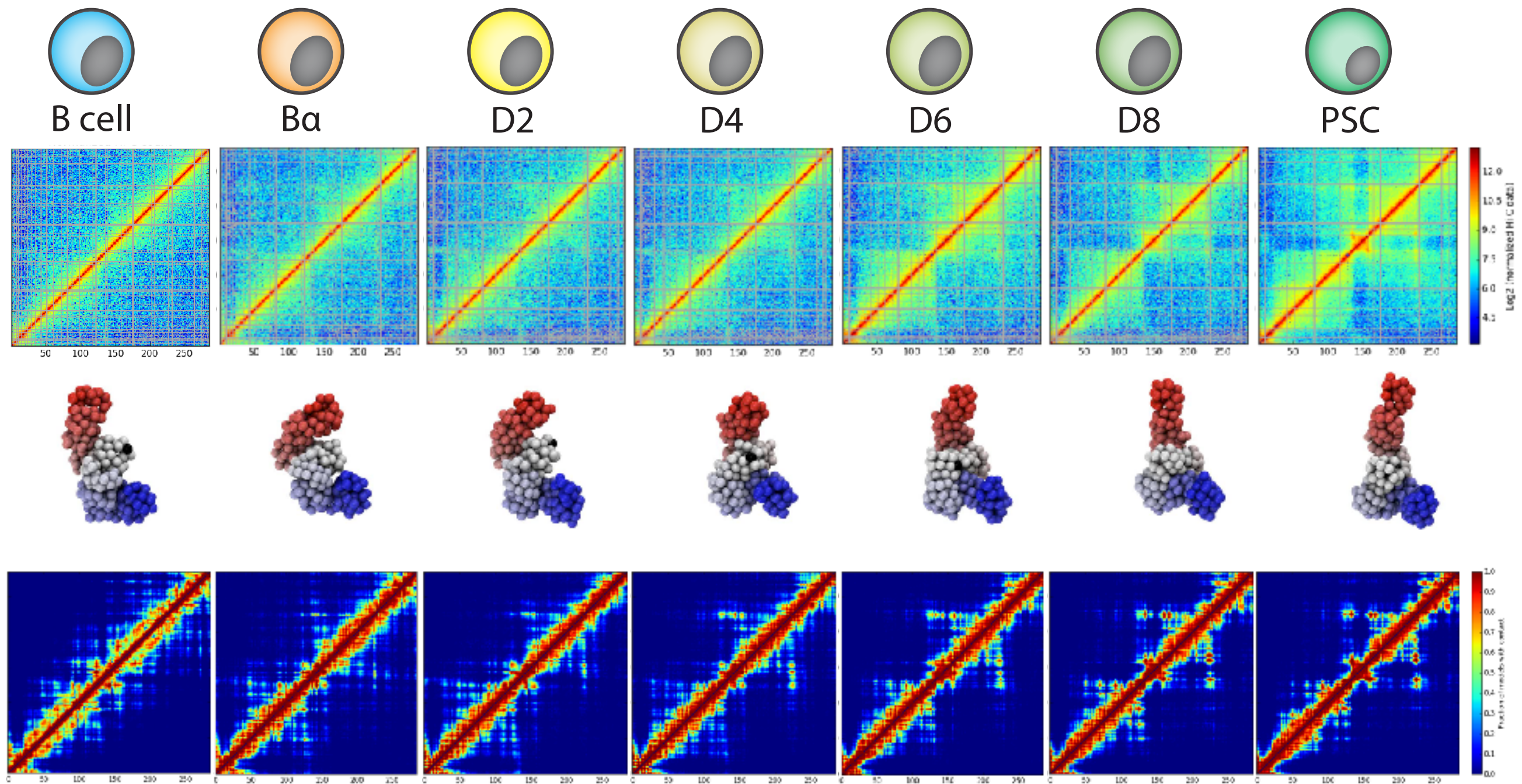


Optimal IMP parameters

lowfreq=0 , upfreq=1 , maxdist=200nm, dcutoff=125nm, particle size=50nm (5kb)

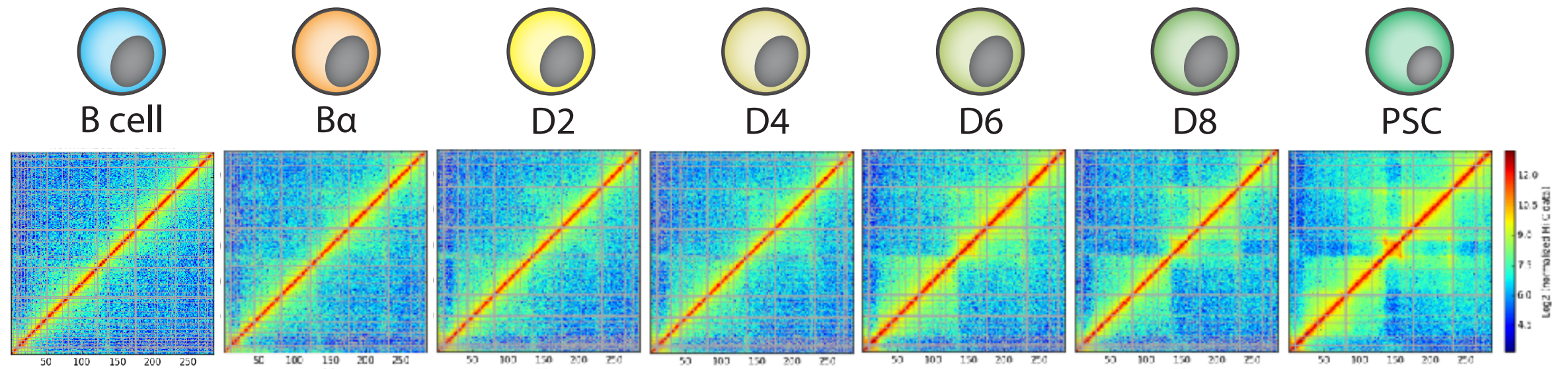
Models of reprogramming from B to PSC

The SOX2 locus



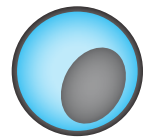
TADdyn: from time-series Hi-C maps to dynamic restraints

The SOX2 locus

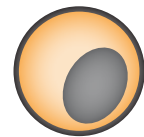


TADdyn: from time-series Hi-C maps to dynamic restraints

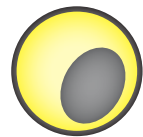
The SOX2 locus



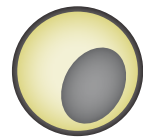
B cell



Bα



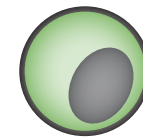
D2



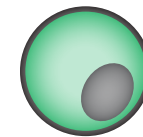
D4



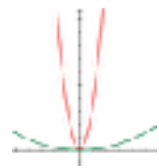
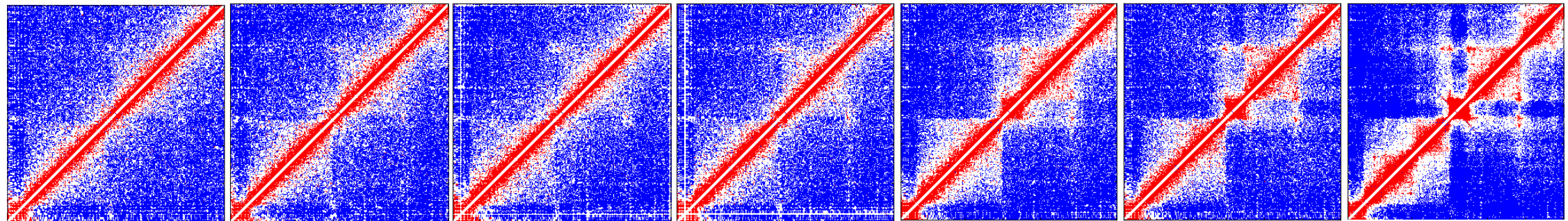
D6



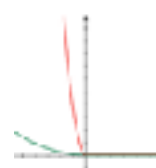
D8



PSC



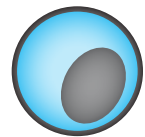
Harmonic



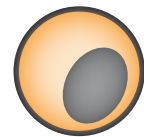
HarmonicLowerBound

TADdyn: from time-series Hi-C maps to dynamic restraints

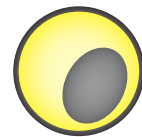
The SOX2 locus



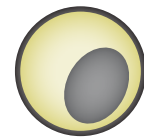
B cell



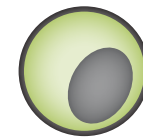
B α



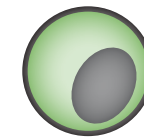
D2



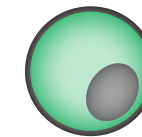
D4



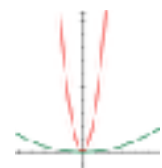
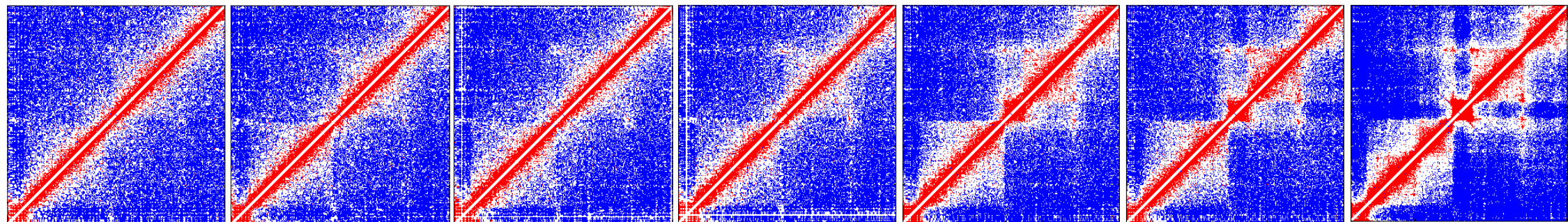
D6



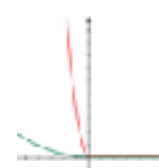
D8



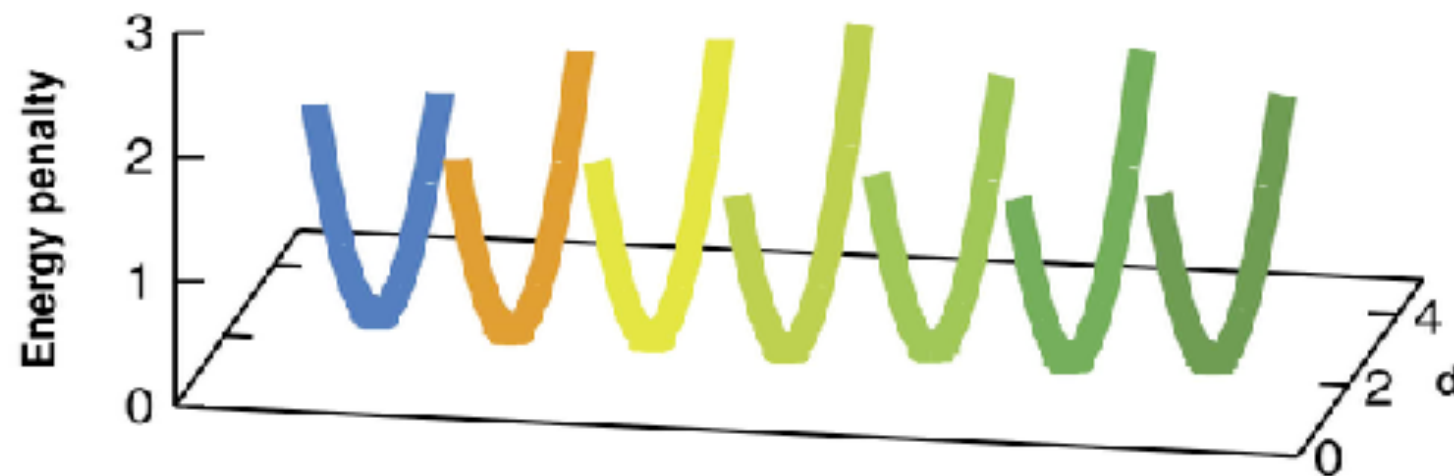
PSC



■ Harmonic



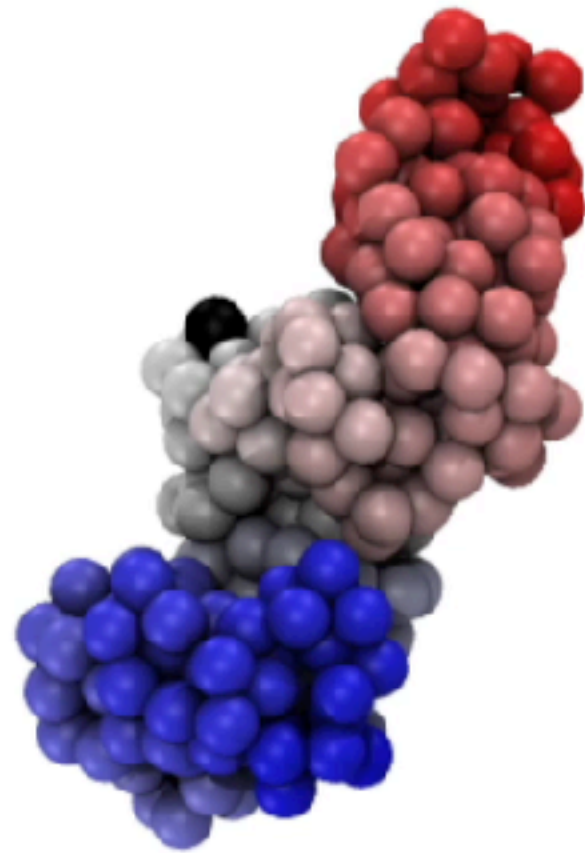
■ HarmonicLowerBound



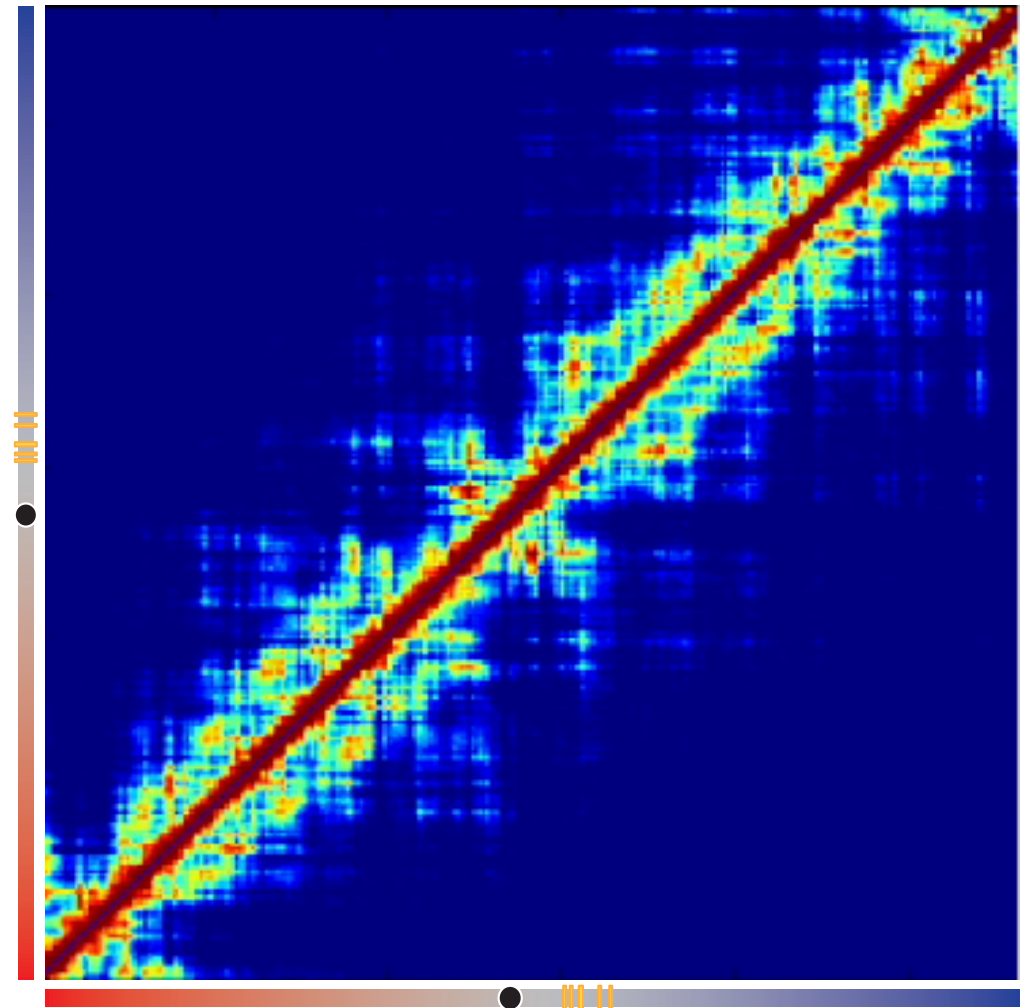
Transition	Stable	Vanishing	Raising
B -> B α	18,612	6,984	7,290
B α -> D2	18,512	7,390	6,687
D2 -> D4	18,369	6,830	6,893
D4 -> D6	18,971	6,291	7,289
D6 -> D8	20,167	6,093	6,250
D8 -> ES	20,679	5,738	6,173

SOX2 locus structural changes from B to PSC

Contacts

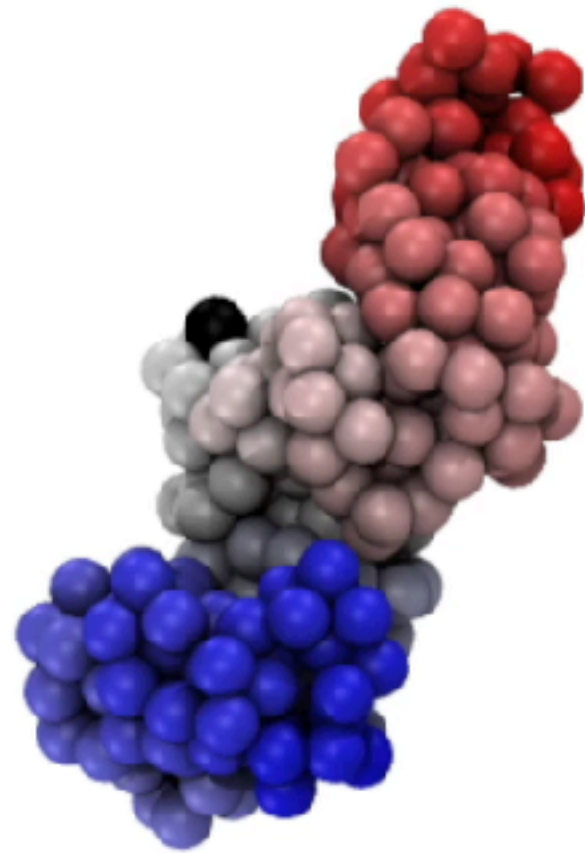


SE
SOX2

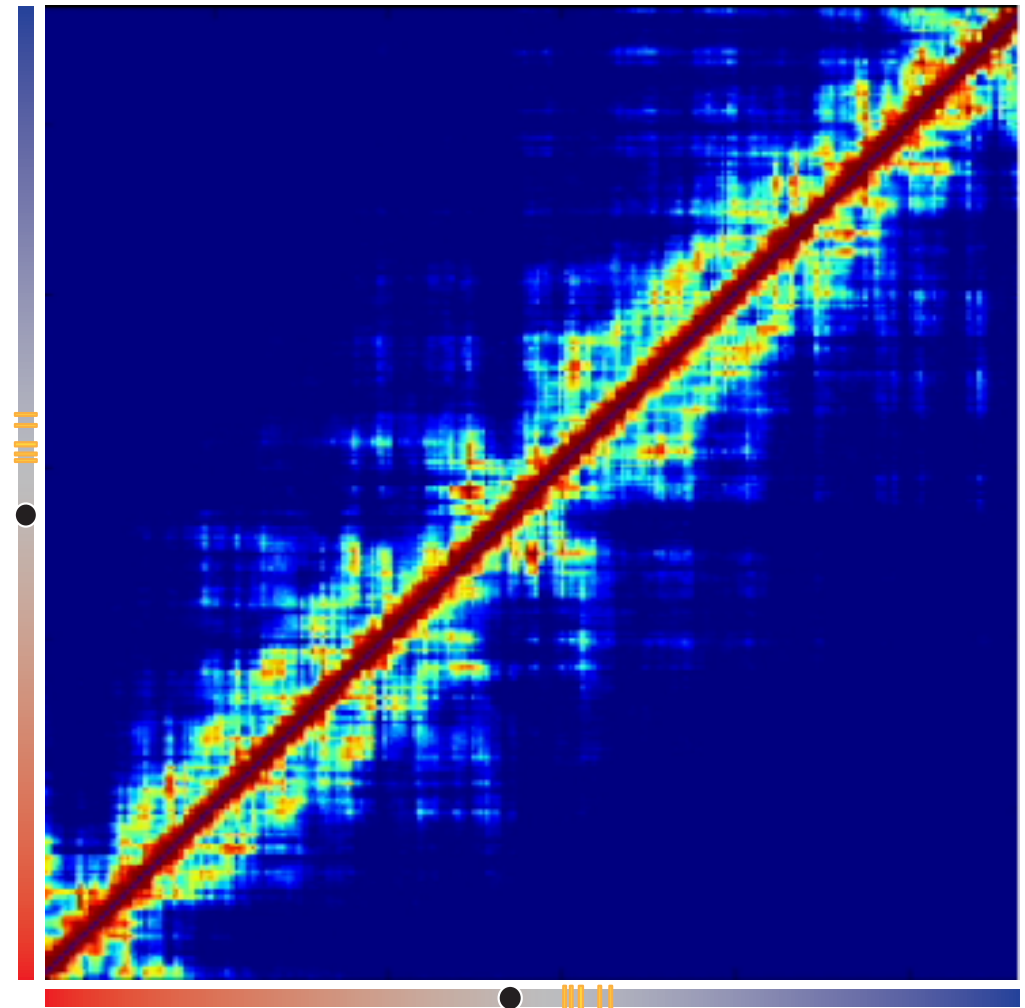


SOX2 locus structural changes from B to PSC

Contacts

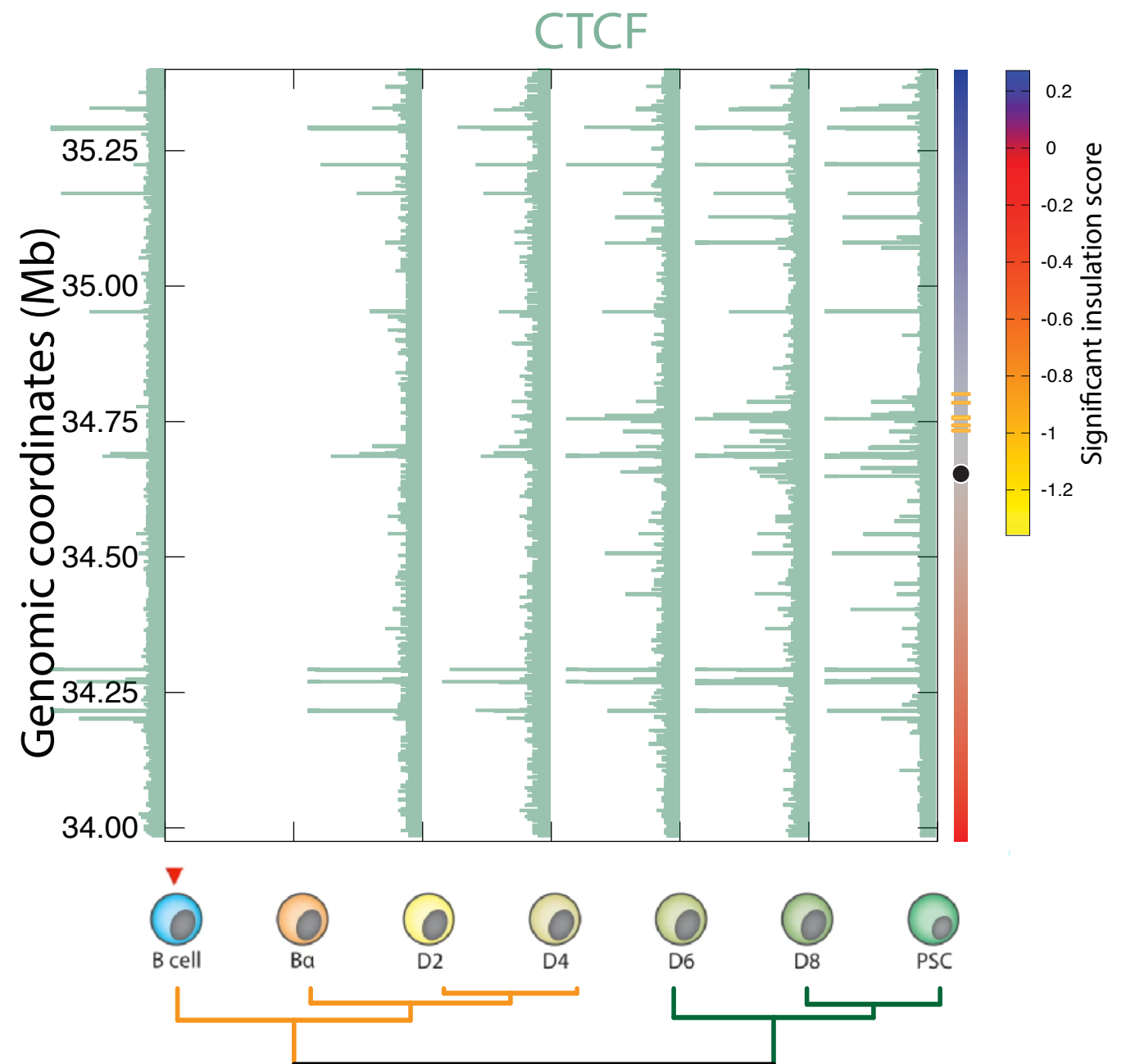
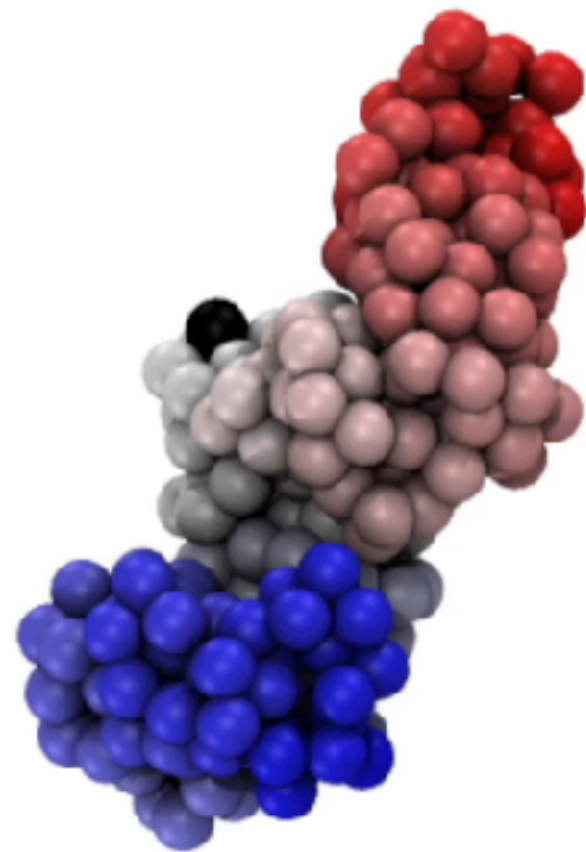


SE
SOX2



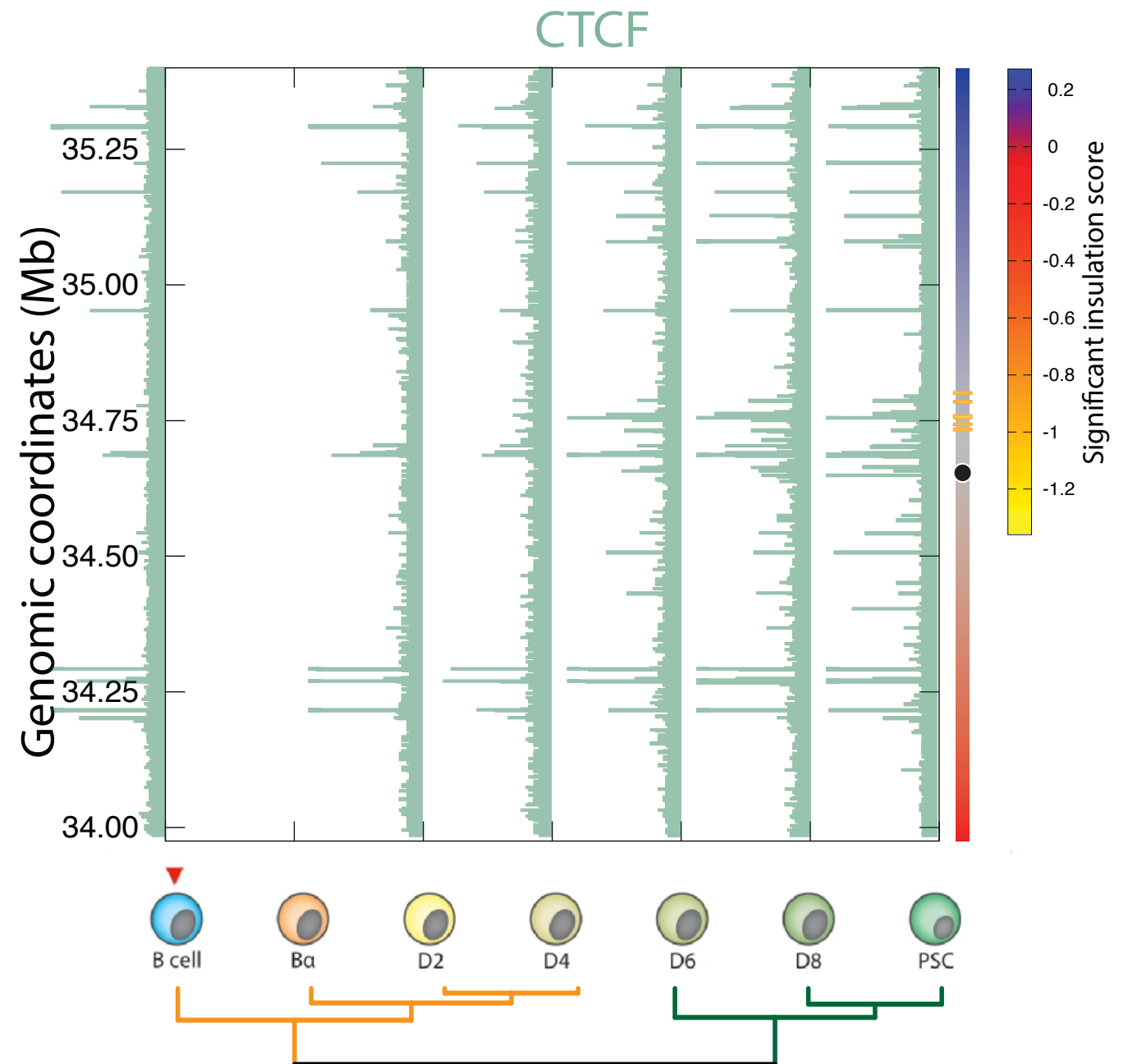
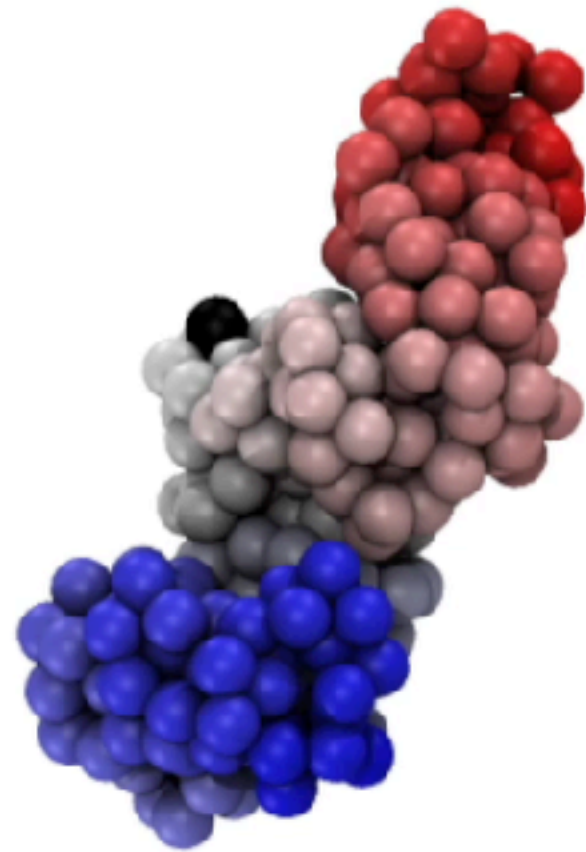
SOX2 locus structural changes from B to PSC

TAD borders



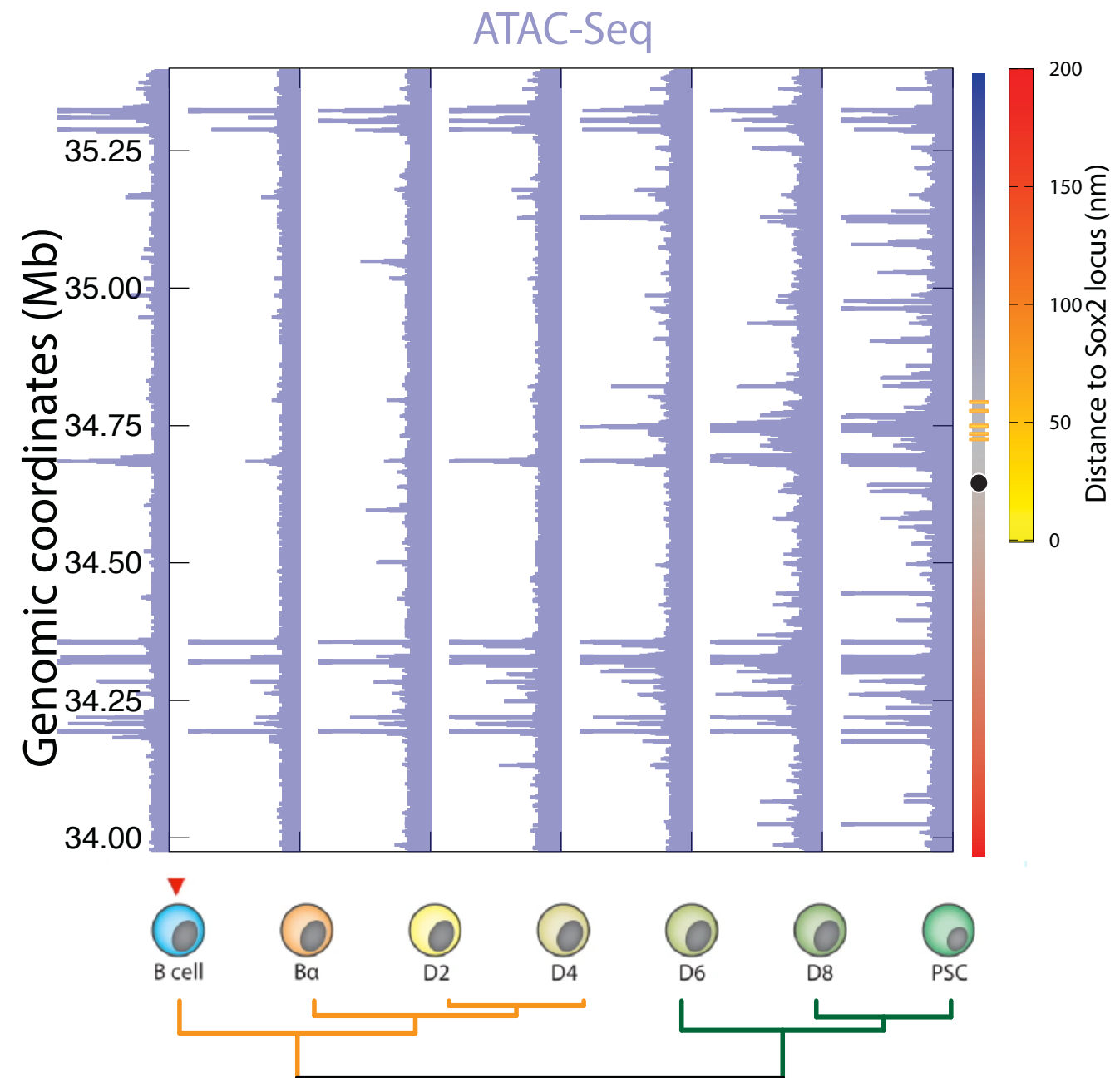
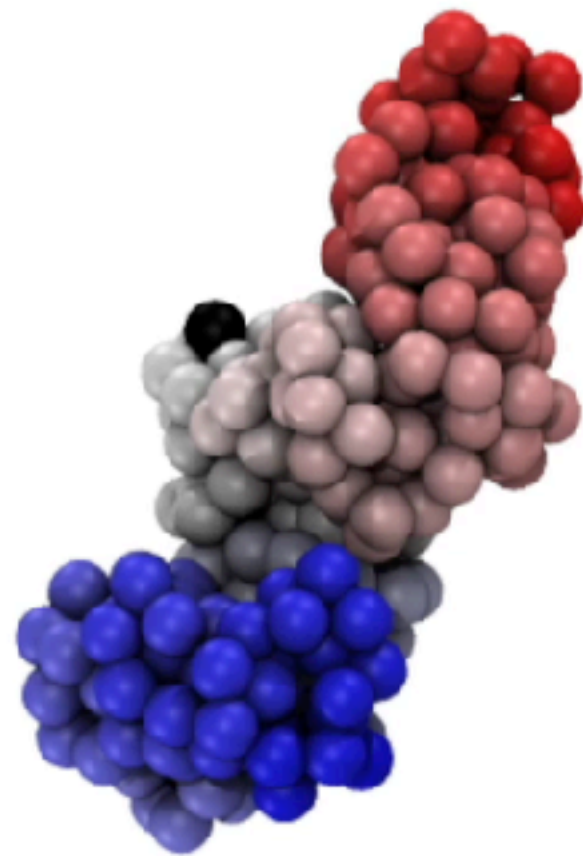
SOX2 locus structural changes from B to PSC

TAD borders



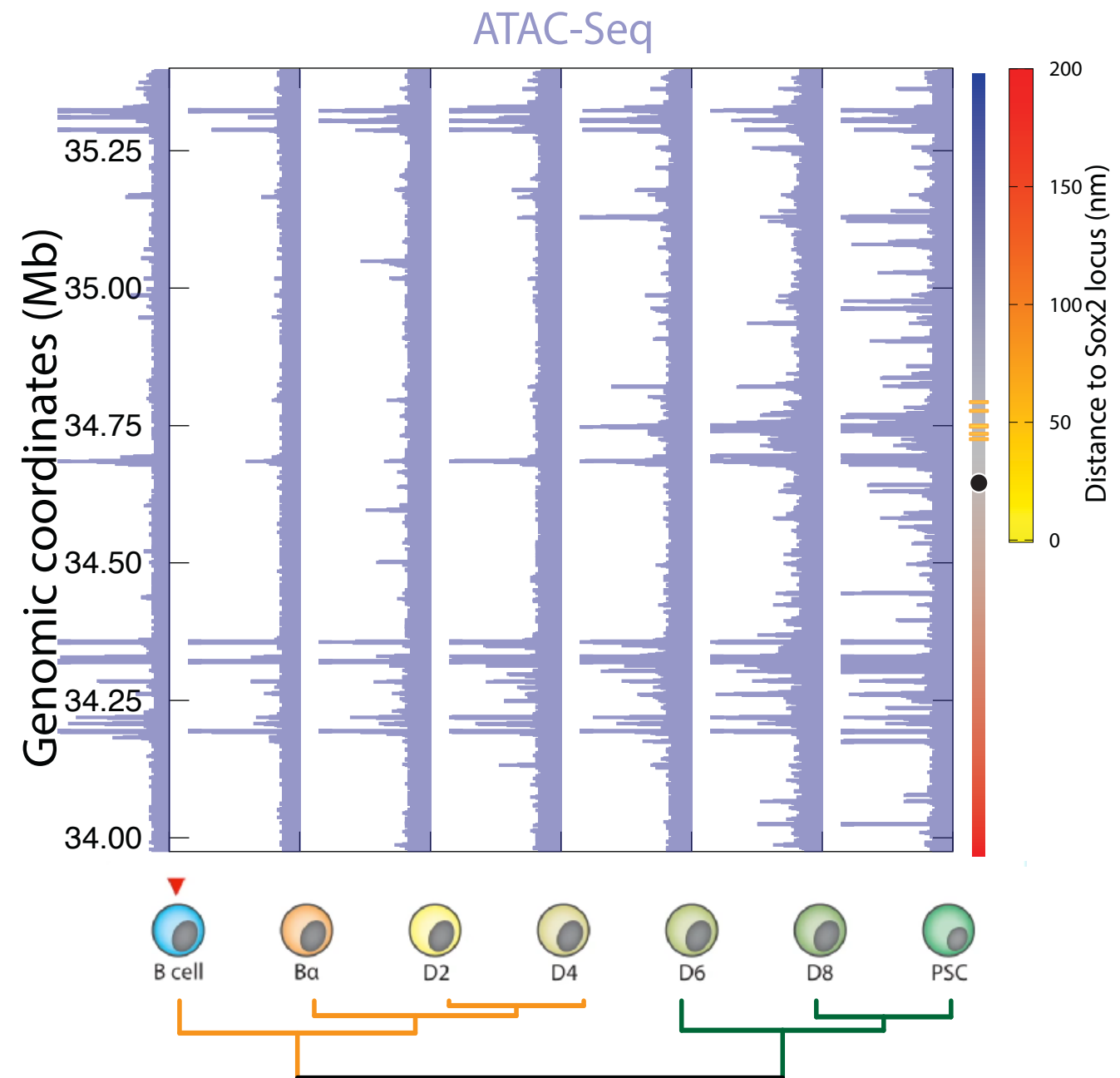
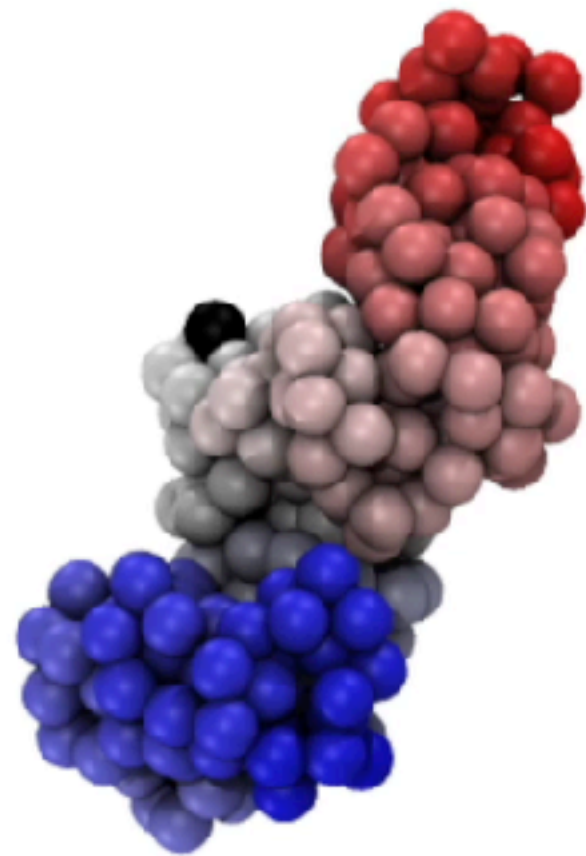
SOX2 locus structural changes from B to PSC

Distance to regulatory elements



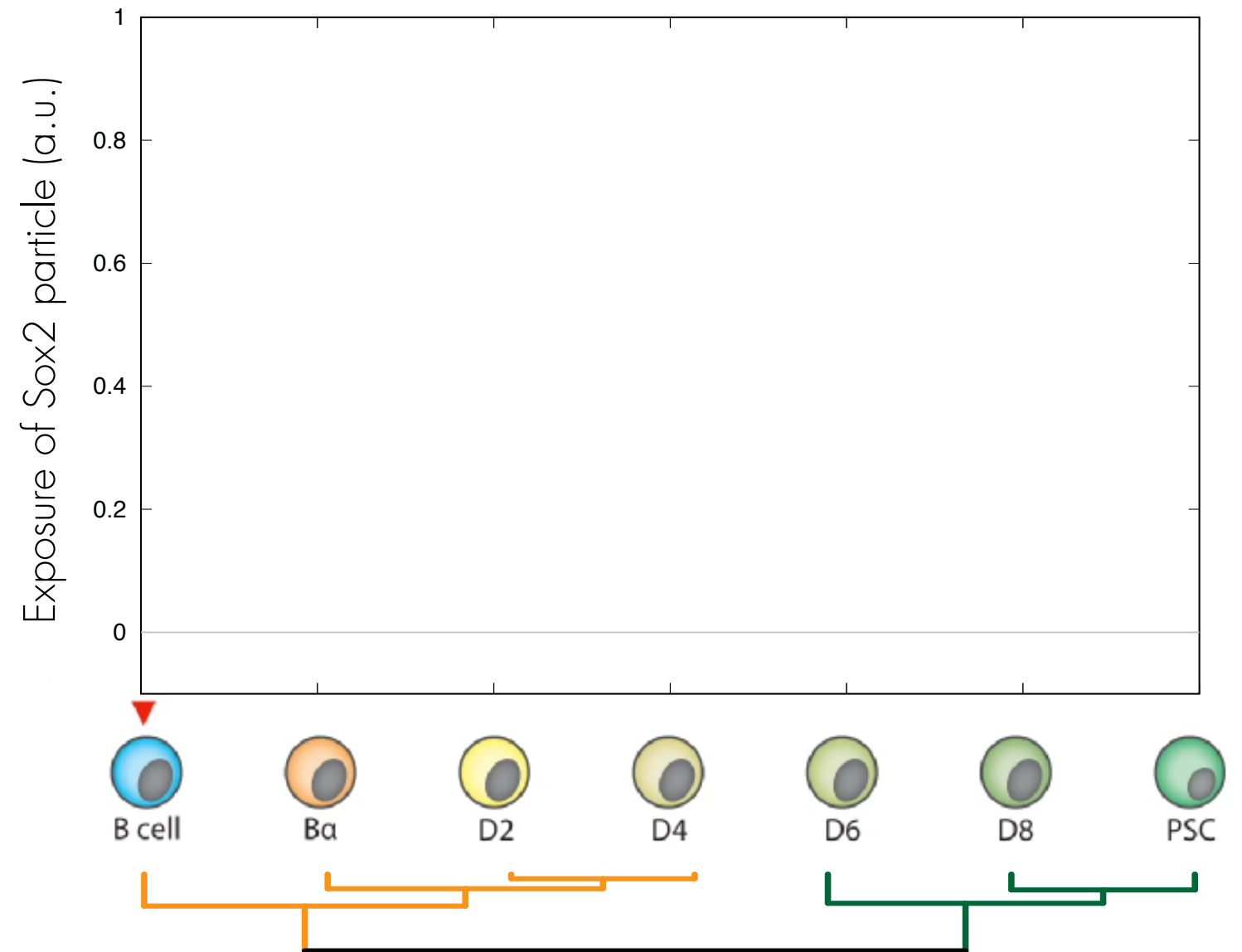
SOX2 locus structural changes from B to PSC

Distance to regulatory elements



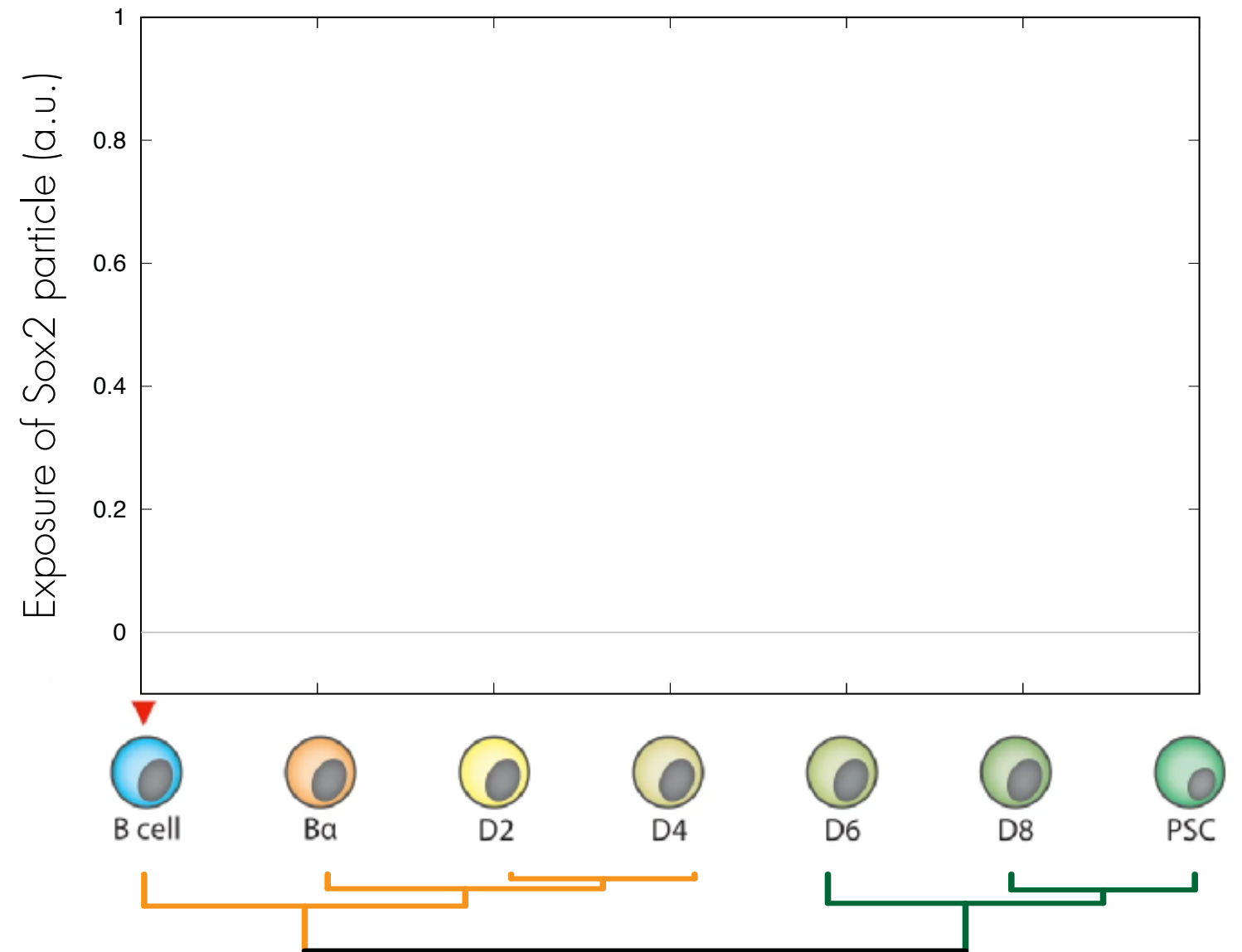
SOX2 locus structural changes from B to PSC

Structural exposure



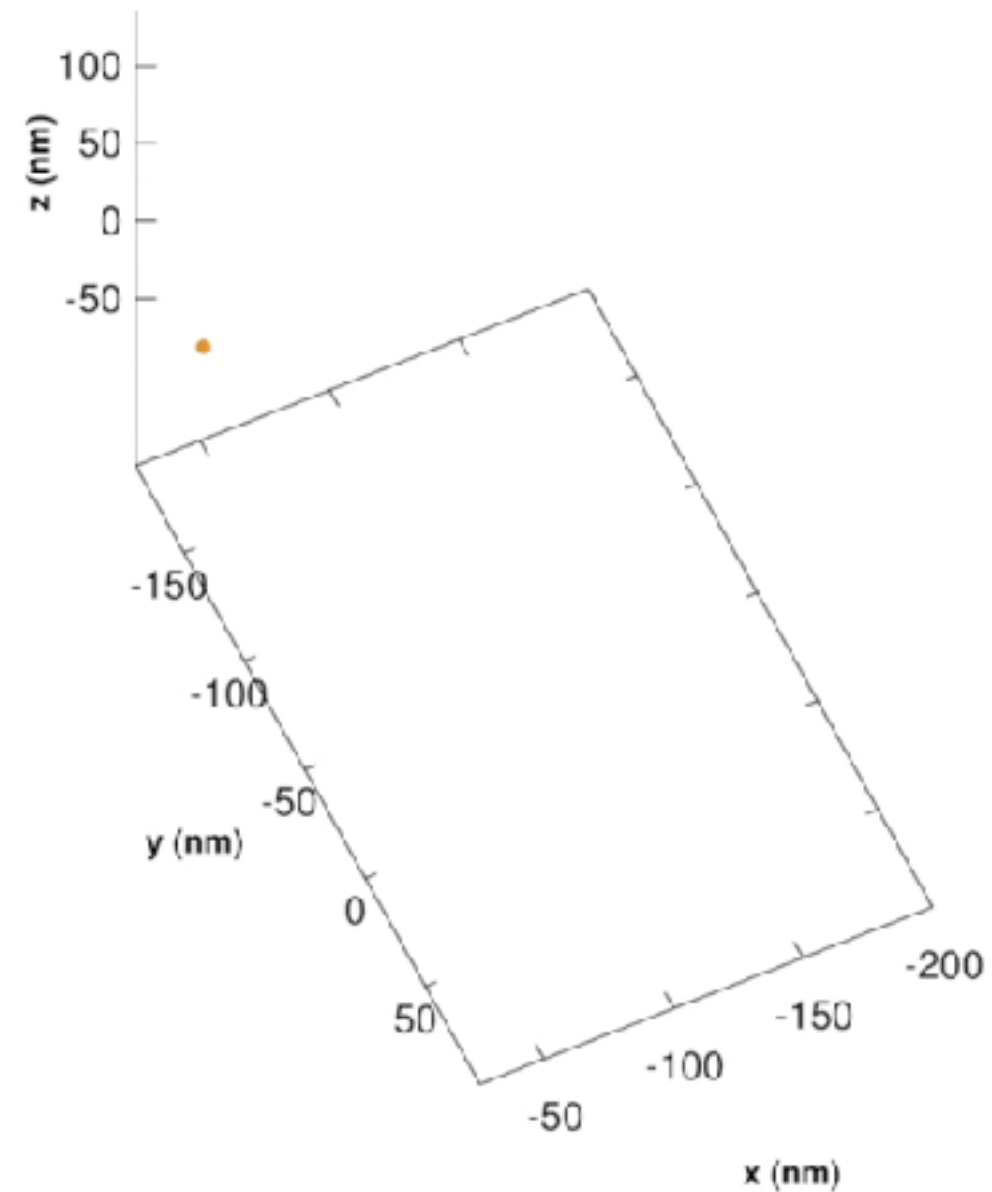
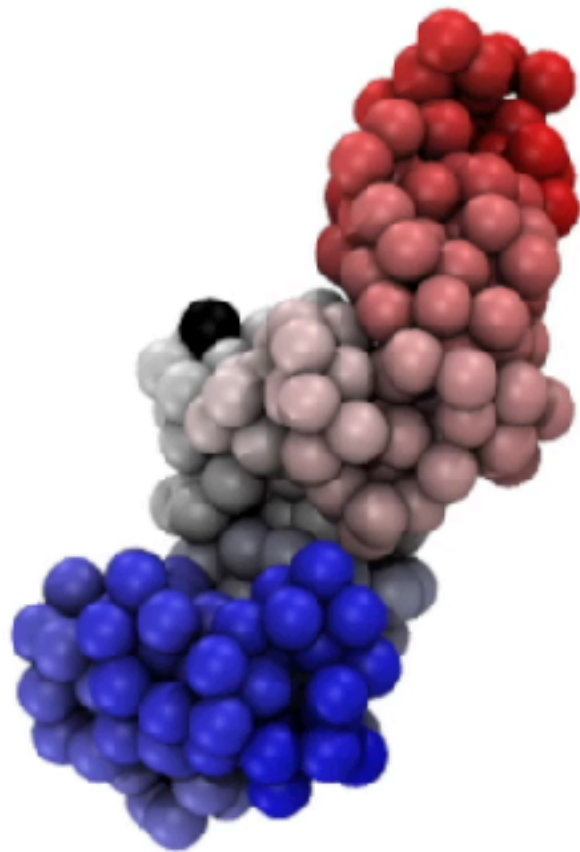
SOX2 locus structural changes from B to PSC

Structural exposure



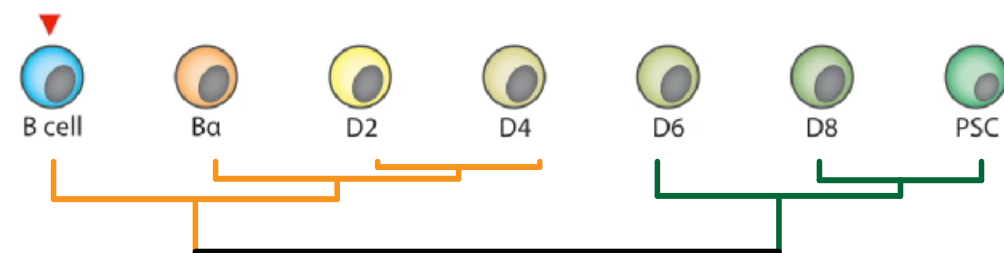
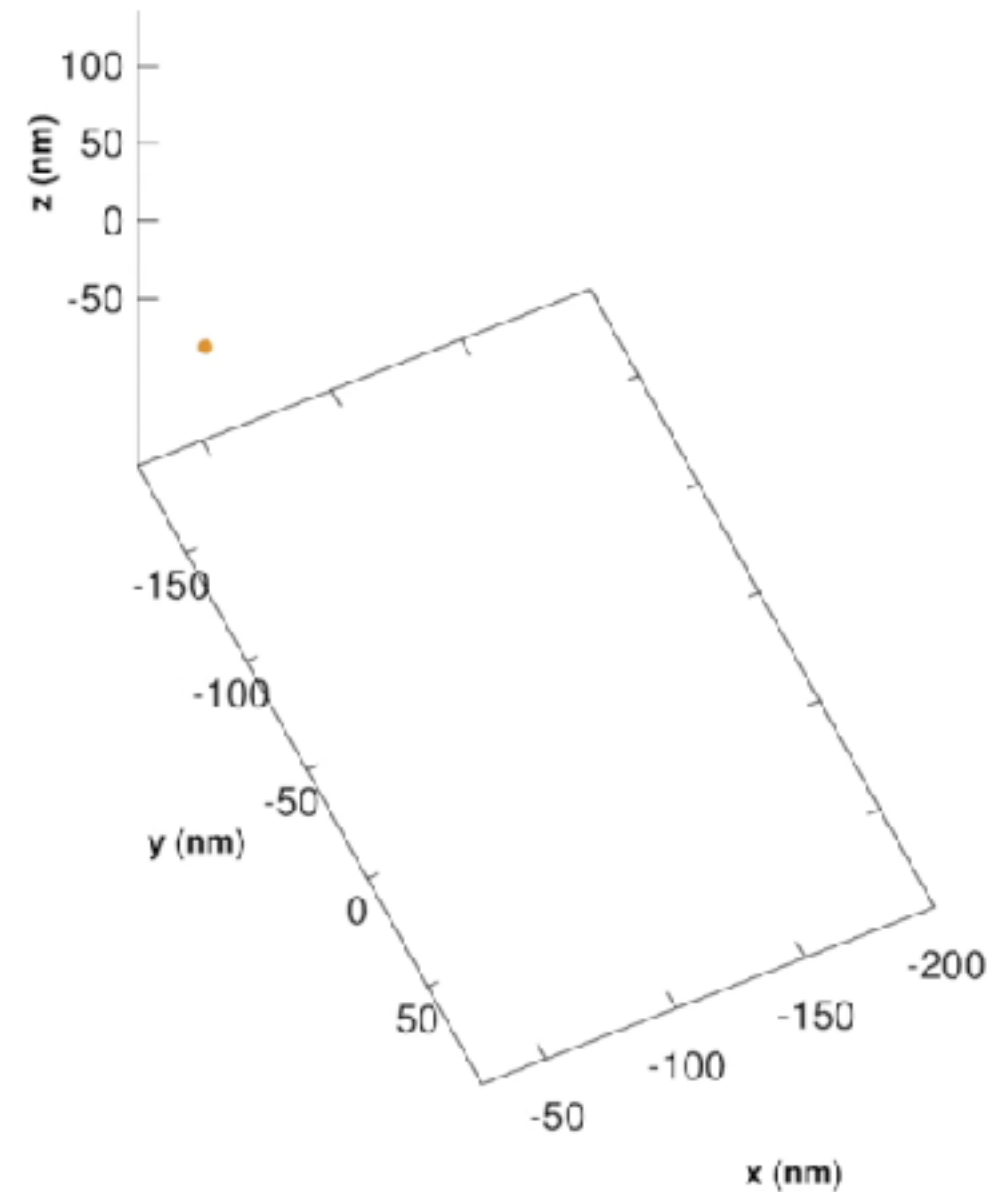
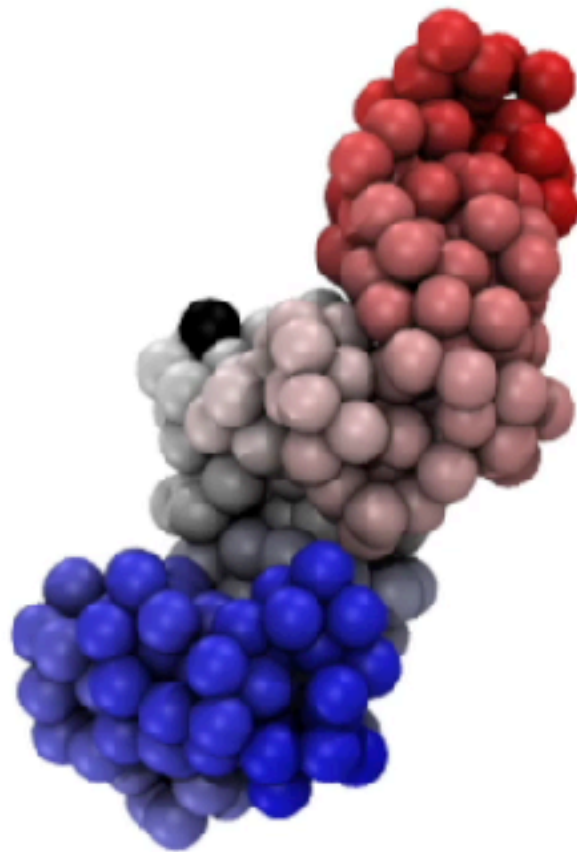
SOX2 locus dynamics changes from B to PSC

SOX2 displacement



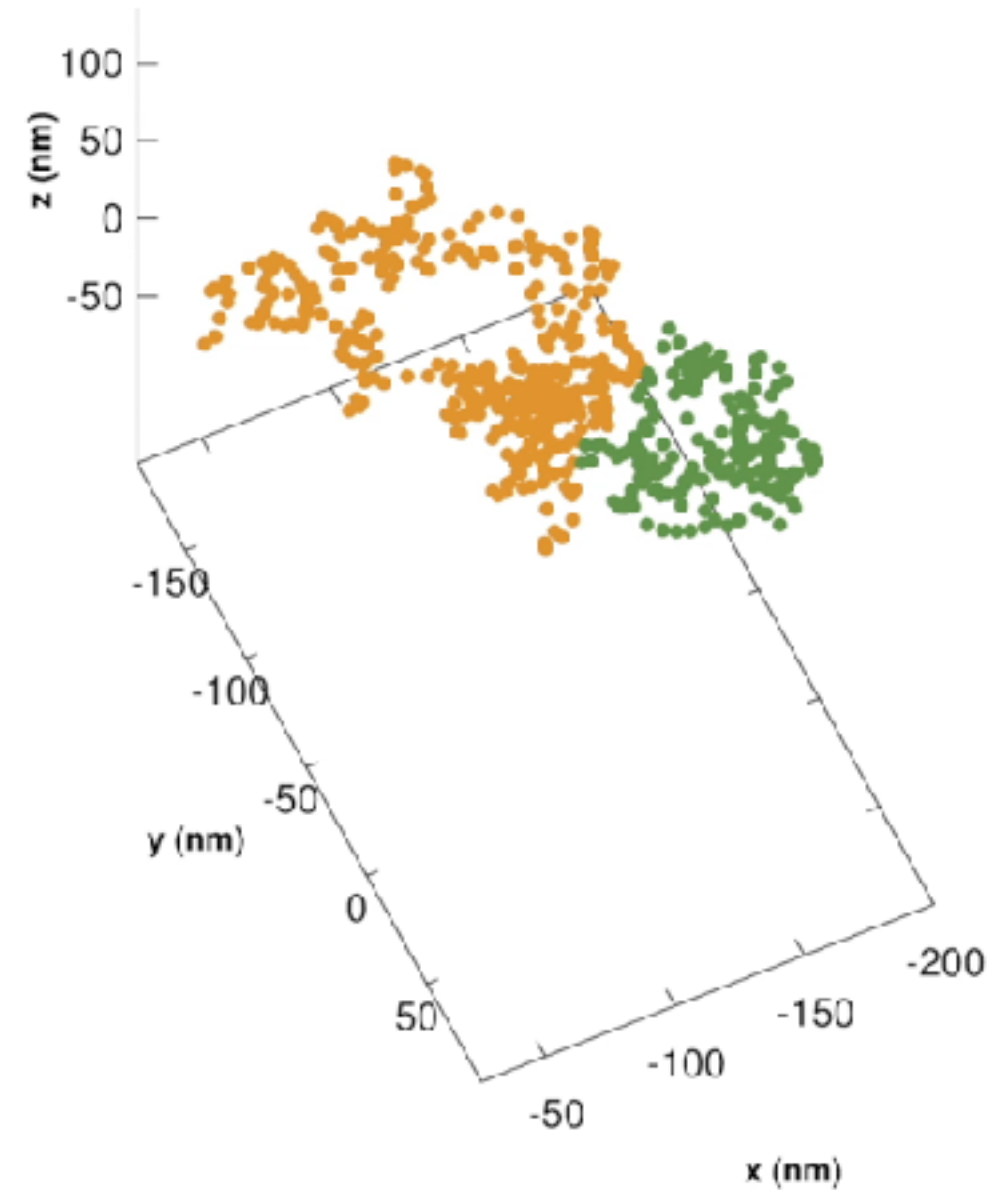
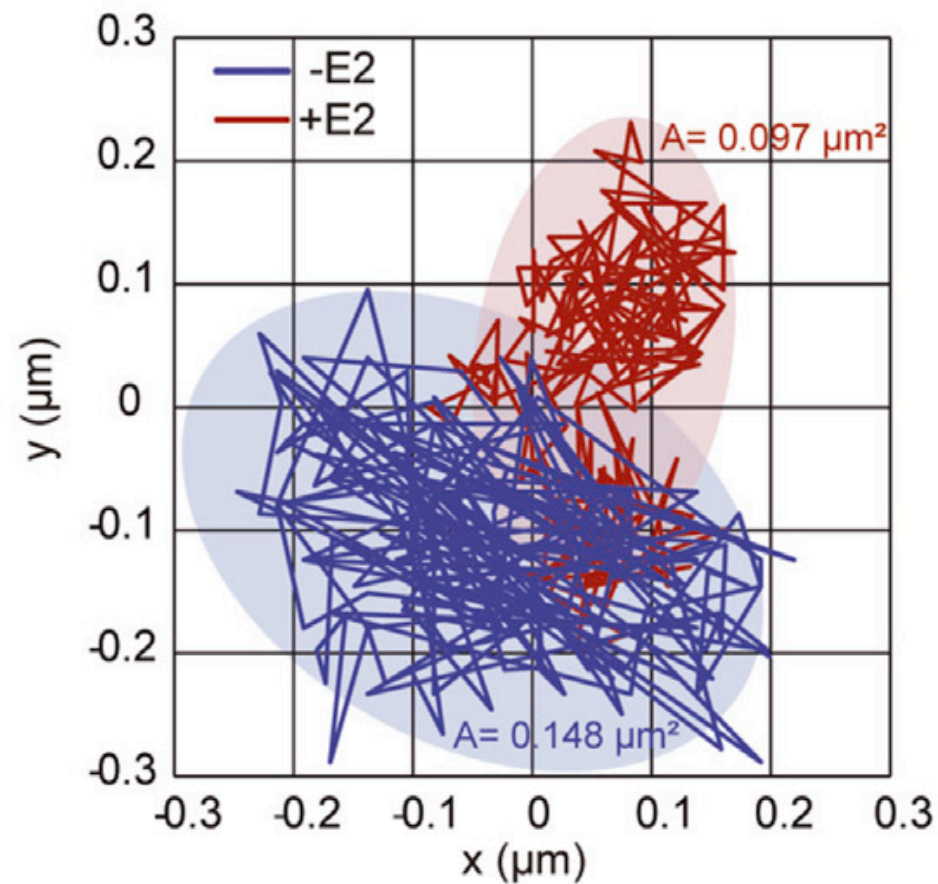
SOX2 locus dynamics changes from B to PSC

SOX2 displacement



SOX2 locus dynamics changes from B to PSC

SOX2 displacement

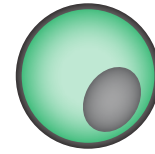
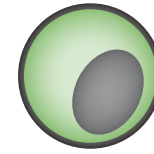
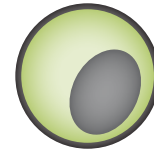
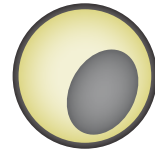
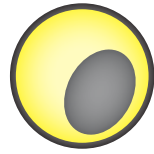
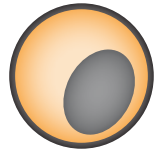
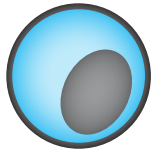


Two dimensional trajectories and area explored over 50s of the CCND1 locus recorded before -E2 and after +E2 activation.

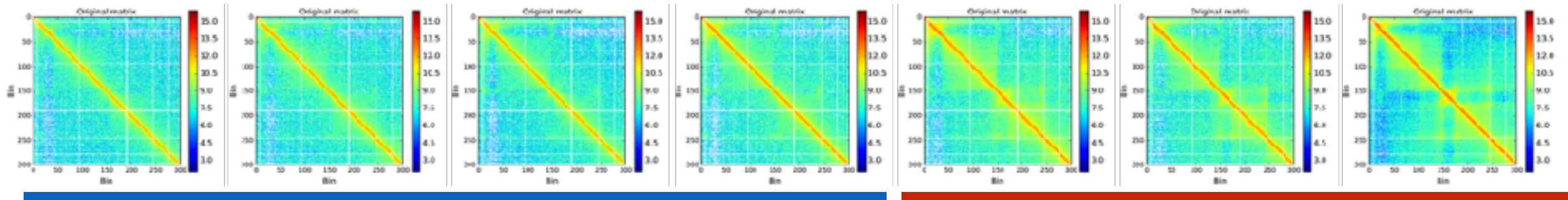
Germier, T., et al, Biophys J. 113, 1383–1394 (2017).



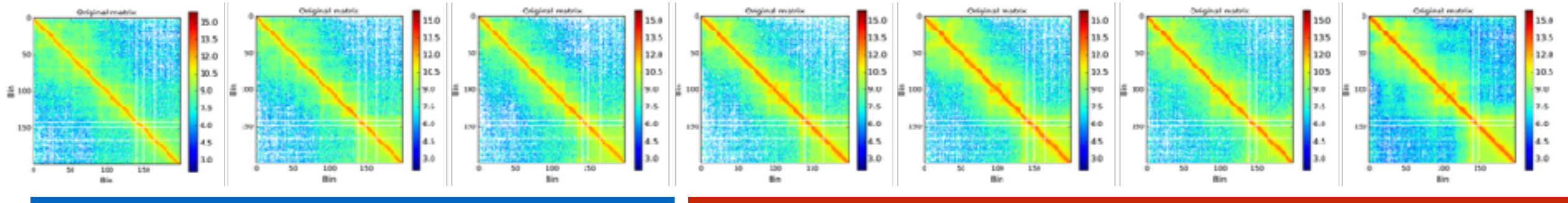
Other regions...



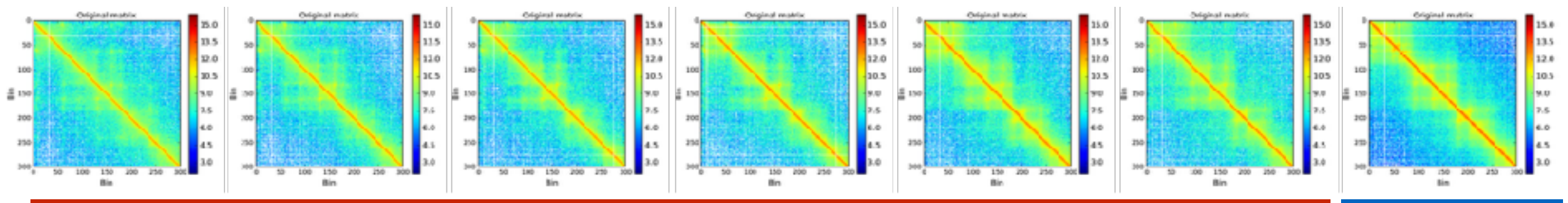
Sox2 chr3:34649995-34652460



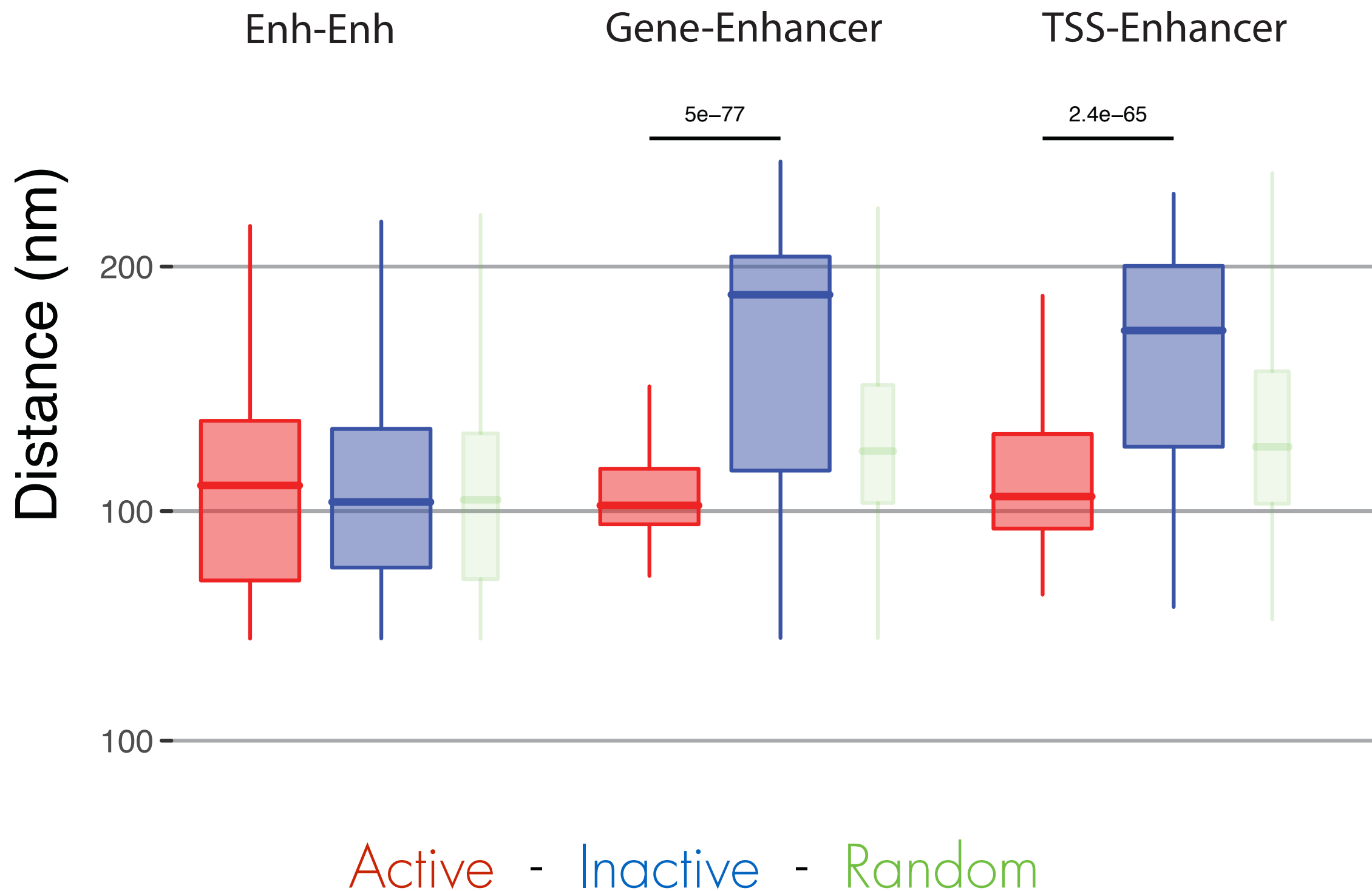
Nanog chr6:122707565-122714633



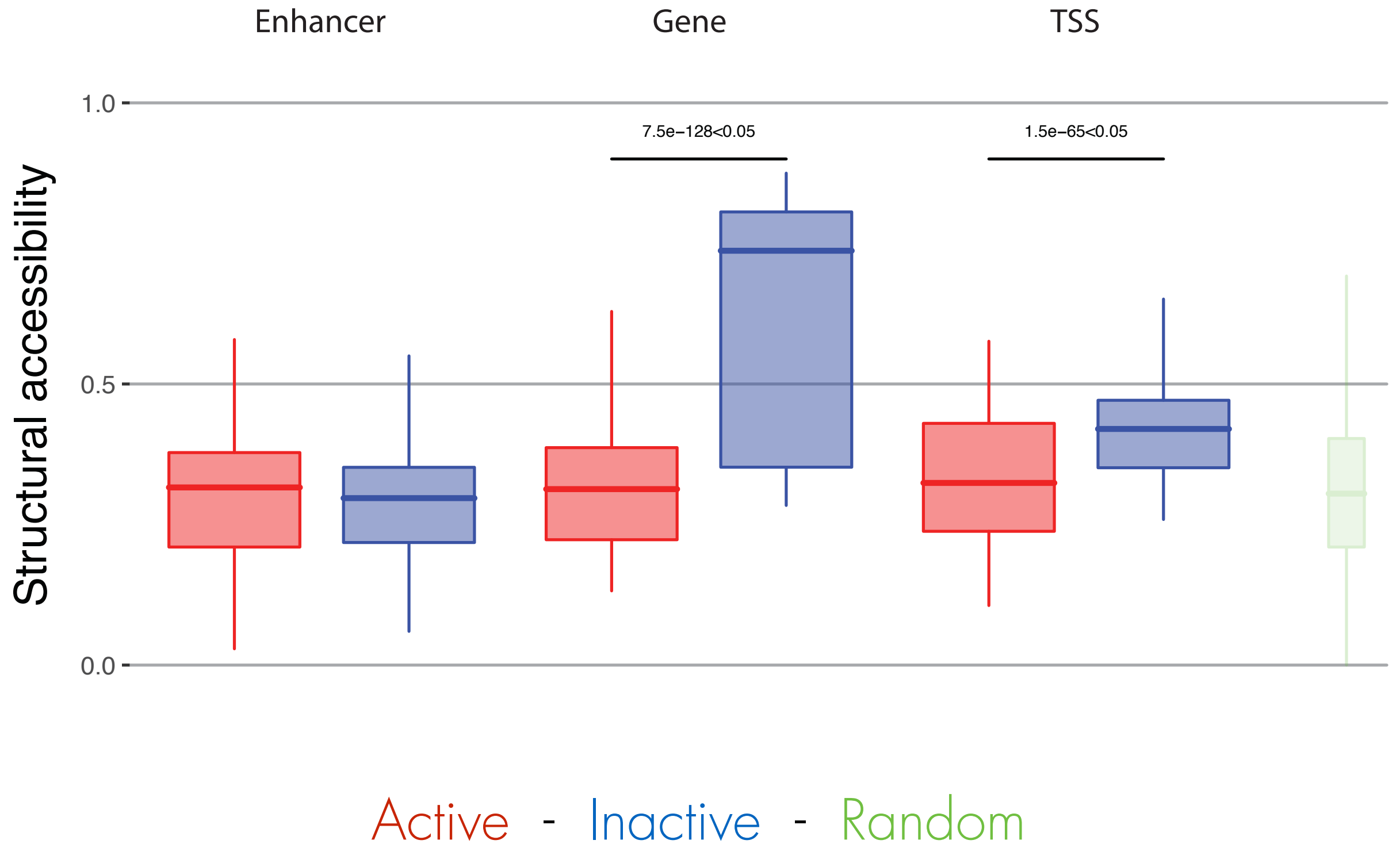
CEBPα chr7:35119293-35121931



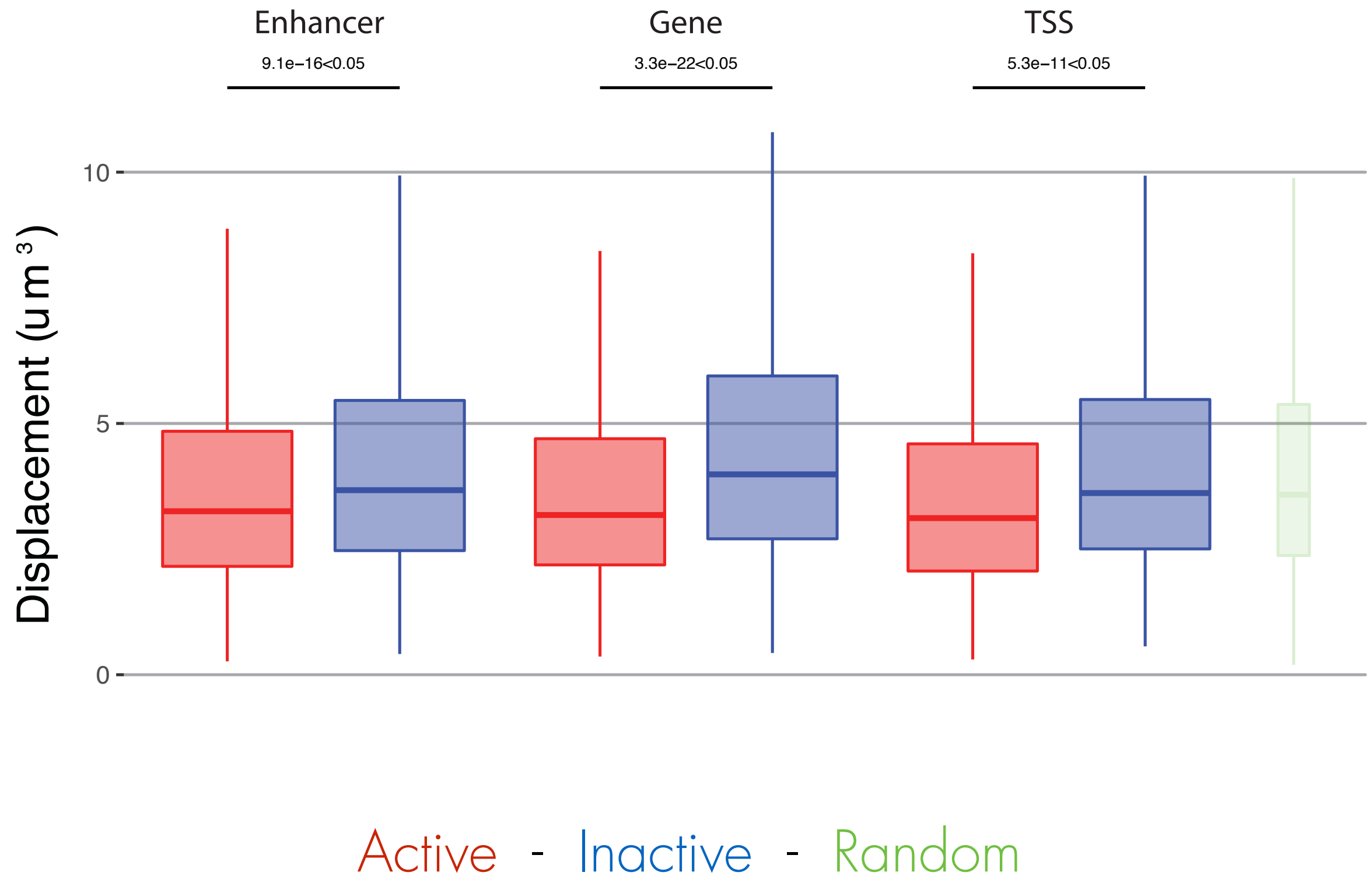
Distance to enhancers



Accessibility



Displacement



A “cage” model for transcriptional activation



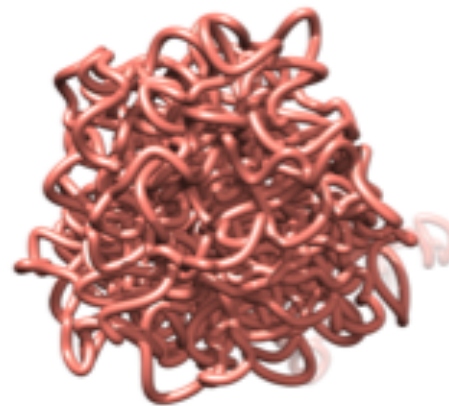
stpRNAs, a new type of structural RNAs?



Irene Farabella

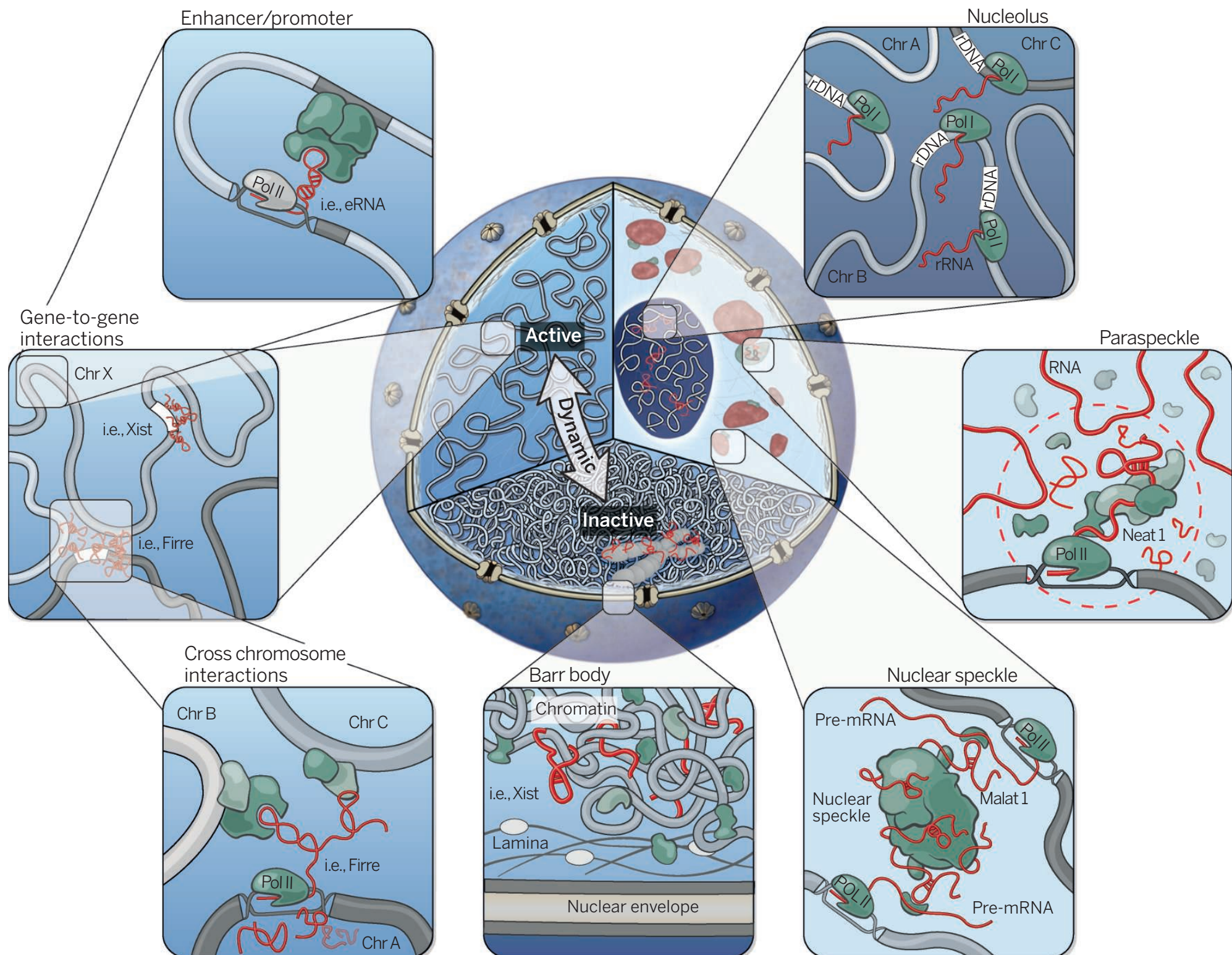


Marco di Stefano

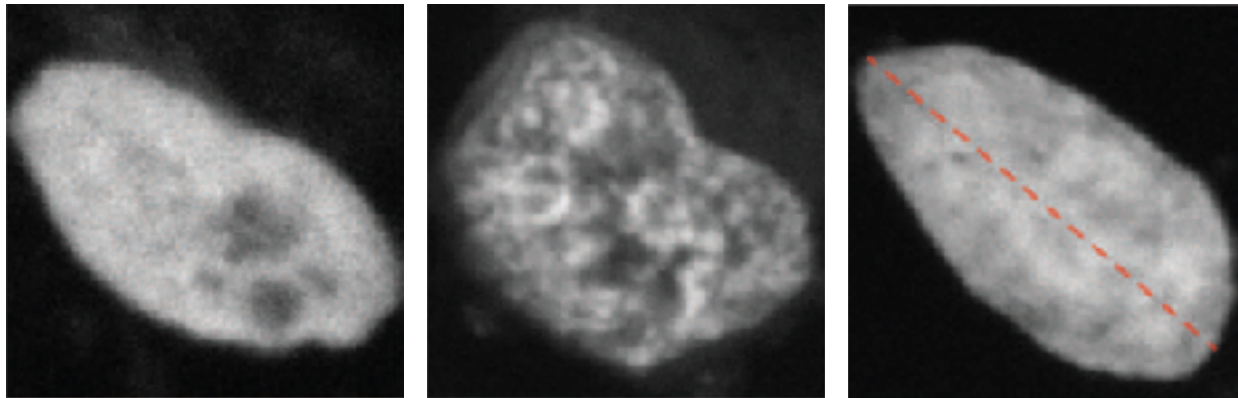


RNA, nuclear organisation dynamics and architecture

Rinn and Guttman, Science; 345(6202):1240–1241 (2014)

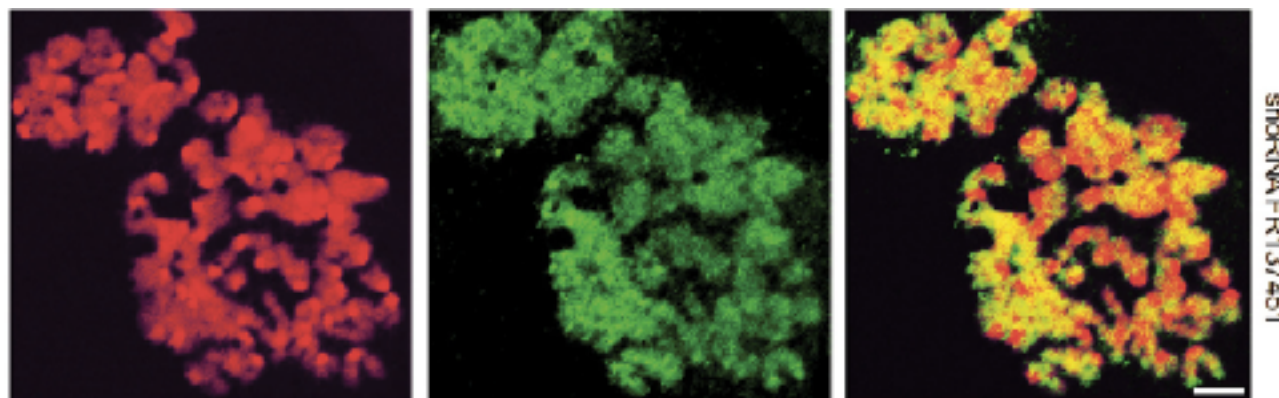


RNA, nuclear organisation dynamics and architecture



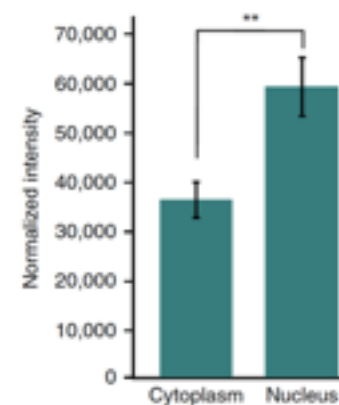
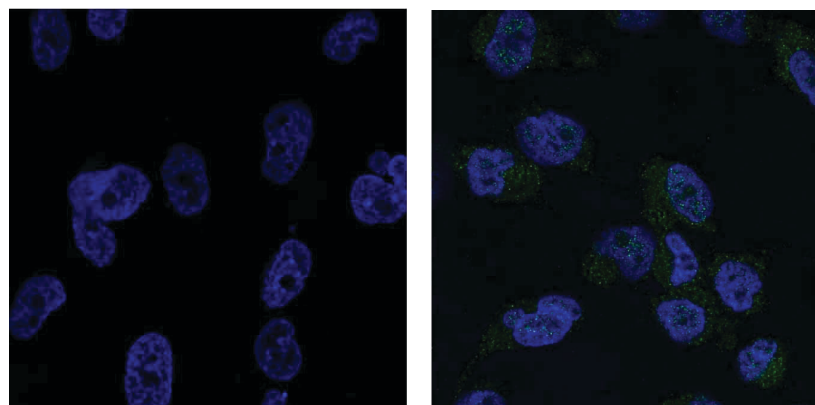
Chromatin-interlinking lncRNAs

Adapted from: Caudron-Herger et al., Nucleus ;2(5):410-24 (2011)



Mitotic chromosome-associated RNAs

Adapted from: Meng et al., Nucleic Acids Res. ;44(10):4934-46 (2016)



RNA-DNA triplex *in vivo* and *in vitro*

Adapted from: Mondal et al. Nat Commun. 6:7743 (2015)

Are there lncRNAs that act as global architectural factor for chromatin organisation?

Hypothesis: such lncRNA may interact with DNA through triplex formation.

lncRNA selection

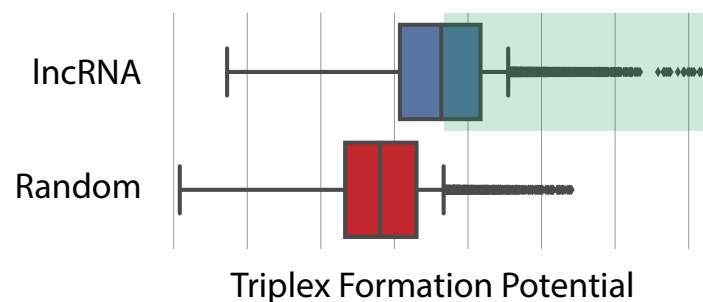


GENCODE v19 mapped of GRCh37
lncRNA genes 13,870



K562 & GM12878
Nuclear localisation
 ≥ 1 RPKM in both replica and in both cells line

≥ 2 exons
>200 nt length
nuclear in K562 & GM12878



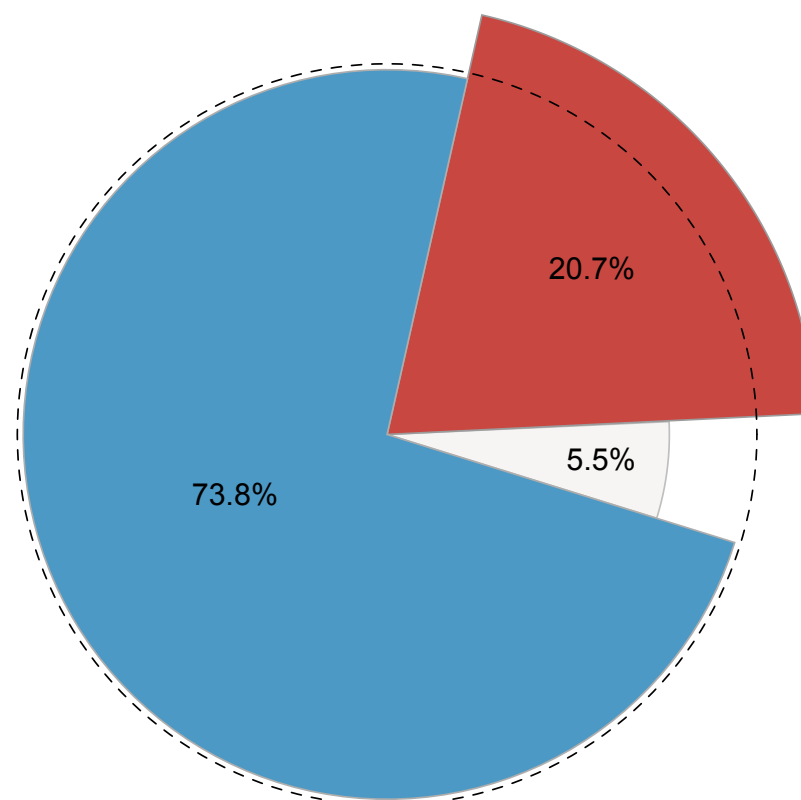
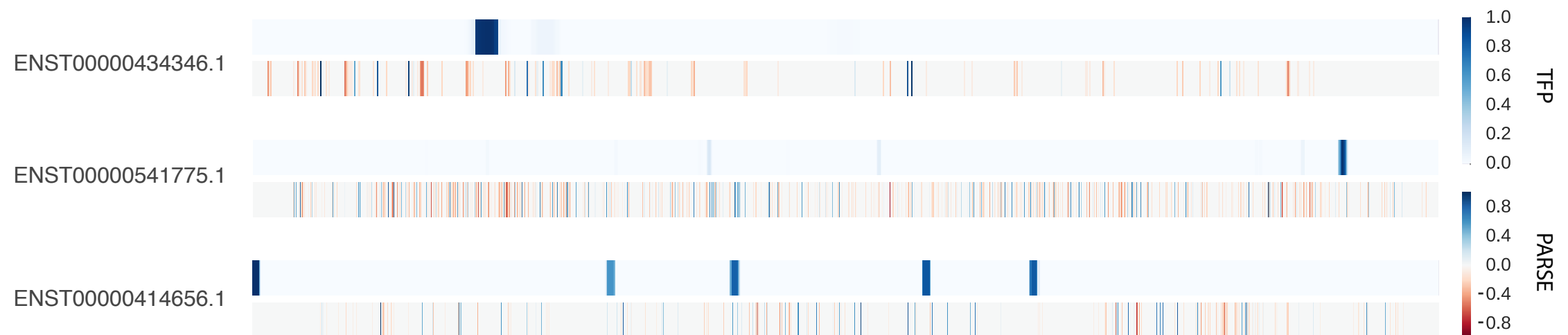
Triplex forming lncRNA
enriched

set of 339 lncRNAs

RNA-Seq expression using
GEM split mapper & Flux capacitor
Marc Dabad & Anna Esteve (CNAG-CRG)

TFO/PARSE IncRNA profiles

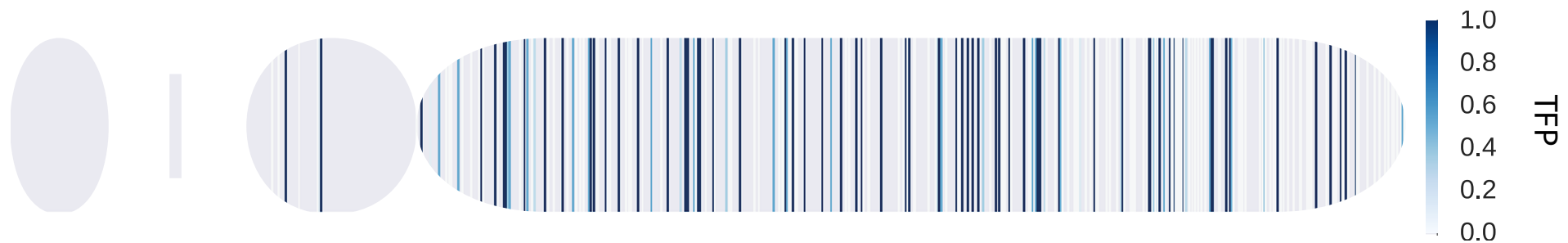
127 triplex forming IncRNA with Secondary Structure information based on PARSE



Data from: Wan et al., Nature 505, 706–709 (2014)

TTS profile on Chromosome

ENST00000505973 in chromosome 22
TTS profile

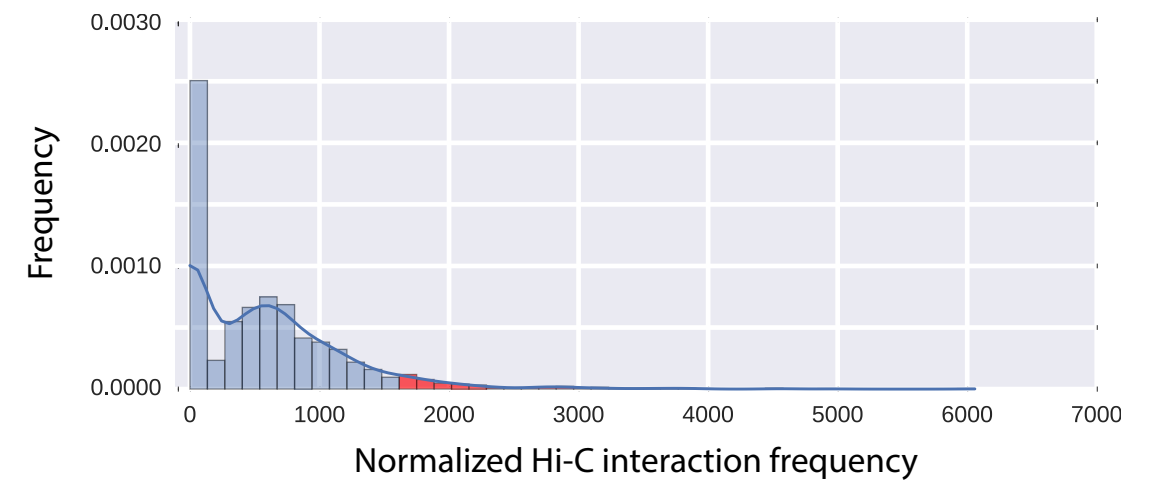
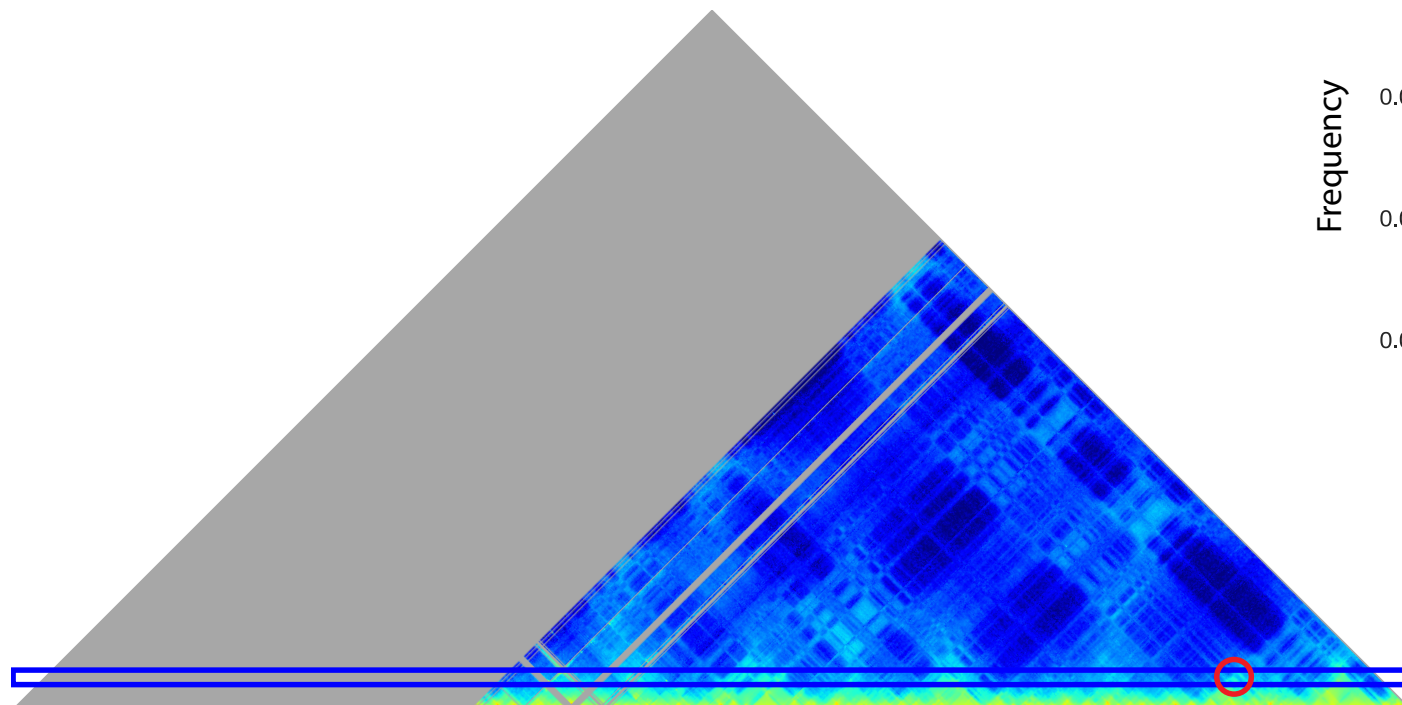


ENST00000434346.1 in chromosome 22
TTS profile



3D Co-localisation of loci

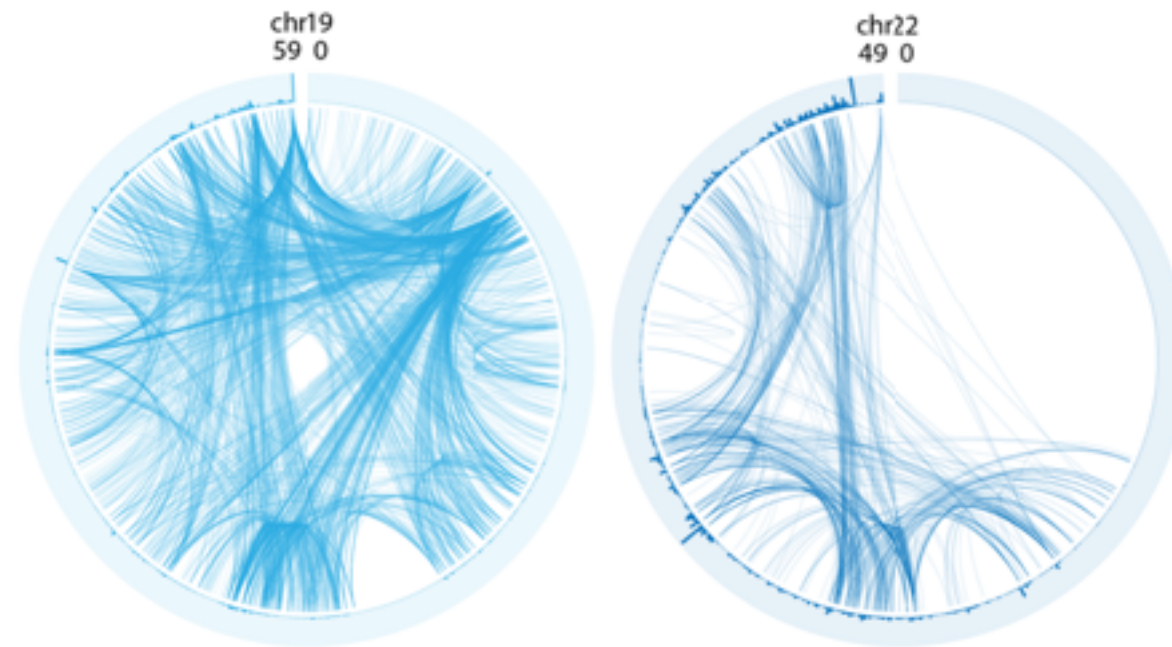
ENST00000505973 in chromosome 22
TTS profile



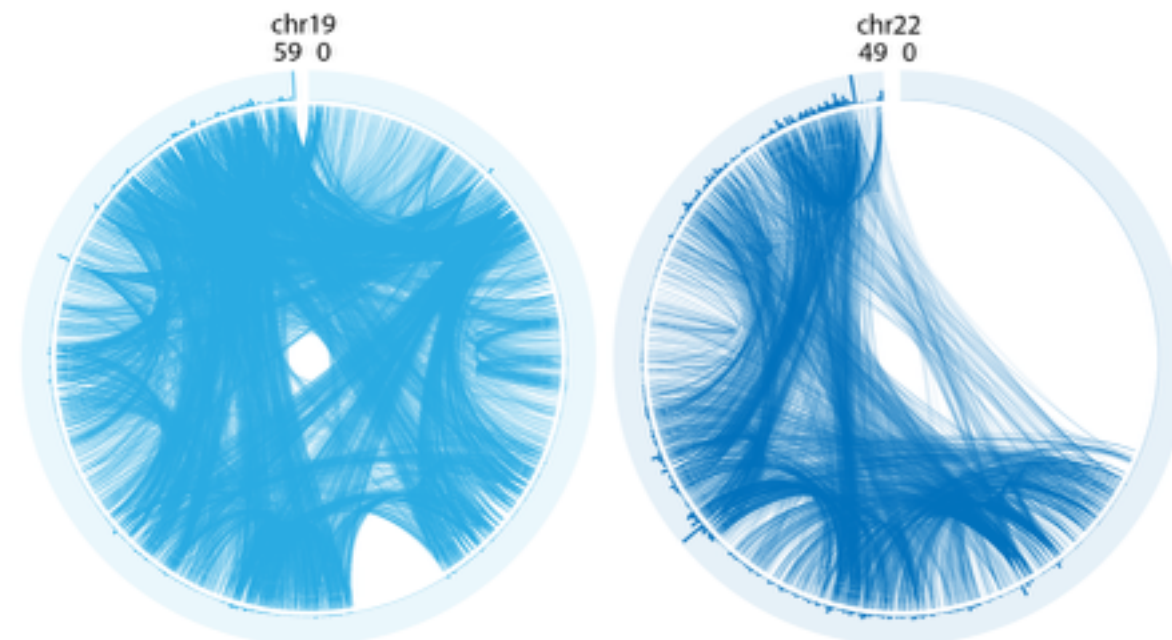
$$S = \log_2 \left(\frac{CTTS_d^{obs}}{CTTS_d^{exp}} \right)$$

Enriched Co-localised TTS site

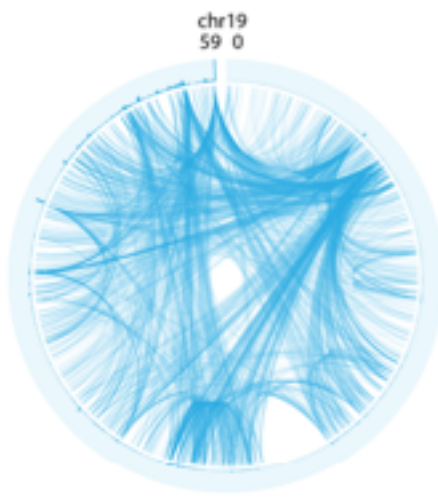
ENST00000434346.1



ENST00000541775.1



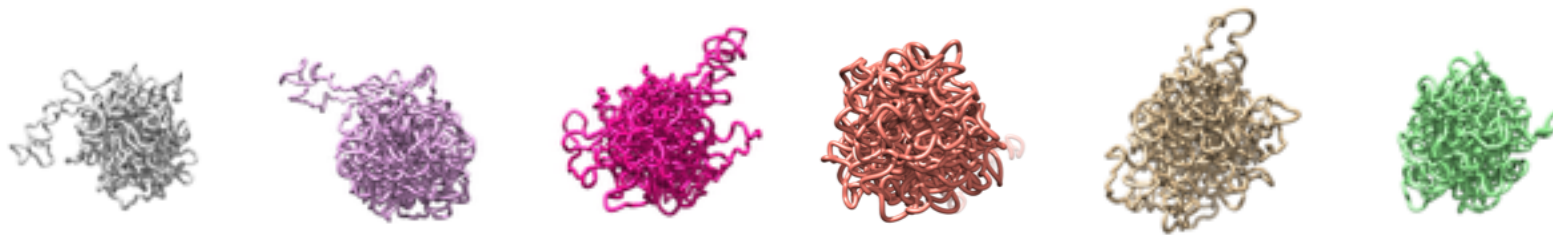
TADdyn modeling



Steered Molecular Dynamics
PLUMED/LAMMPS
with distance dependent harmonic restraints

Kremer and Grest 1991

Structure Population

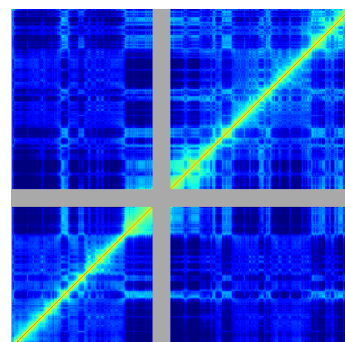


1,000 replicas using 10% of the initial set of restraints

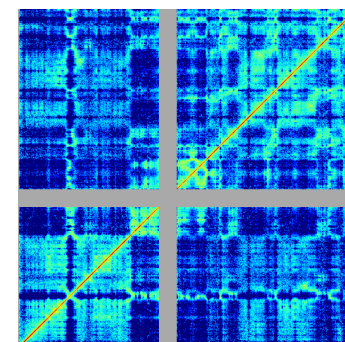


Marco Di Stefano

HiC experiment
(Rao @50kb)

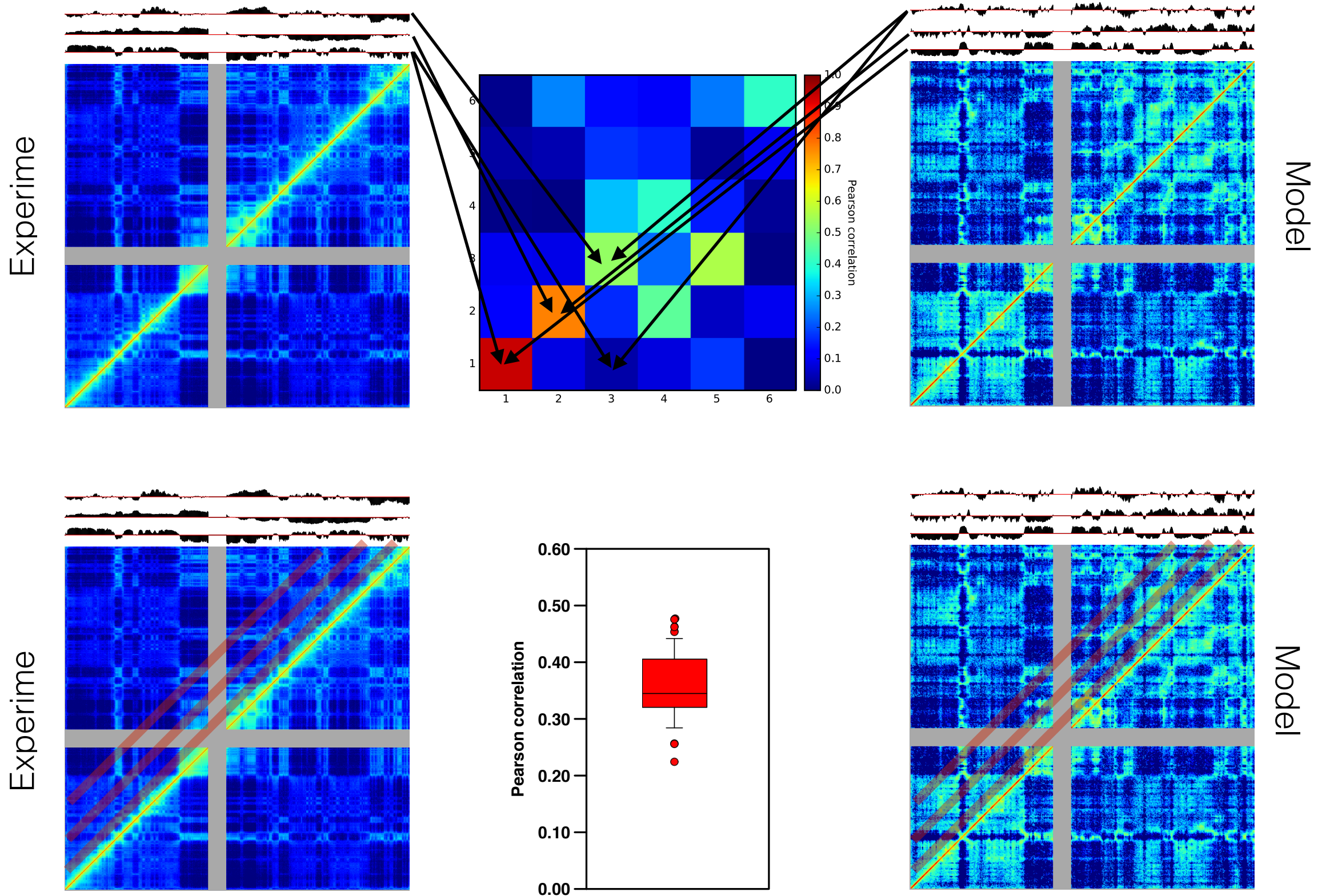


Predicted from
Structure population



Benchmark measures

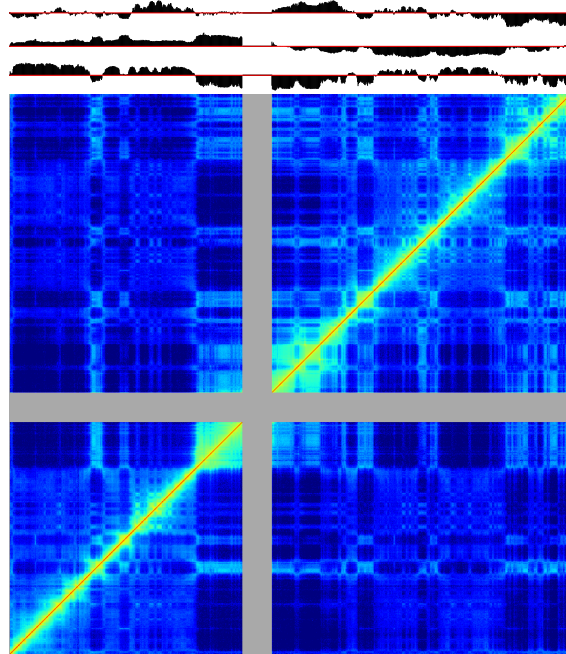
Eigen vector correlation & Diagonal cross correlation



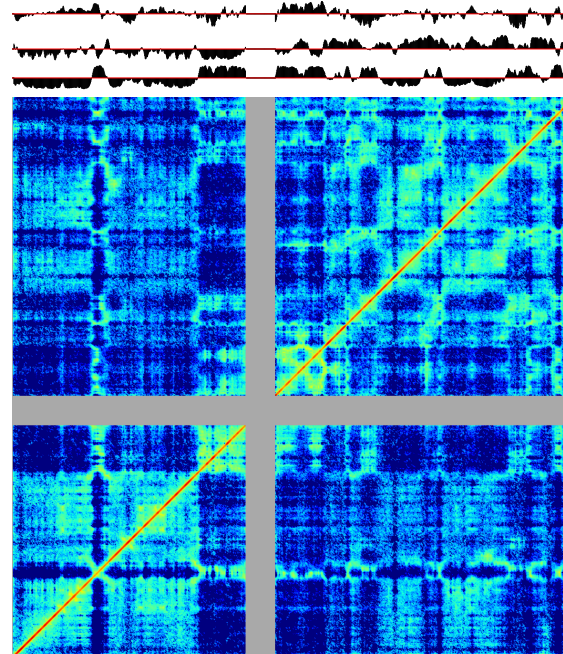
ENST00000434346.1 TTS in Chr19

total of 3,039 restraints over 620,899 possible (0.5%)

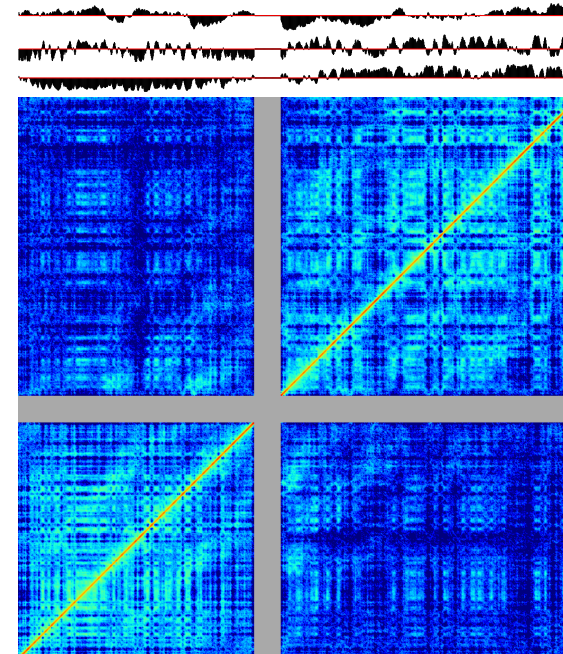
HiC experiment



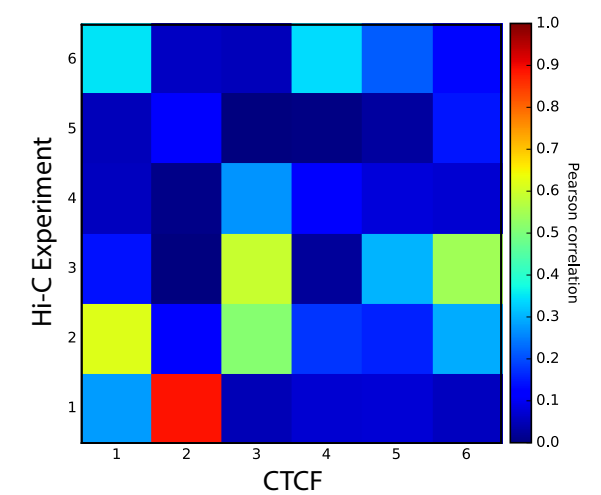
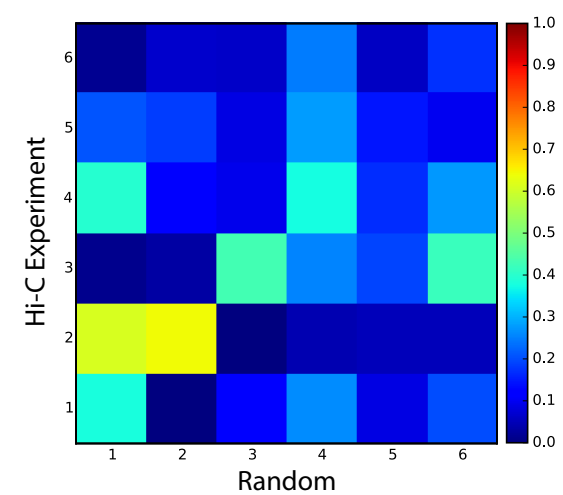
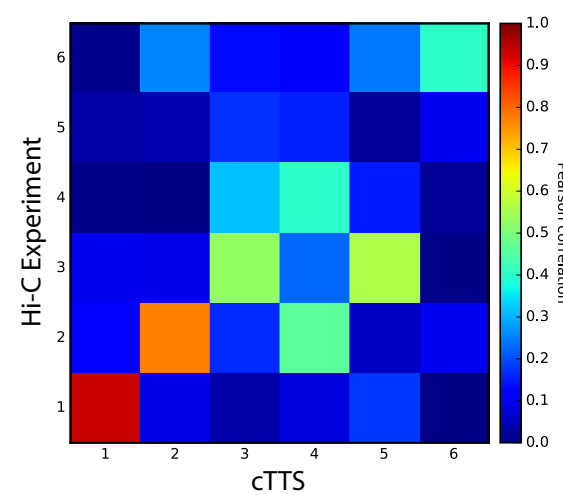
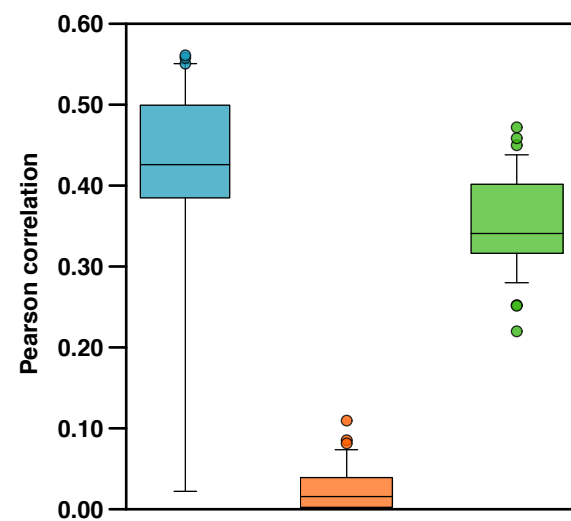
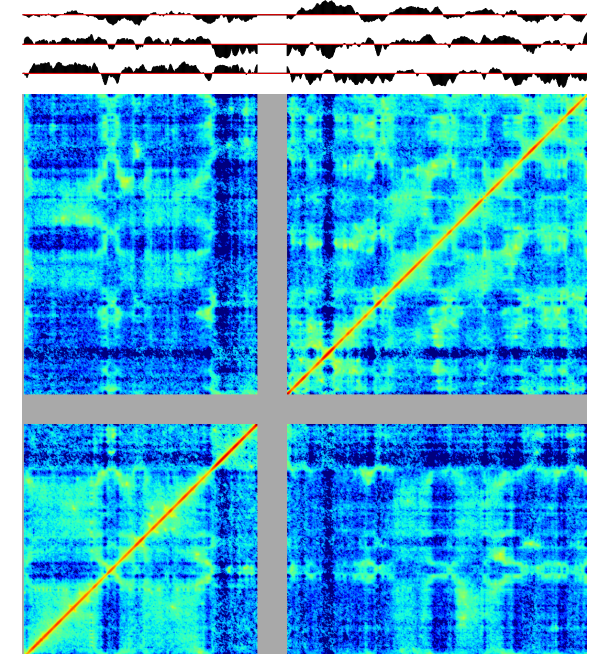
cTTS



Random

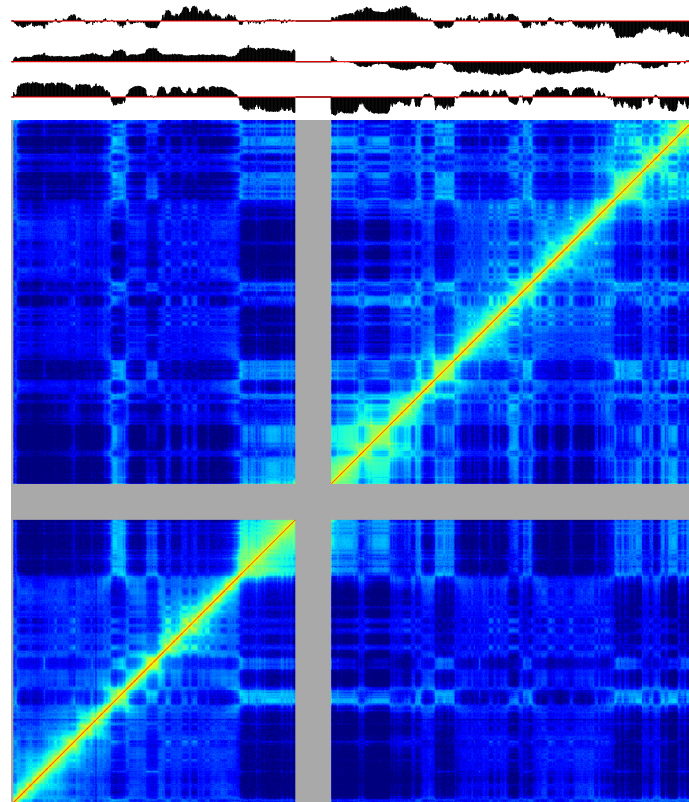


CTCF

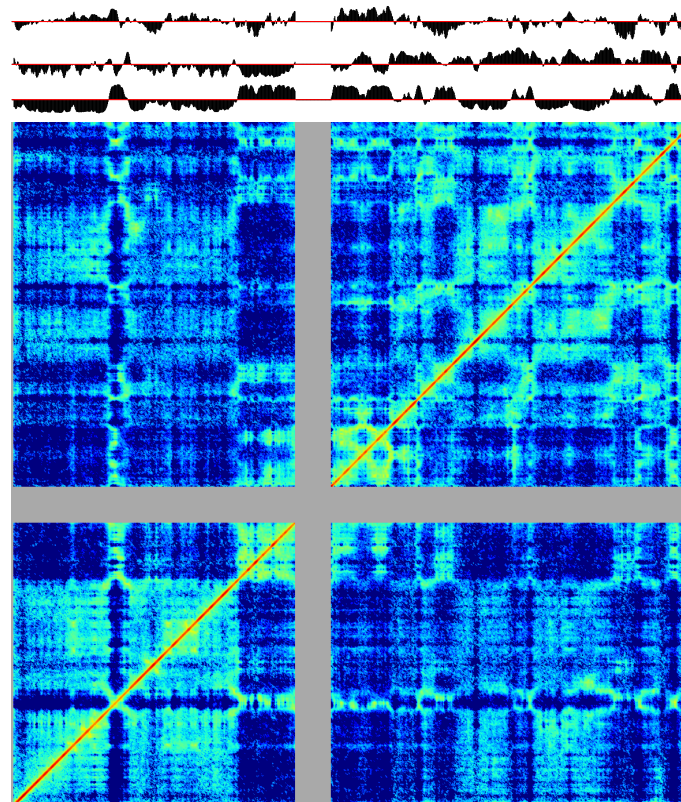


Other stpRNA TTS in Chr19

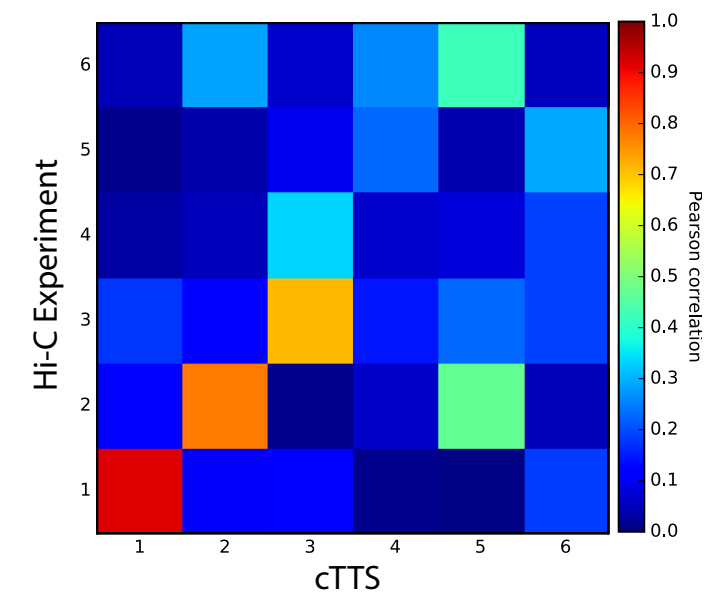
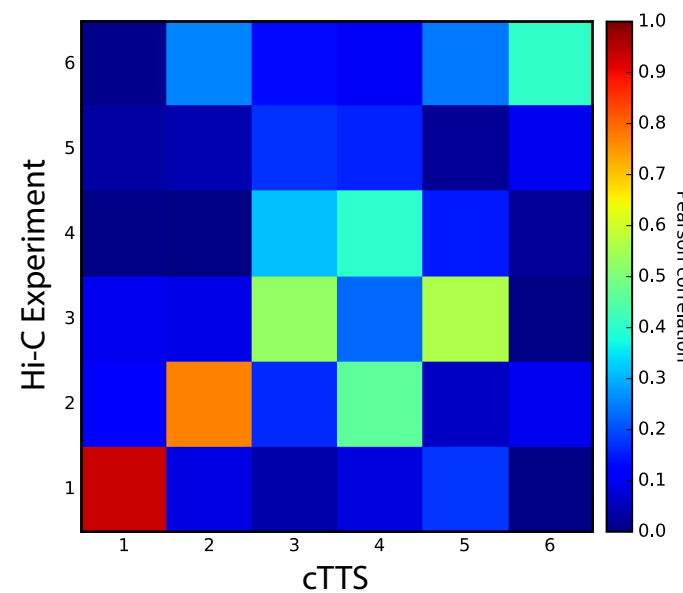
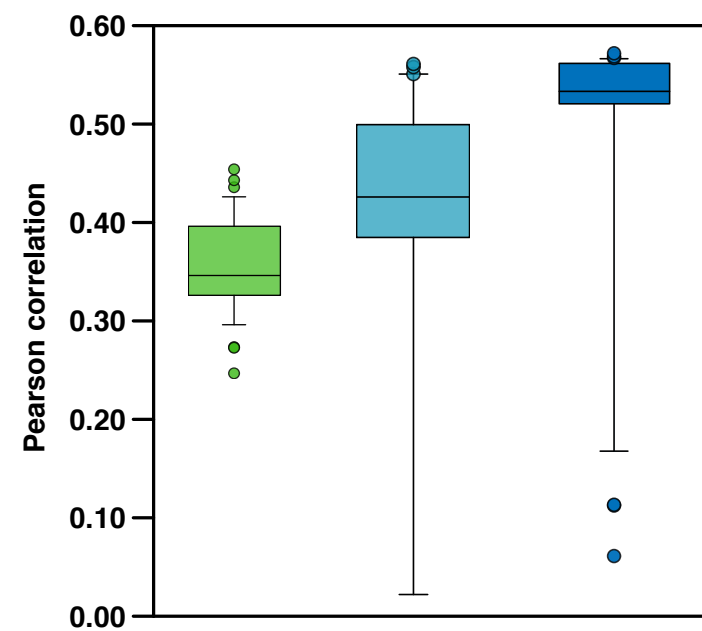
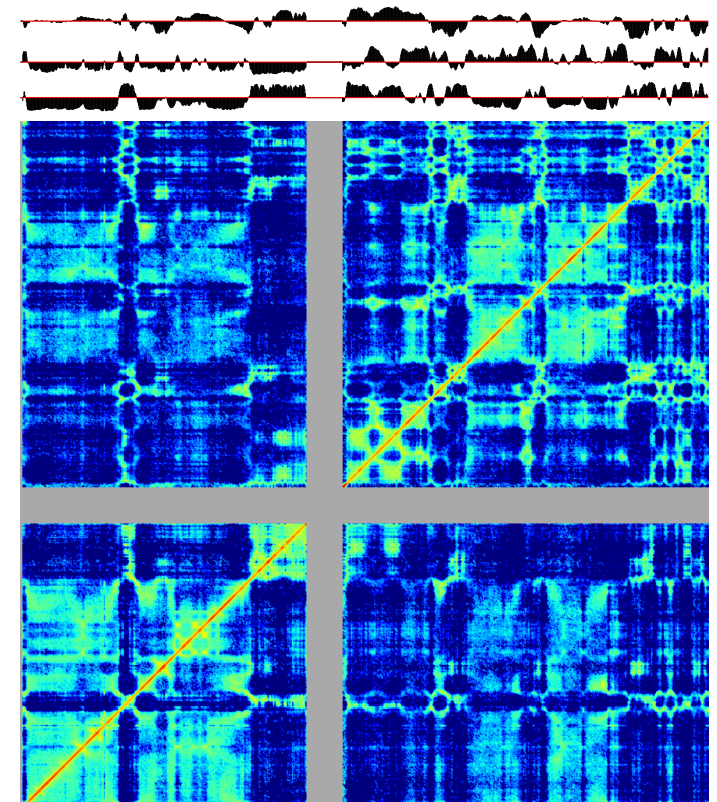
HiC experiment



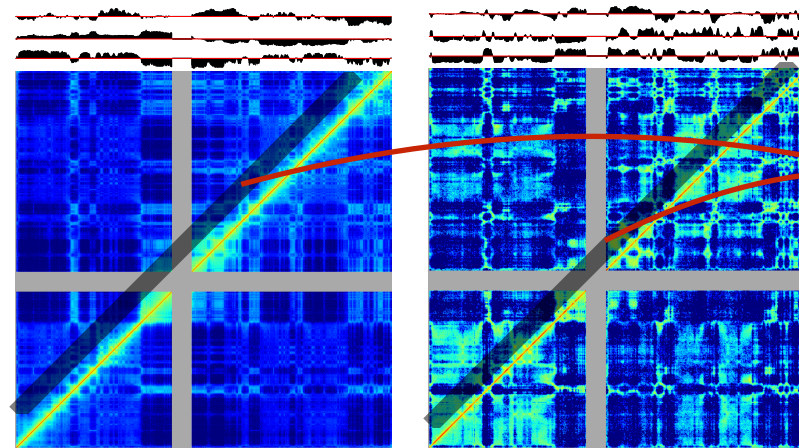
ENST00000434346.1



ENST00000541775.1



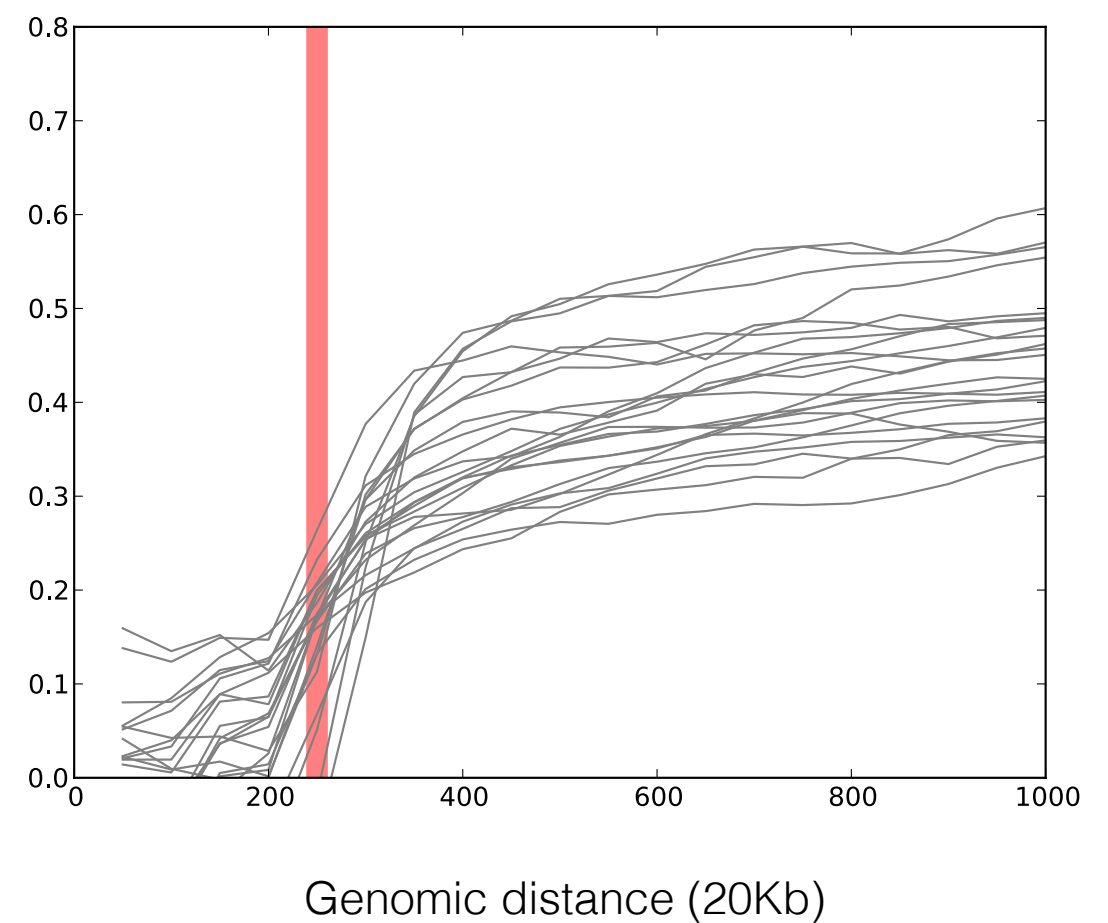
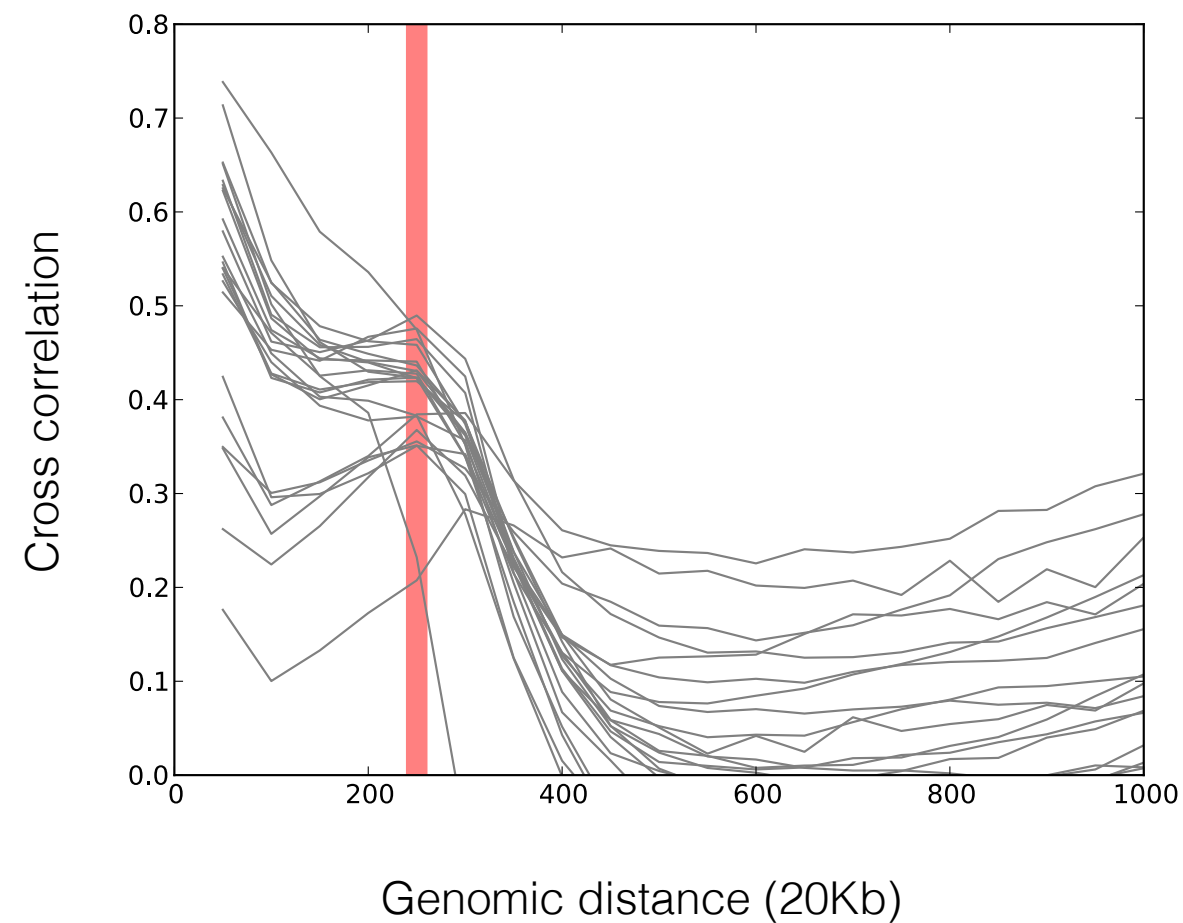
RNA & nuclear architecture



diagonal cross
correlation

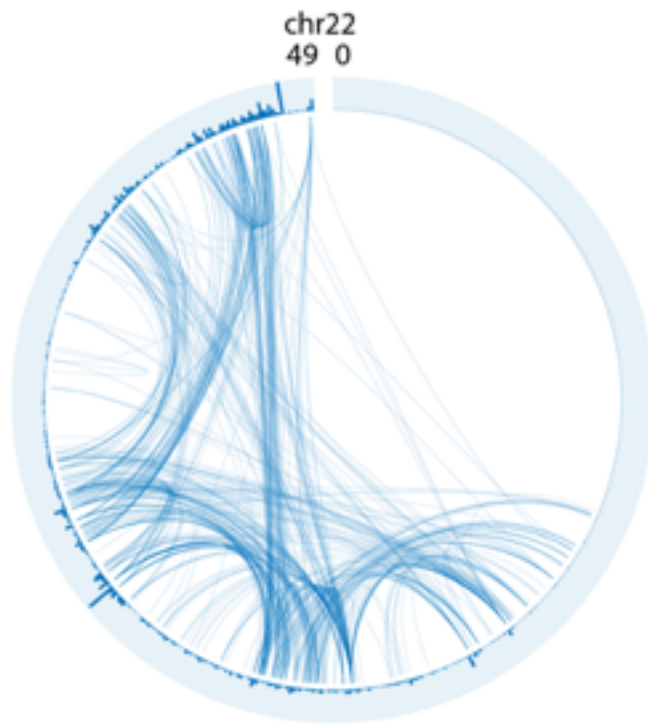
CTCF

ENST00000561611.2

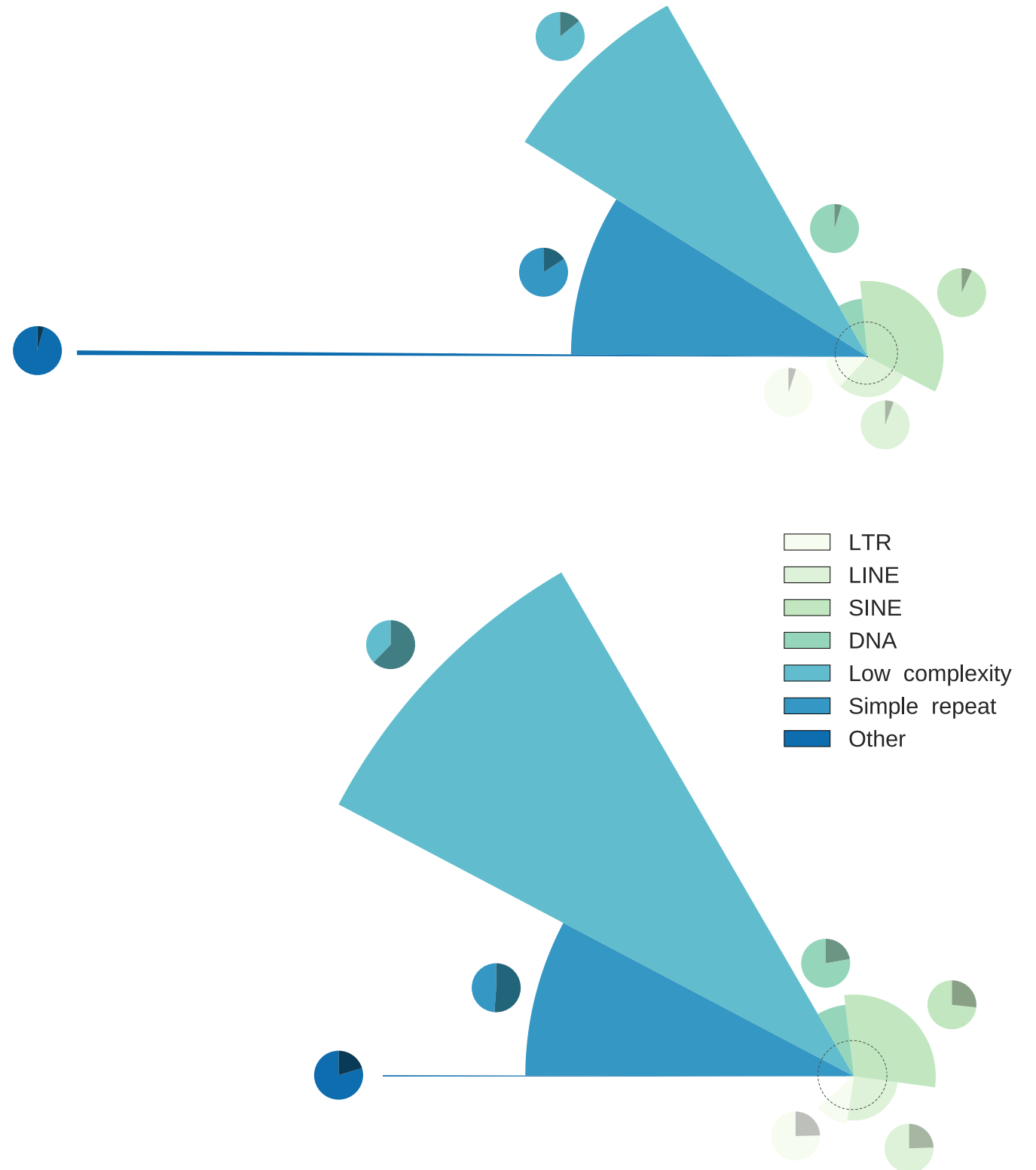
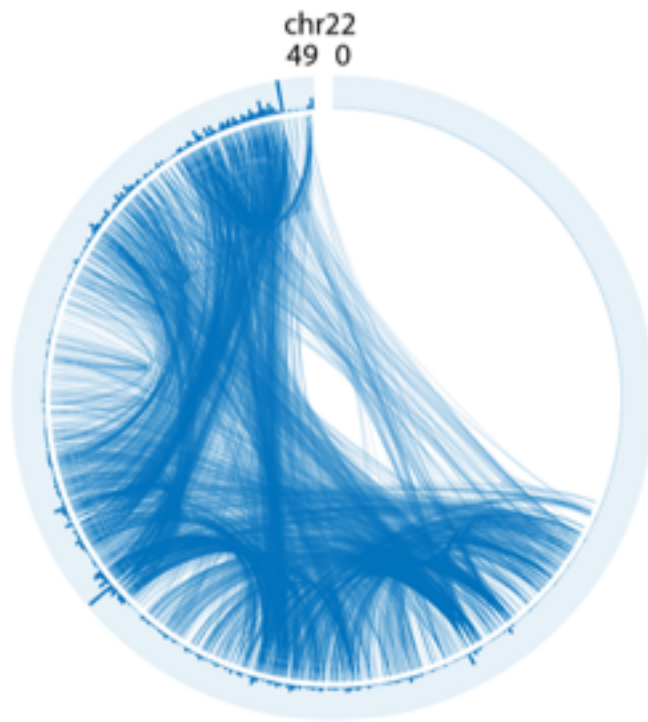


TTS and repetitive elements

ENST000000434346.1



ENST000000541775.1



Staple RNA to hold DNA?



<http://marciuslab.org>
<http://3DGenomes.org>
<http://cnag.crg.eu>



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François Serra
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Aleksandra Sparavier

In collaboration with Ralph Stadhouders (Erasmus MC) and Thomas Graf (CRG)

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