



3DGenomics

Marc A. Marti-Renom

CNAG-CRG · ICREA

<http://marciuslab.org>
<http://3DGenomes.org>
<http://cnag.crg.eu>

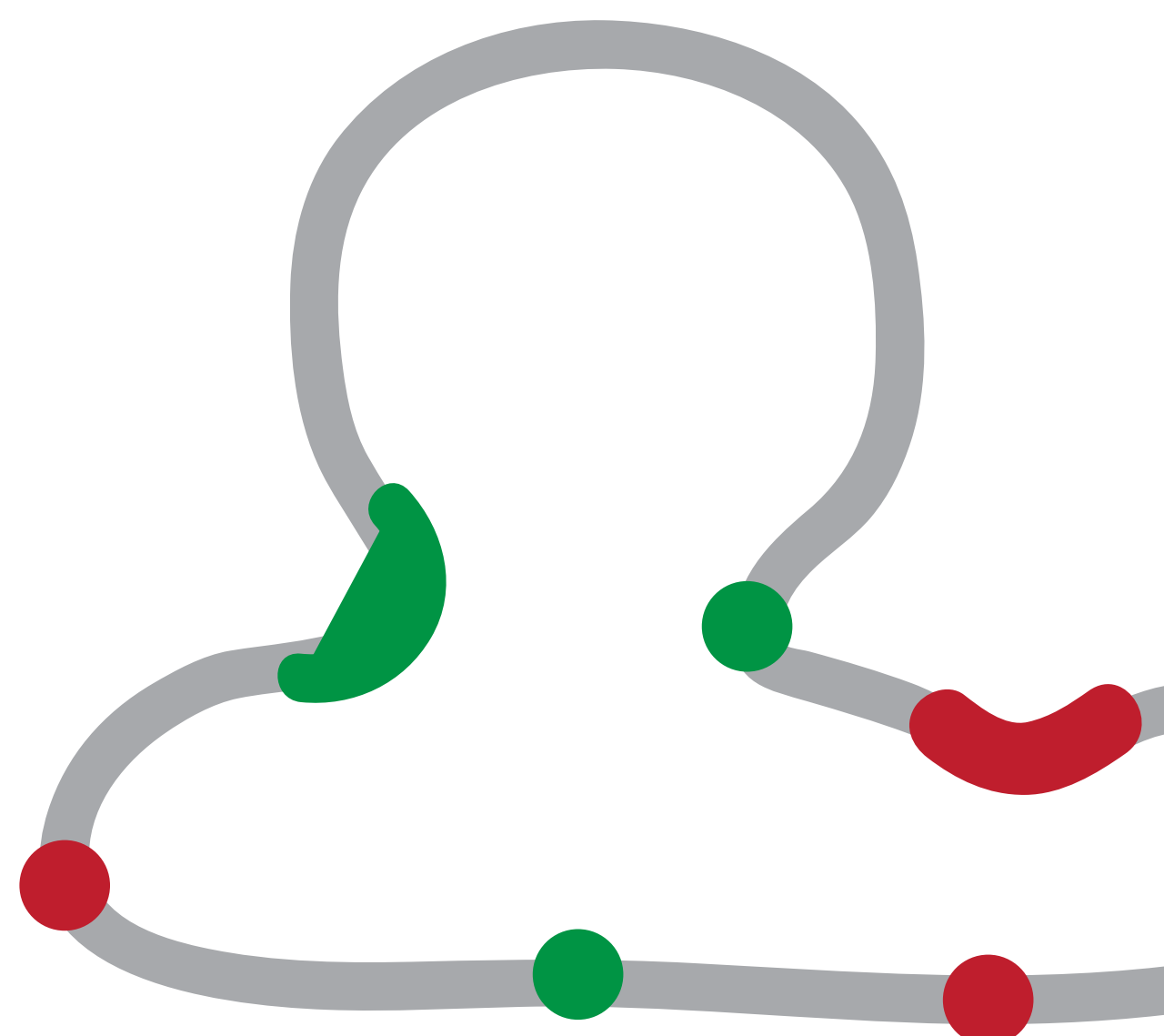
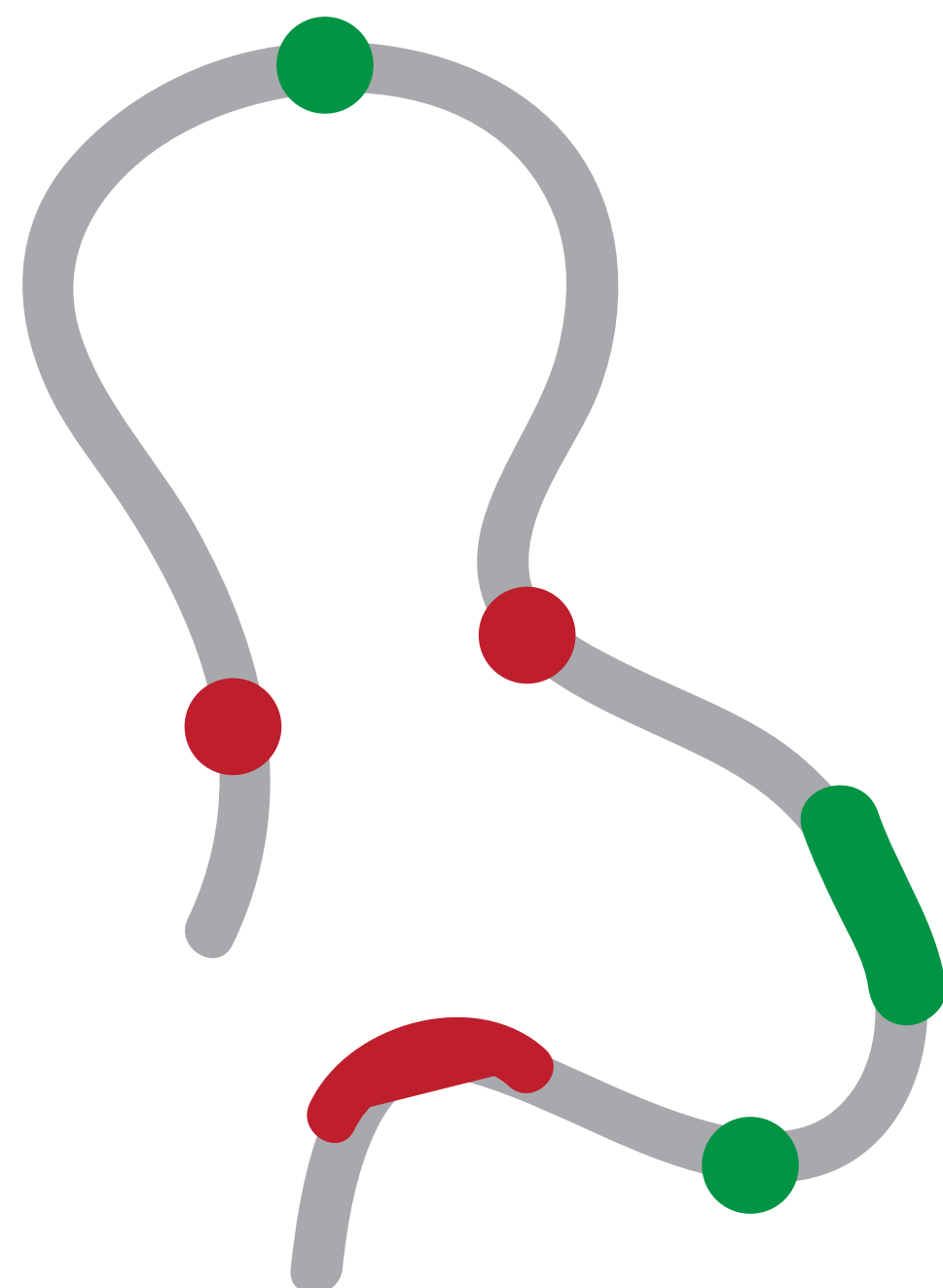
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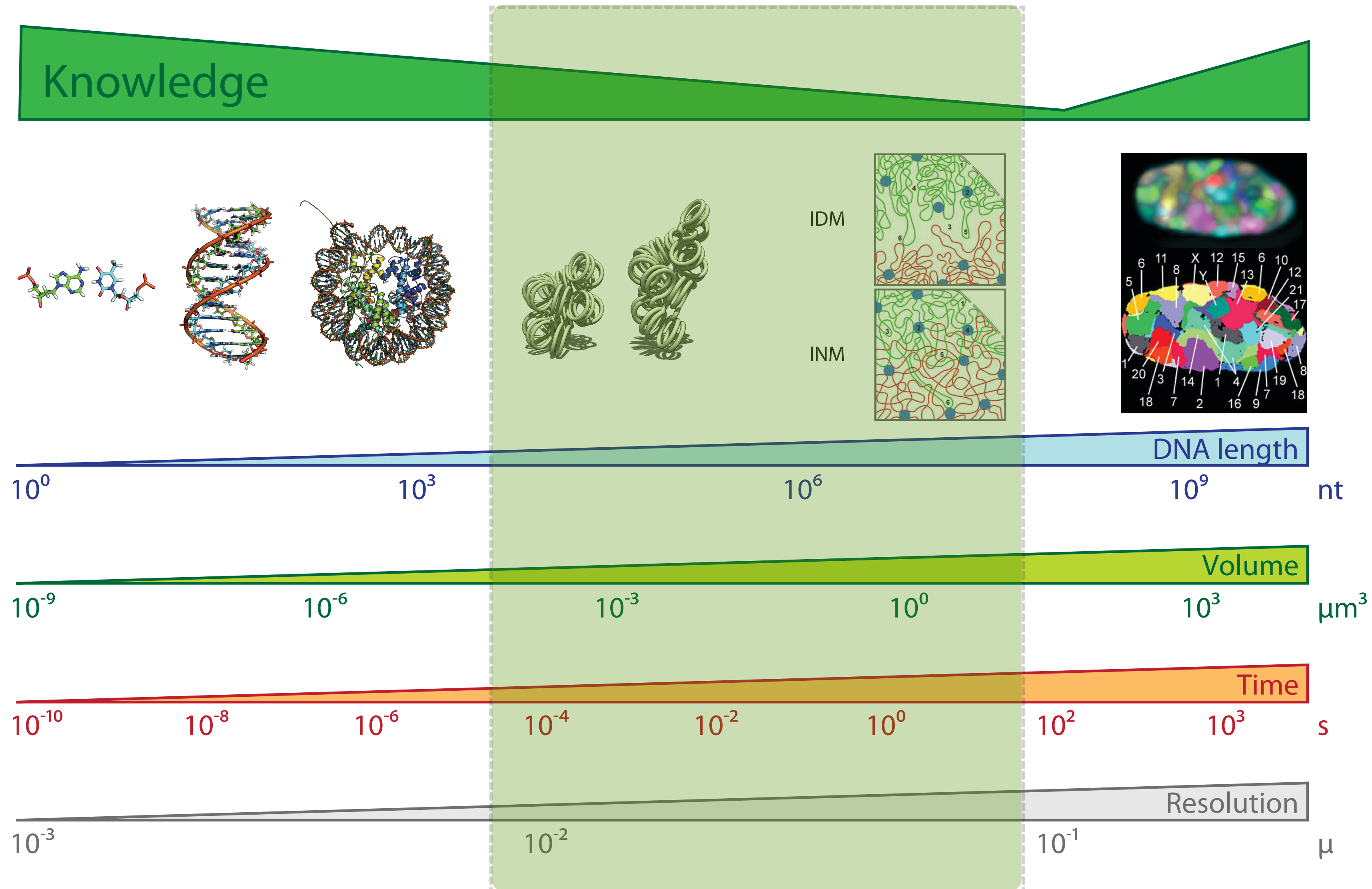
I encourage you to:

You can ask for question any time



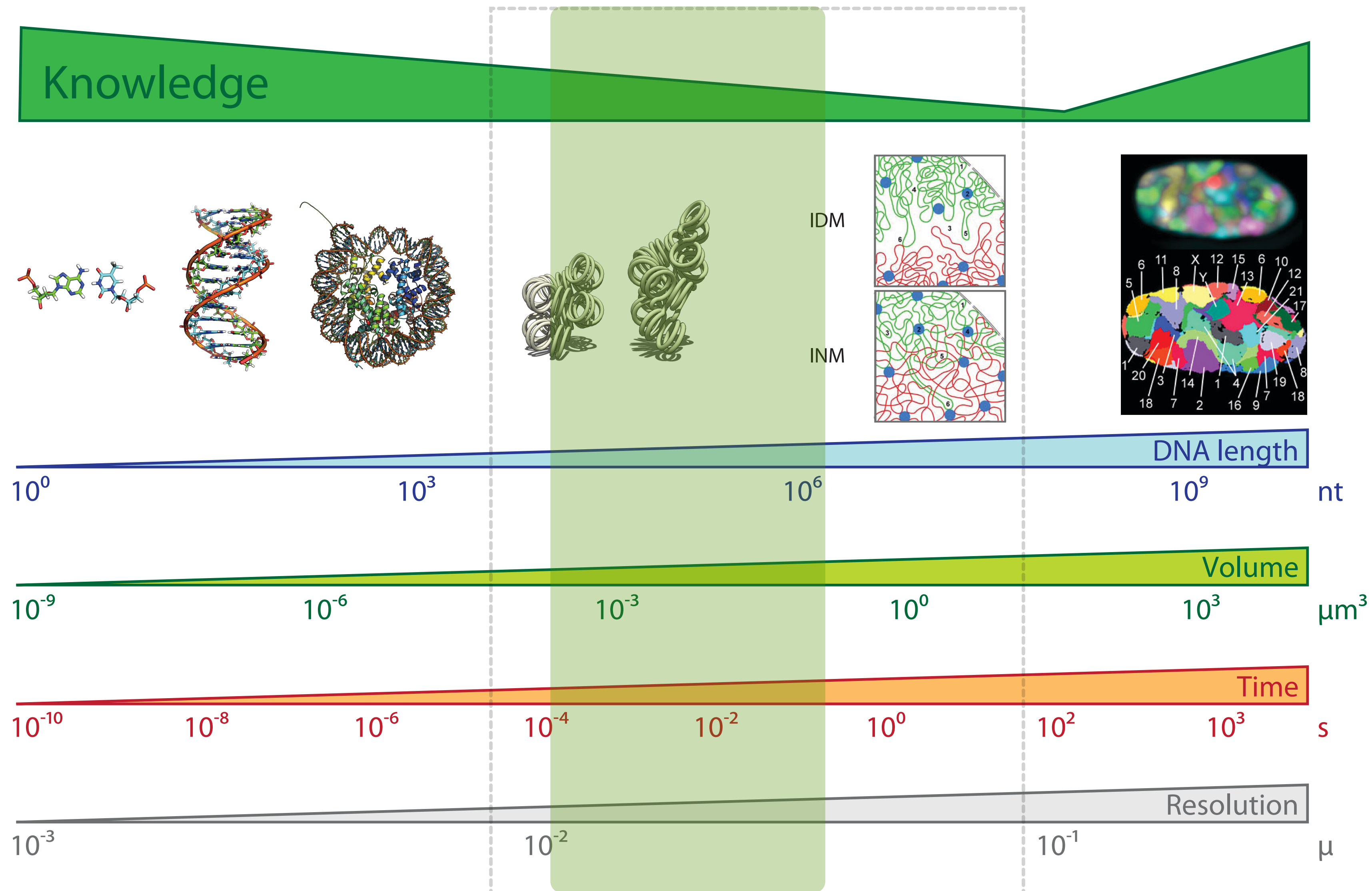
Resolution Gap

Marti-Renom, M. A. & Mirny, L. A. PLoS Comput Biol 7, e1002125 (2011)



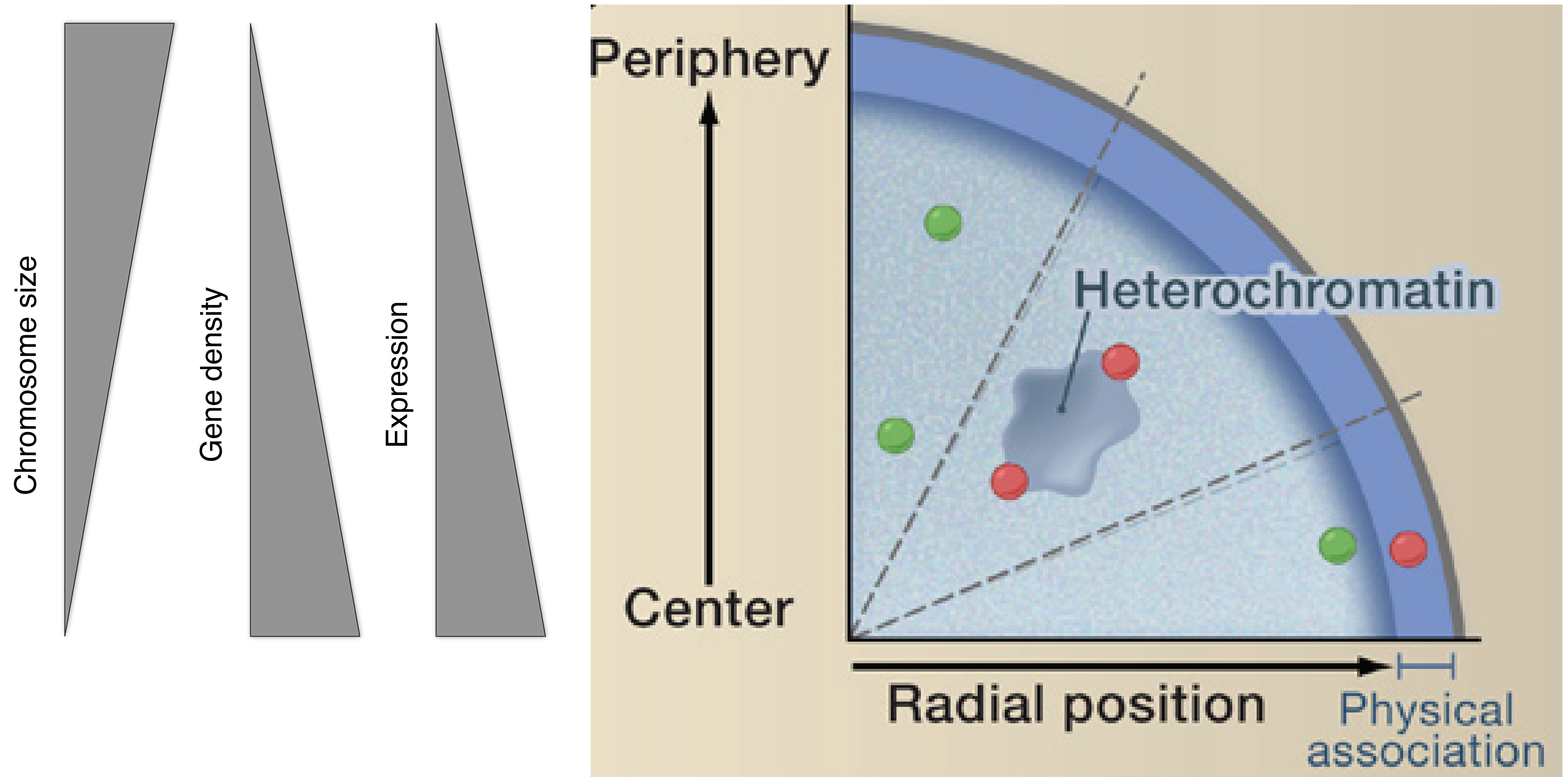
Resolution Gap

Marti-Renom, M. A. & Mirny, L. A. PLoS Comput Biol 7, e1002125 (2011)



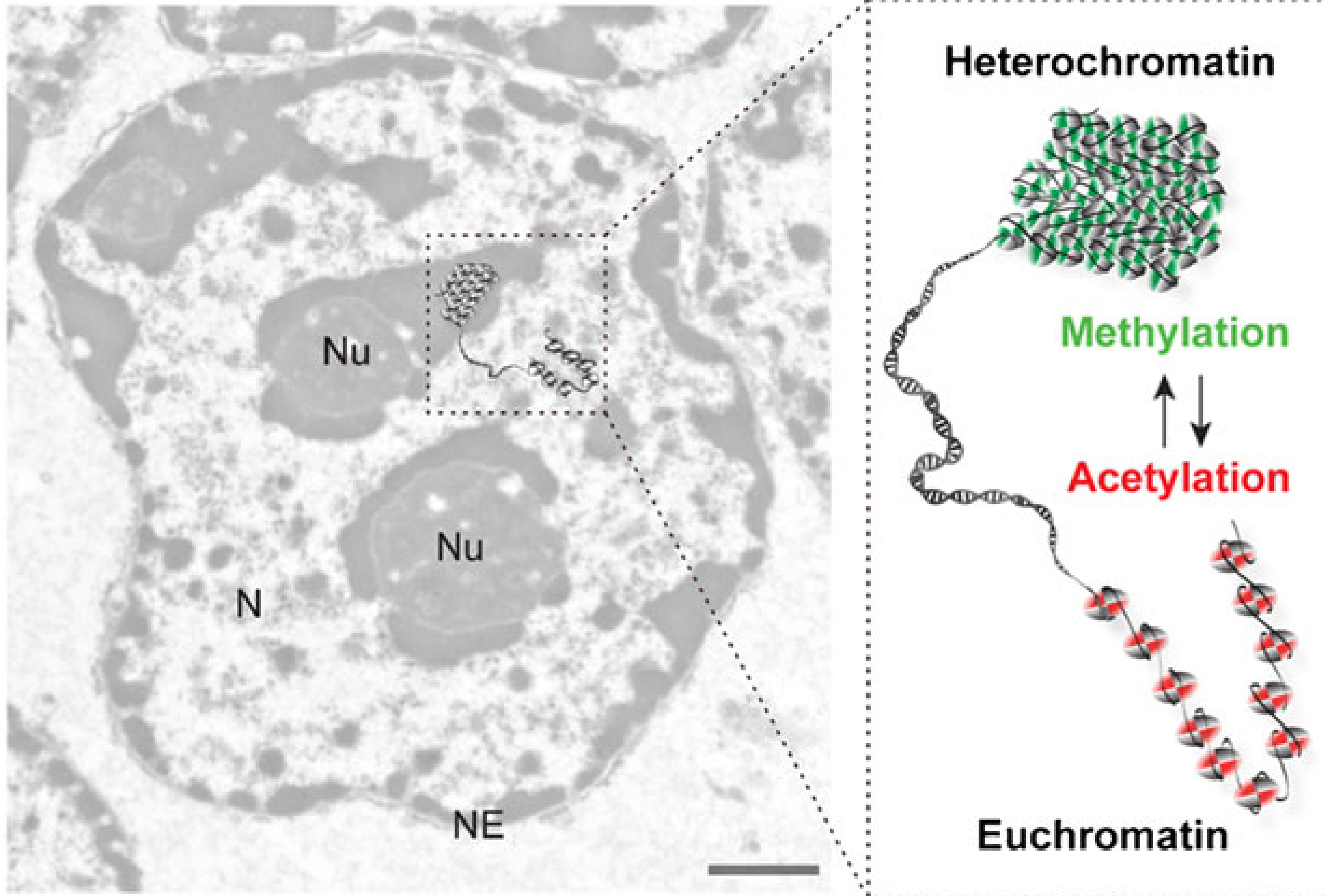
Level I: Radial genome organization

Takizawa, T., Meaburn, K. J. & Misteli, T. The meaning of gene positioning. Cell 135, 9–13 (2008).

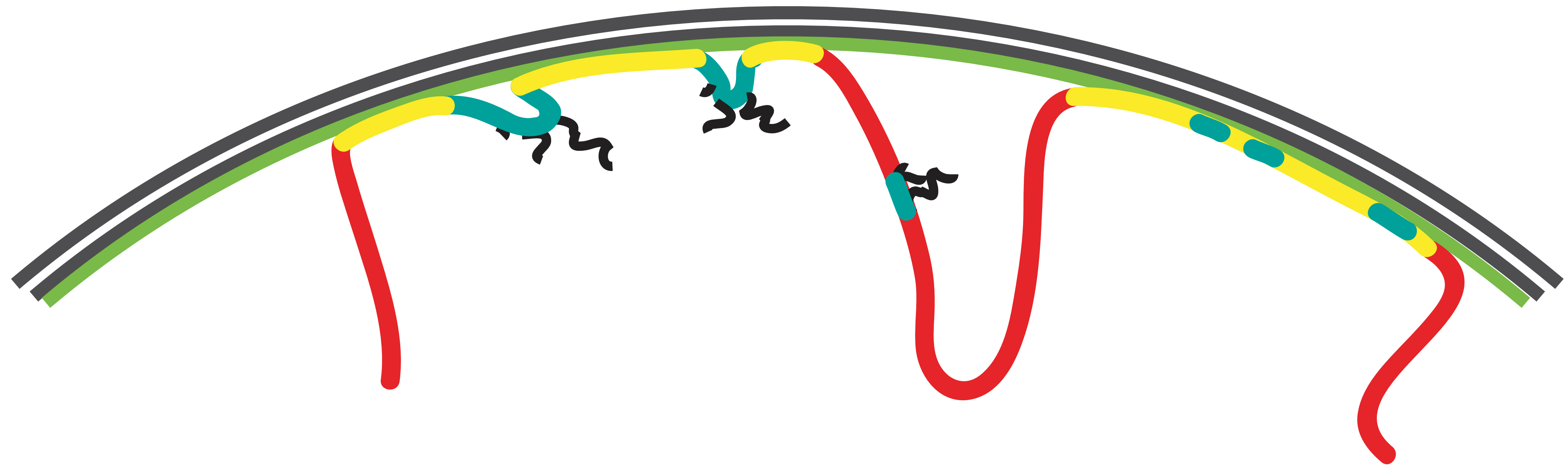


Level II: Euchromatin vs heterochromatin

Electron microscopy



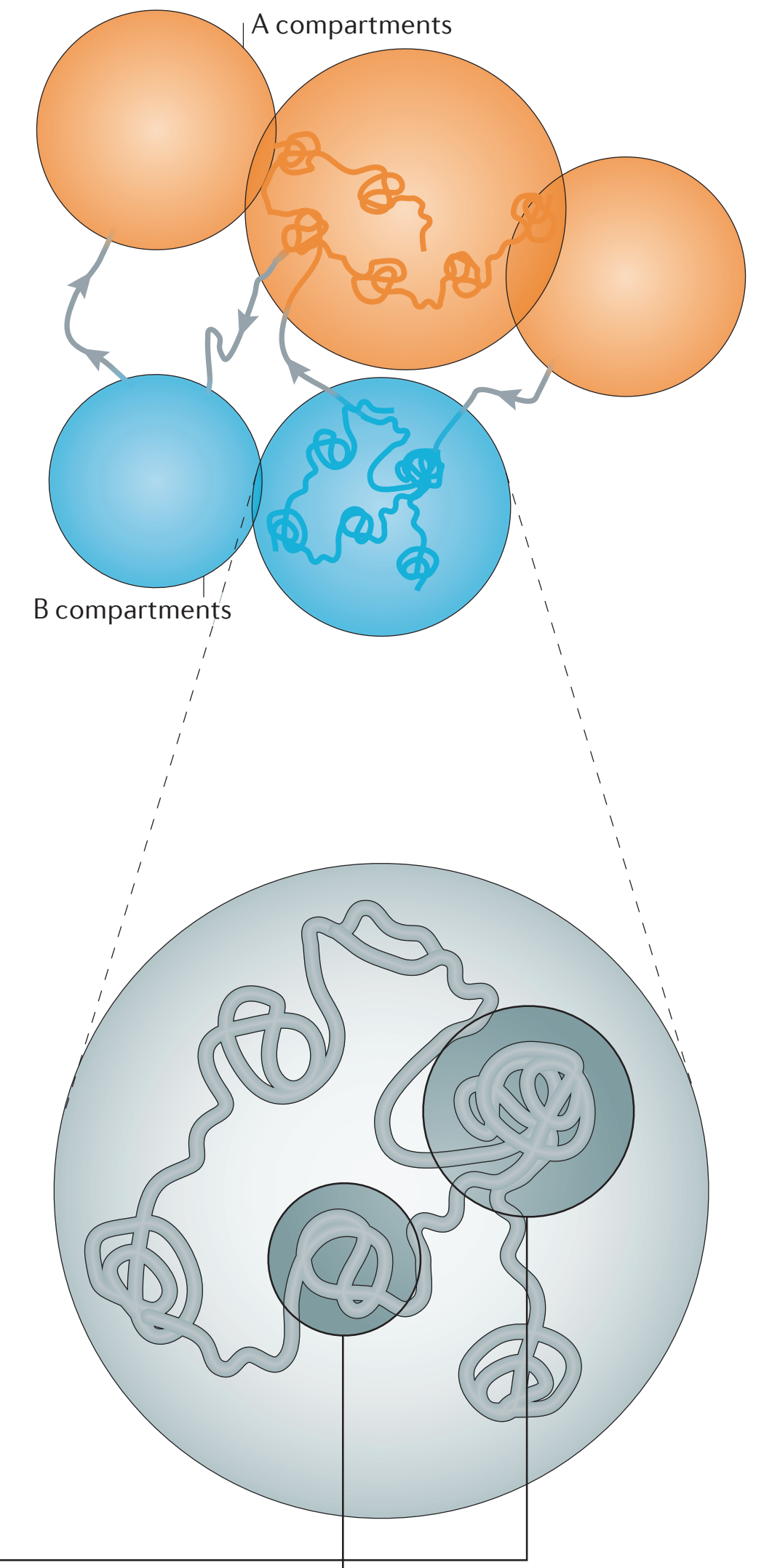
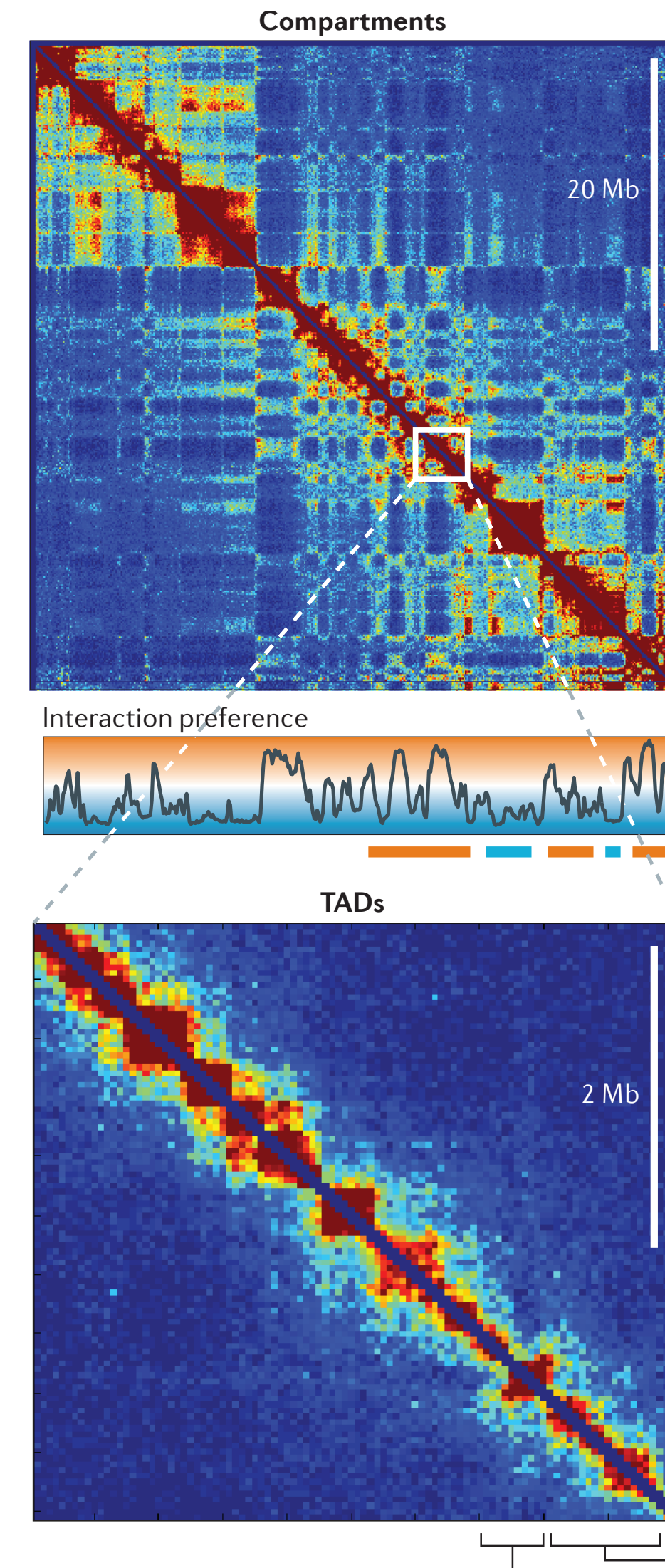
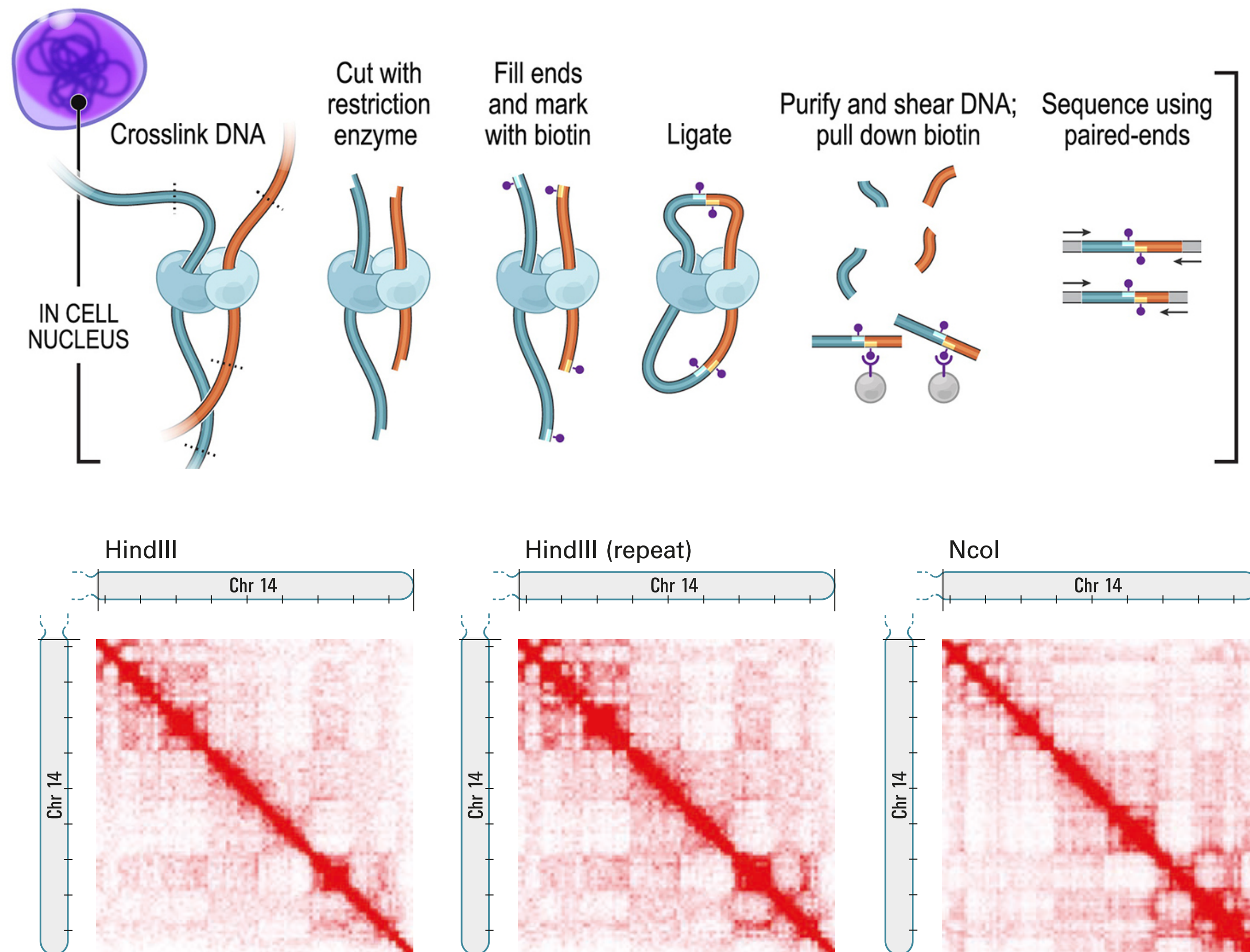
Level III: Lamina-genome interactions



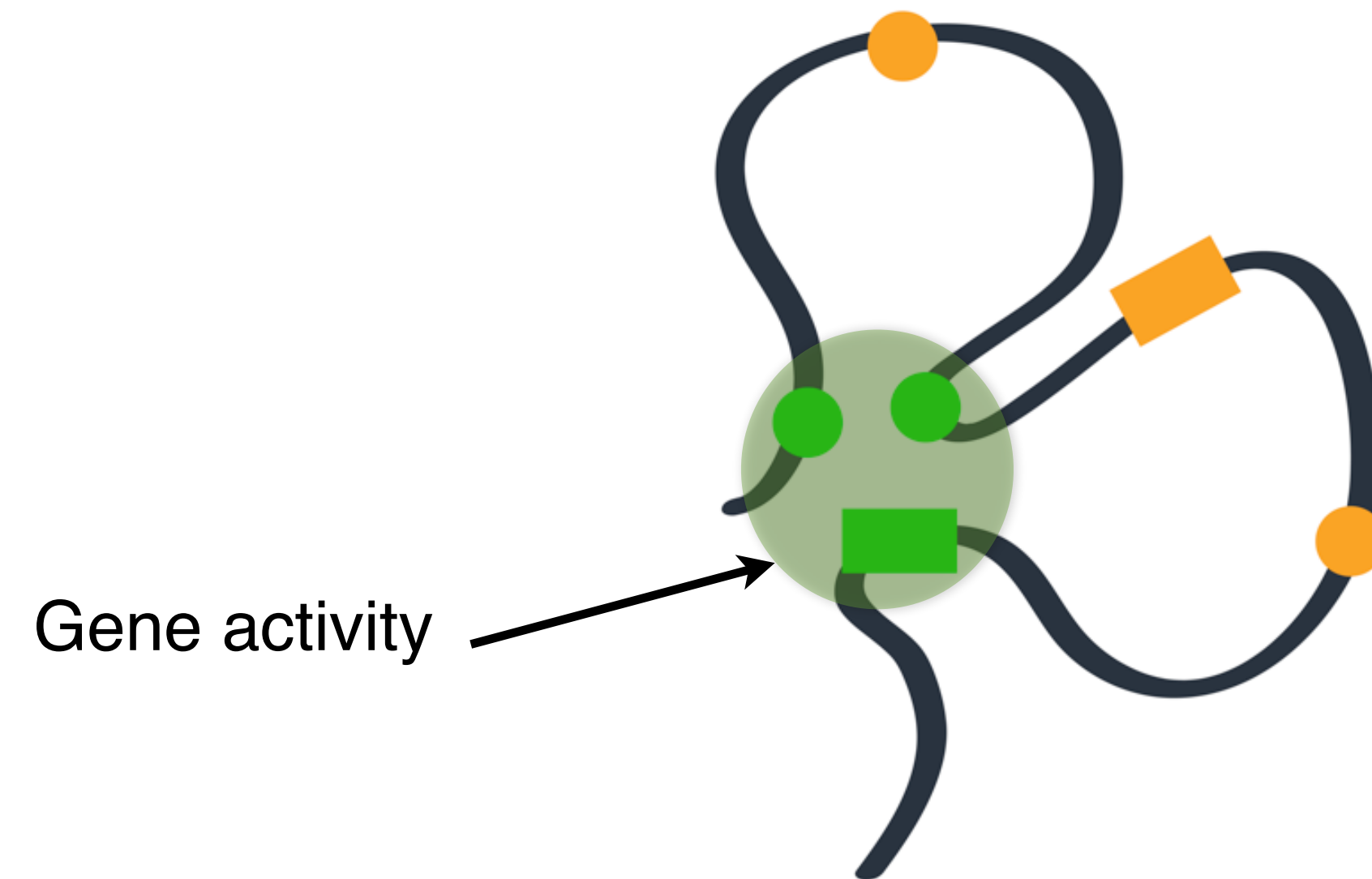
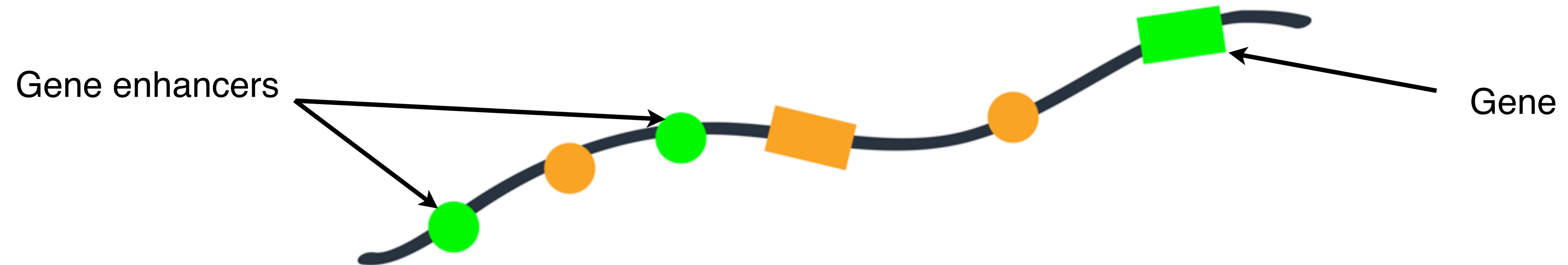
- nuclear membrane
- nuclear lamina
- internal chromatin (mostly active)
- lamina-associated domains (repressed)
- Genes
- mRNA

Level IV: Higher-order organization

Dekker, J., Marti-Renom, M. A. & Mirny, L. A. Nat Rev Genet 14, 390–403 (2013).



Level V: Chromatin loops

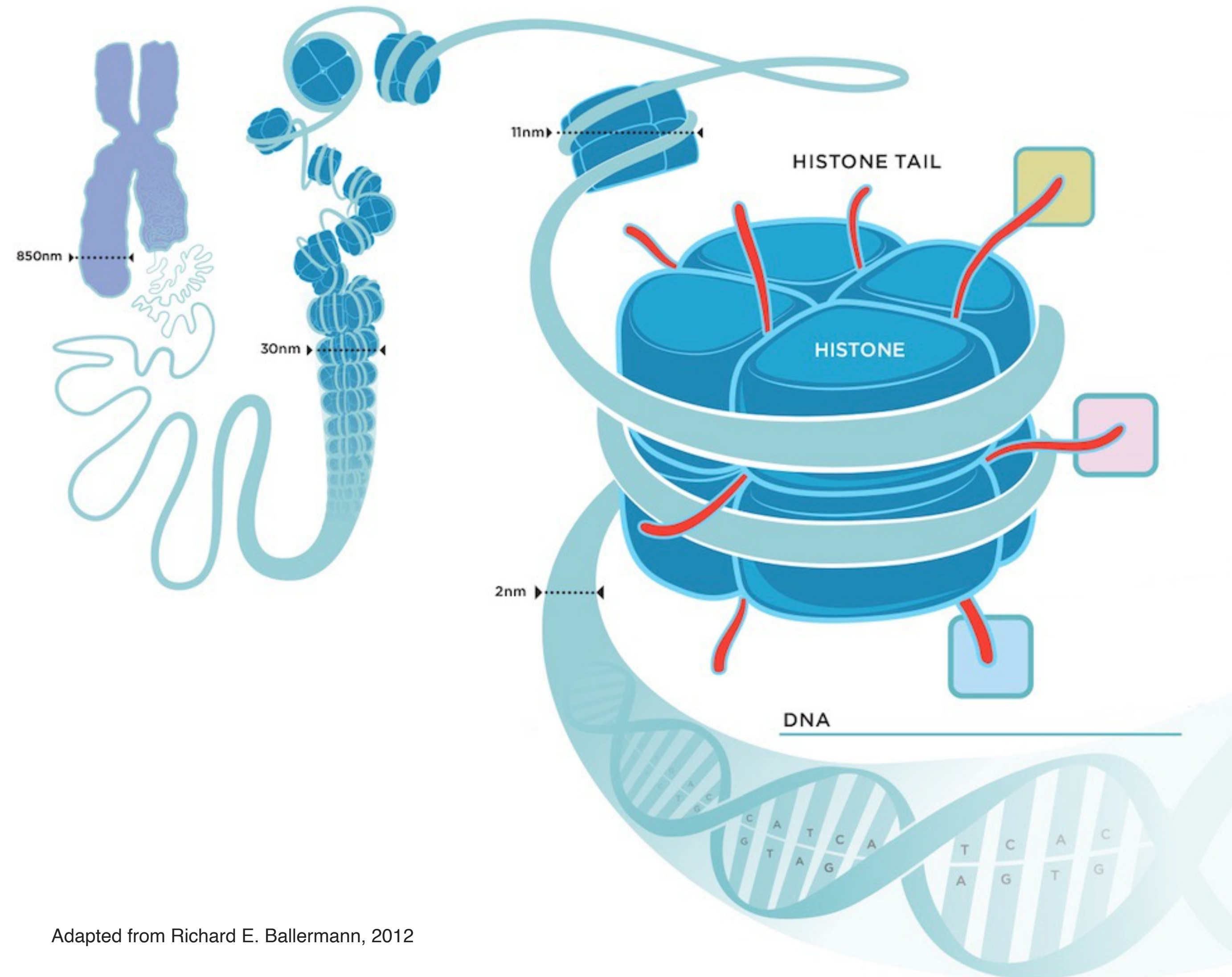


Level VI: Nucleosome

Chromosome

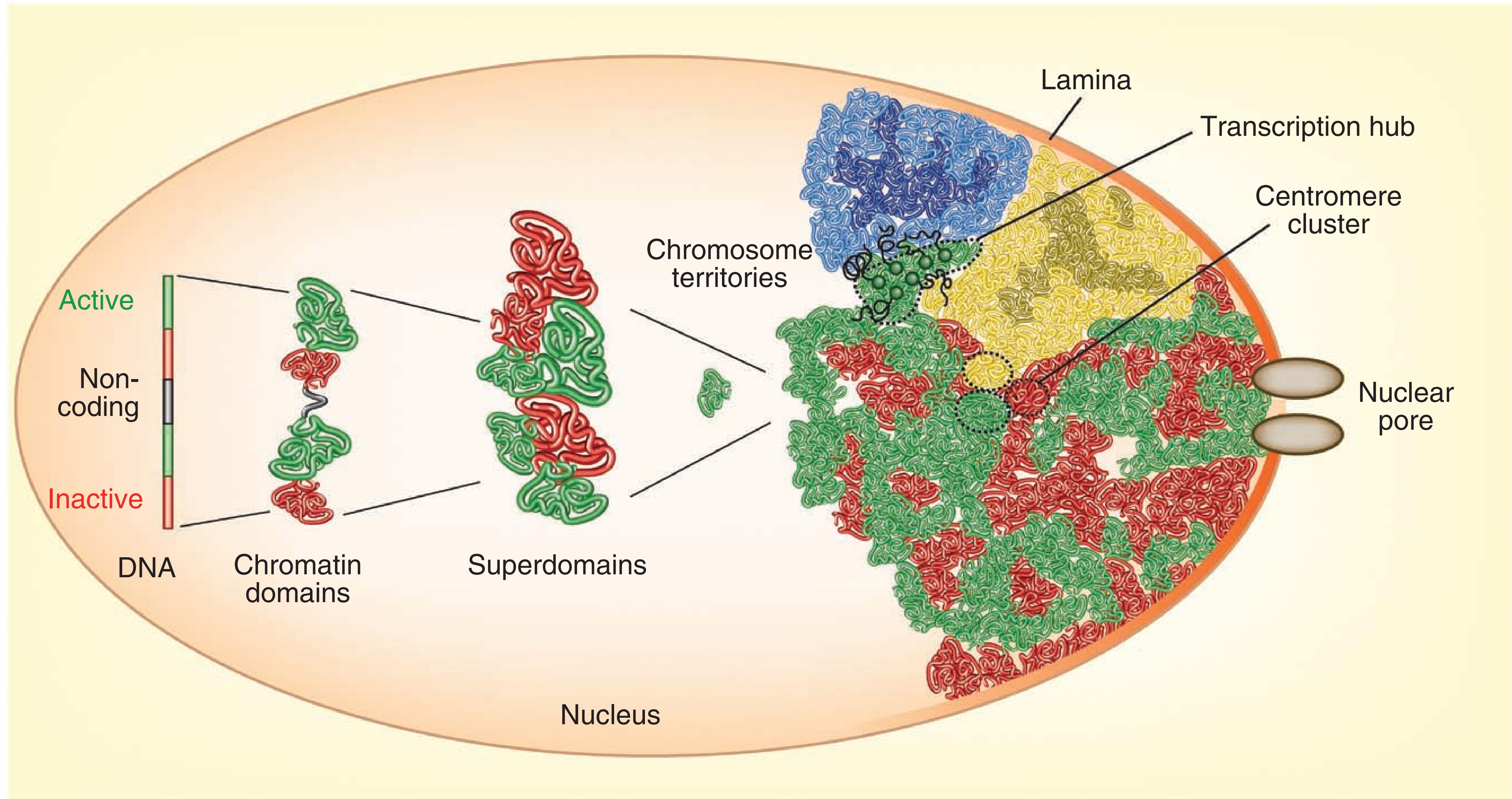
Chromatin fibre

Nucleosome

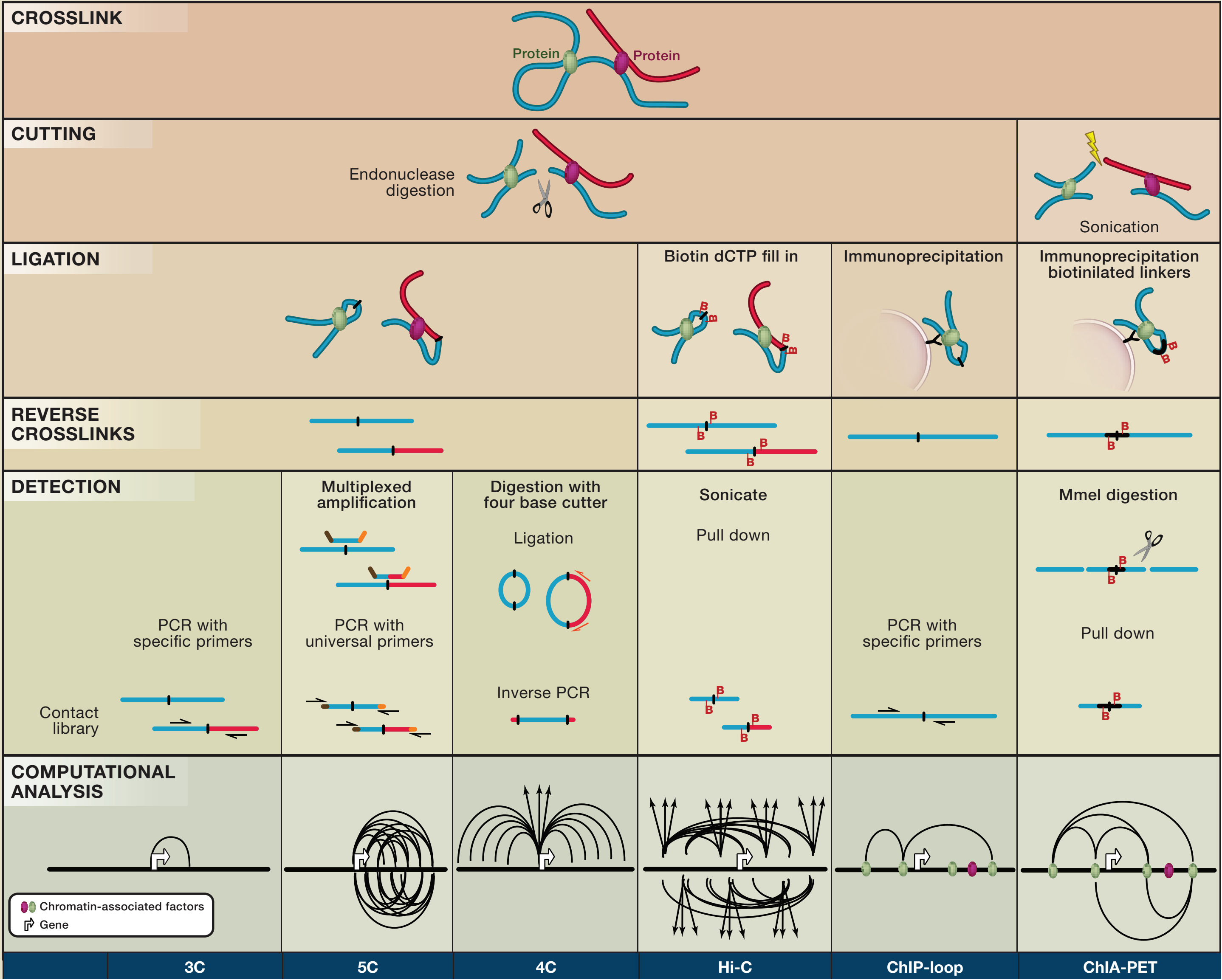


Complex genome organization

Cavalli, G. & Misteli, T. Functional implications of genome topology. Nat Struct Mol Biol 20, 290–299 (2013).



Chromosome Conformation Capture



ARTICLE doi:10.1038/nature12593

Single-cell Hi-C reveals cell-to-cell variability in chromosome structure

Takashi Nagano^{1*}, Yaniv Lubling^{2*}, Tim J. Stevens^{3*}, Stefan Schoenfelder¹, Eitan Yaffe², Wendy Dean⁴, Ernest D. Laue³, Amos Tanay² & Peter Fraser²

LETTER doi:10.1038/nature20158

Capturing pairwise and multi-way chromosomal conformations using chromosomal walks

Pedro Olivares-Chauvet¹, Zohar Mukamel¹, Aviezer Lifshitz¹, Omer Schwartzman¹, Noa Oded Elkayam¹, Yaniv Lubling¹, Gintaras Deikus², Robert P. Sebra² & Amos Tanay¹

nature genetics ARTICLES https://doi.org/10.1038/s41588-018-0161-5

Enhancer hubs and loop collisions identified from single-allele topologies

Amin Allahyar^{1,2,7}, Carlo Vermeulen^{3,7}, Britta A. M. Bouwman³, Peter H. L. Krijger³, Marjon J. A. M. Versteegen³, Geert Geeven³, Melissa van Kranenburg³, Mark Pieterse³, Roy Straver³, Judith H. I. Haarhuis⁴, Kees Jalink⁵, Hans Teunissen⁶, Ivo J. Renkens¹, Wigard P. Kloosterman¹, Benjamin D. Rowland⁴, Elzo de Wit⁵, Jeroen de Ridder^{3*} and Wouter de Laat^{3*}

Cell Resource

Higher-Order Inter-chromosomal Hubs Shape 3D Genome Organization in the Nucleus

Graphical Abstract

Authors: Sofia A. Quinodoz, Noah Ollikainen, Barbara Tabak, ..., Patrick McDonel, Manuel Garber, Mitchell Guttman

Correspondence: mguttman@caltech.edu

ARTICLE DOI: 10.1038/s41467-018-06961-0 OPEN

Chromatin conformation analysis of primary patient tissue using a low input Hi-C method

Noelia Díaz¹, Kai Kruse¹, Tabea Erdmann², Annette M. Staiger^{3,4,5}, German Ott³, Georg Lenz² & Juan M. Vaquerizas¹

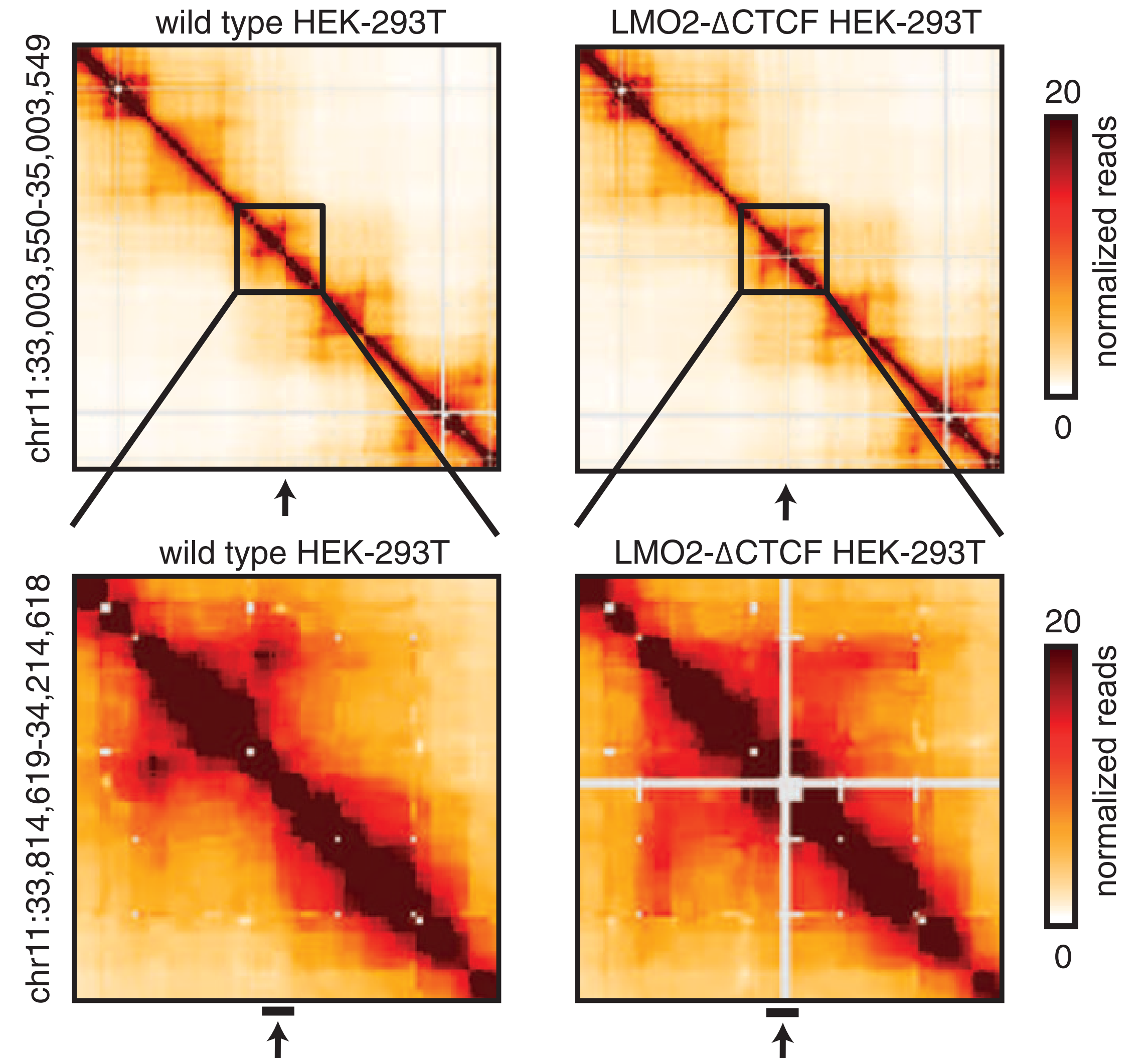
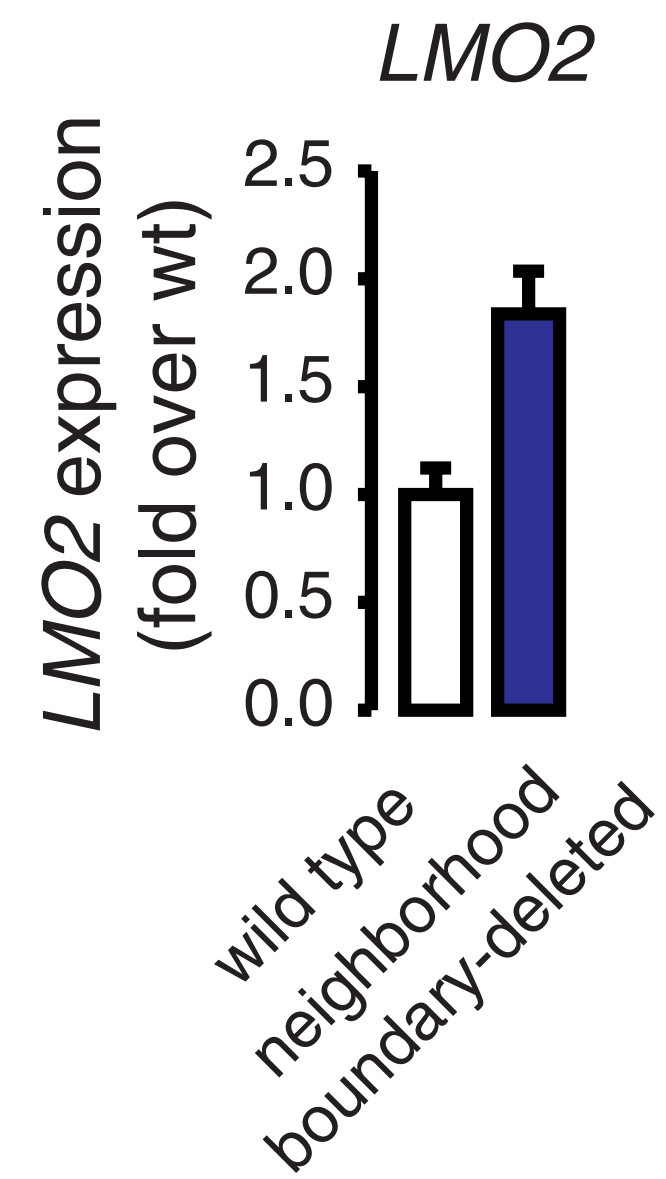
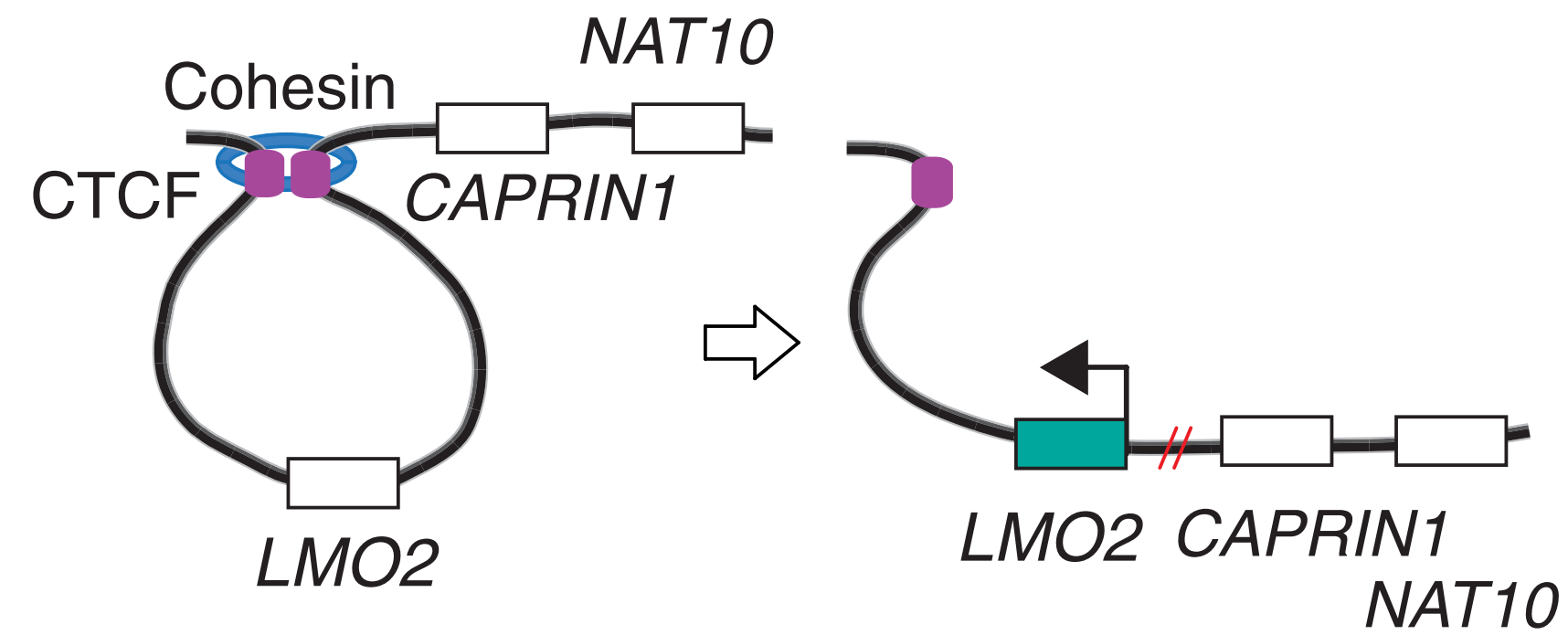
Liquid chromatin Hi-C characterizes compartment-dependent chromatin interaction dynamics

Houda Belaghzal, Tyler Borrmann, Andrew D. Stephens, Denis L. Lafontaine, Sergey V. Venev, Zhiping Weng, John F. Marko & Job Dekker

Nature Genetics 53, 367–378 (2021) | Cite this article
7436 Accesses | 8 Citations | 20 Altmetric | Metrics

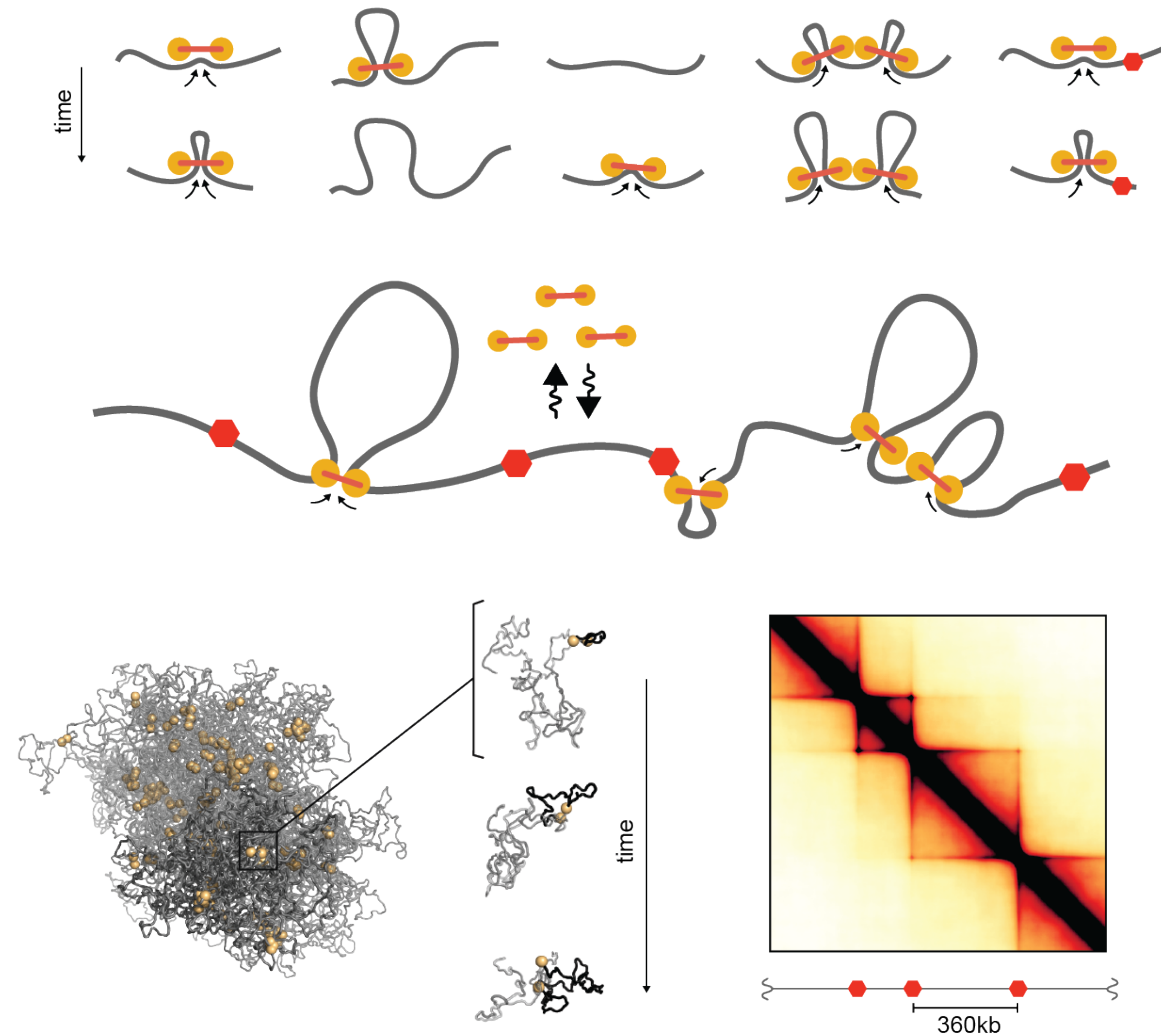
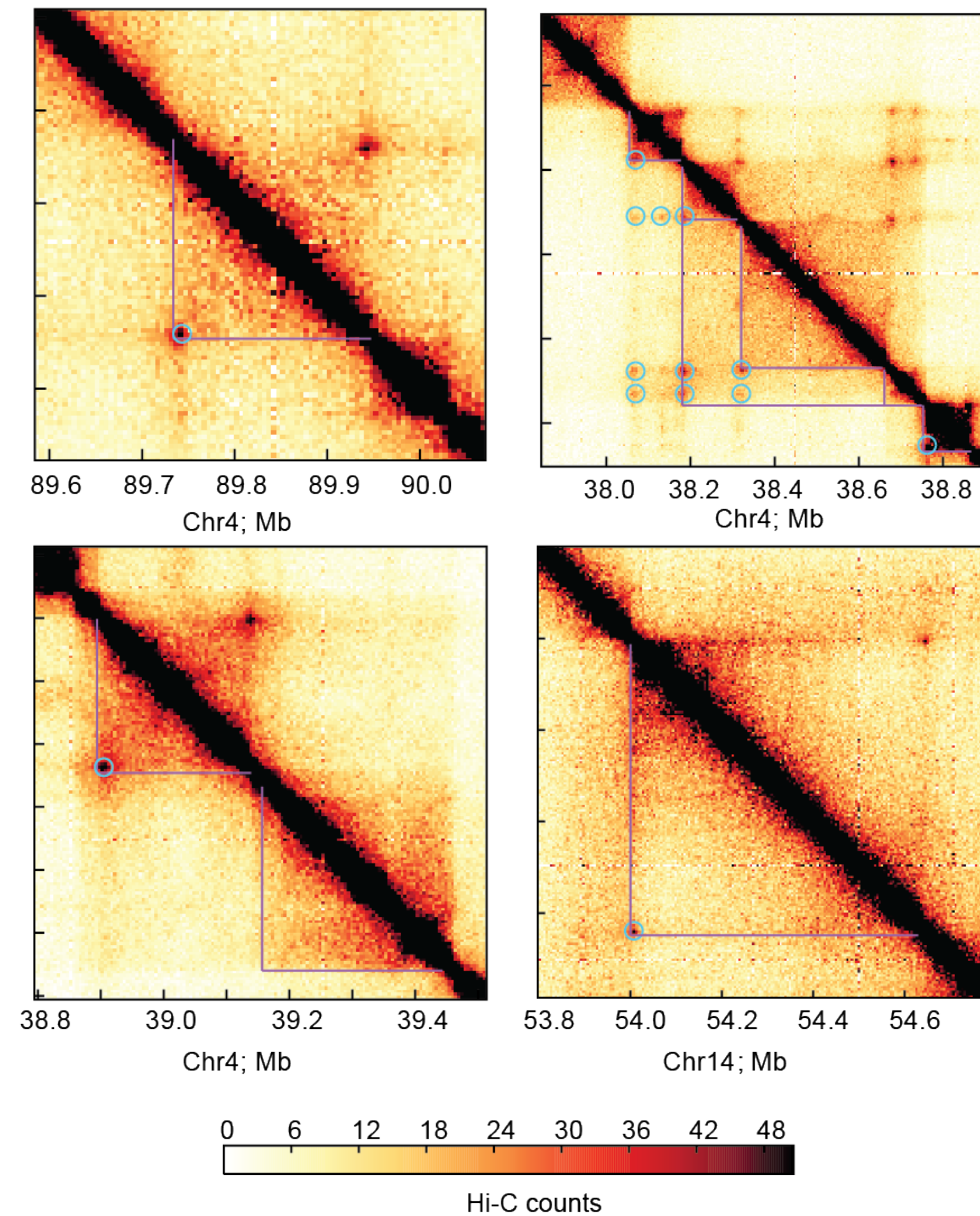
TADs are functional units

Hnisz, D., et al. (2016). Science



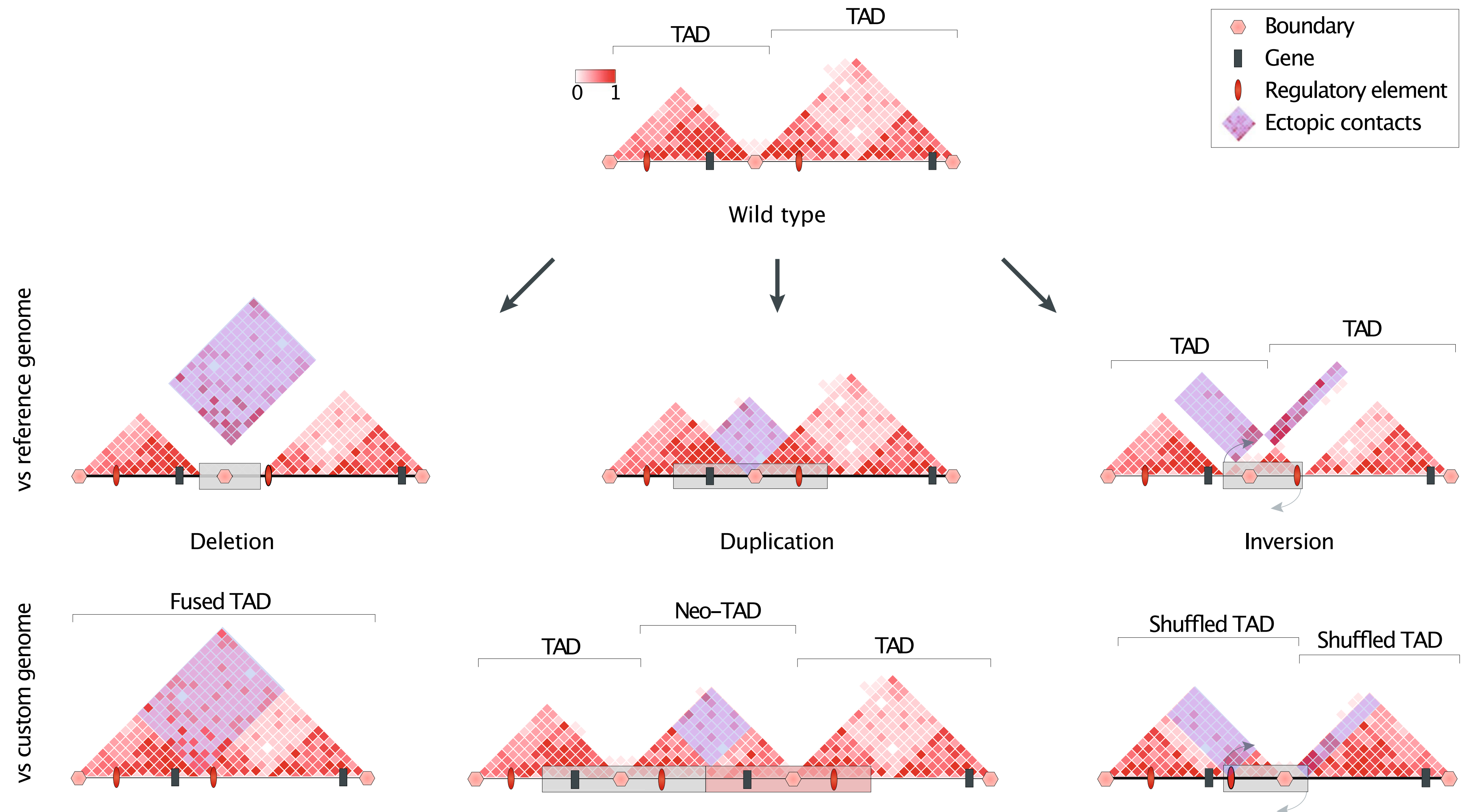
Loop-extrusion as a TAD forming mechanism

Fudenberg, G., Imakaev, M., Lu, C., Goloborodko, A., Abdennur, N., & Mirny, L. A. (2018).
Cold Spring Harb Symp Quant Biol 2017. 82: 45-55



Are TADs functional units?

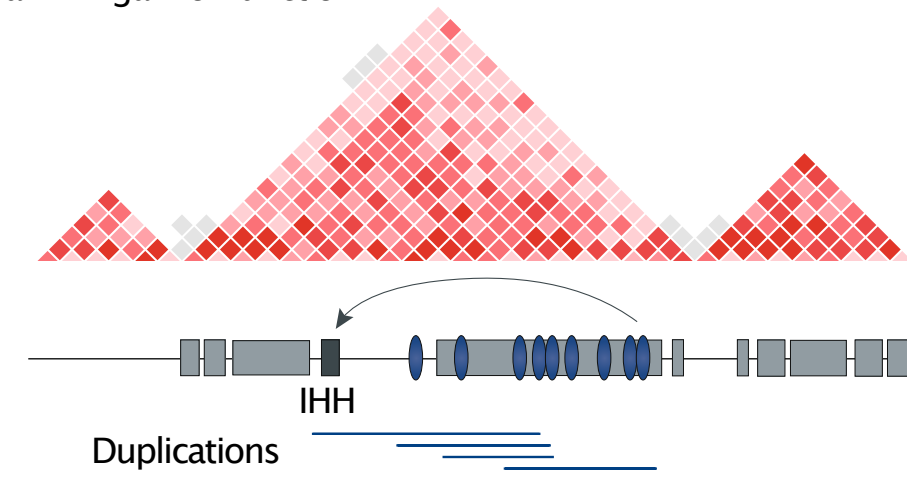
Spielmann Nature Reviews Genetics 2018 (19) 453–467



Clinical examples of structural variants

Spielmann Nature Reviews Genetics 2018 (19) 453–467

Intra-TAD gain of function



Phenotype

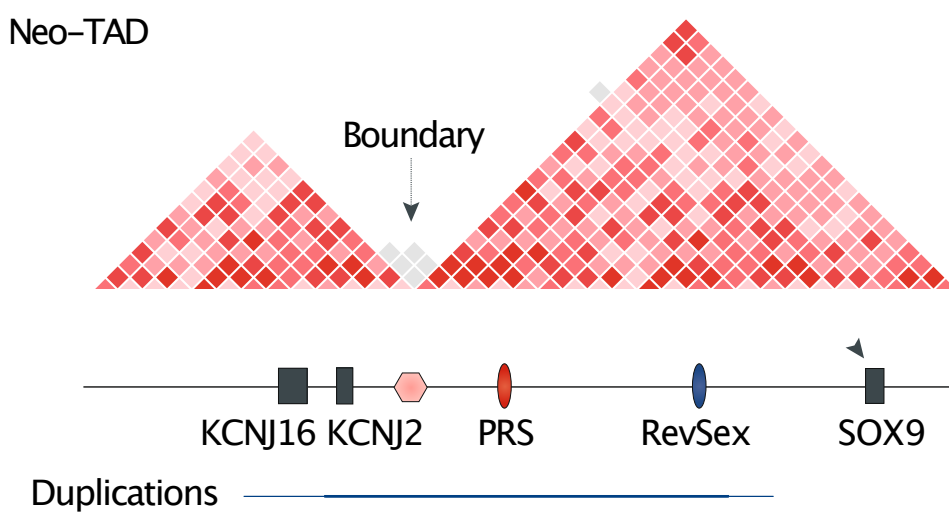


Duplications of enhancer elements cause preaxial synpolydactyly of feet

Examples

- Gain of function:
- SOX9 locus: duplications of gonad enhancer cause 46,XX sex reversal
 - BCL6 locus: duplications of super enhancers cause B cell lymphomas
 - SHH locus: duplications of limb enhancer causes polydactyly
- Loss of function:
- PAX6 locus: aniridia
 - DLX5 and/or DLX6 loci: split hand foot malformation
 - SOX9 locus: deletions of gonad enhancer cause 46,XY sex reversal

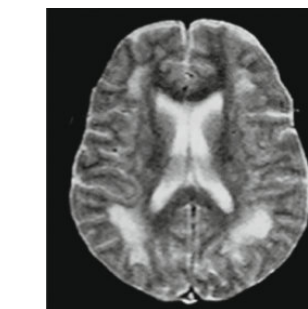
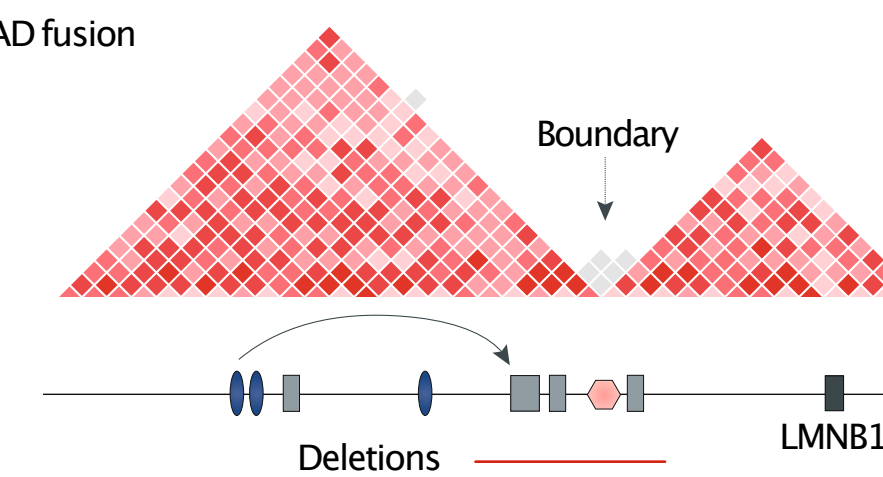
Neo-TAD



Cooks syndrome: Duplications of TAD boundary, KCNJ2 and KCNJ16 cause aplasia of nails and short digits

- FGF2 locus: colorectal cancer
- PRDM6 locus: medulloblastoma

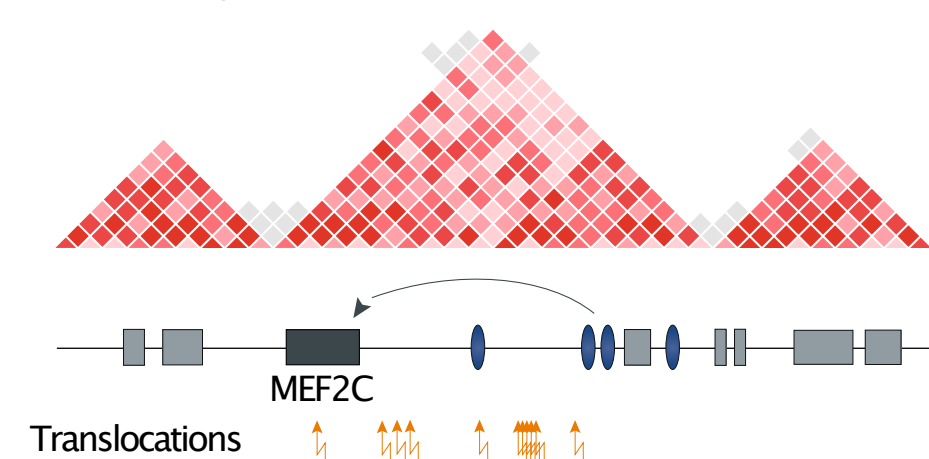
TAD fusion



Adult-onset demyelinating leukodystrophy

- GF11 locus: medulloblastoma
- TAL1 and LMO2 loci: T cell acute lymphoblastic leukaemia
- IRS4 locus: lung squamous carcinoma, sarcoma and cervical squamous carcinoma
- SOX4 locus: mesomelic dysplasia

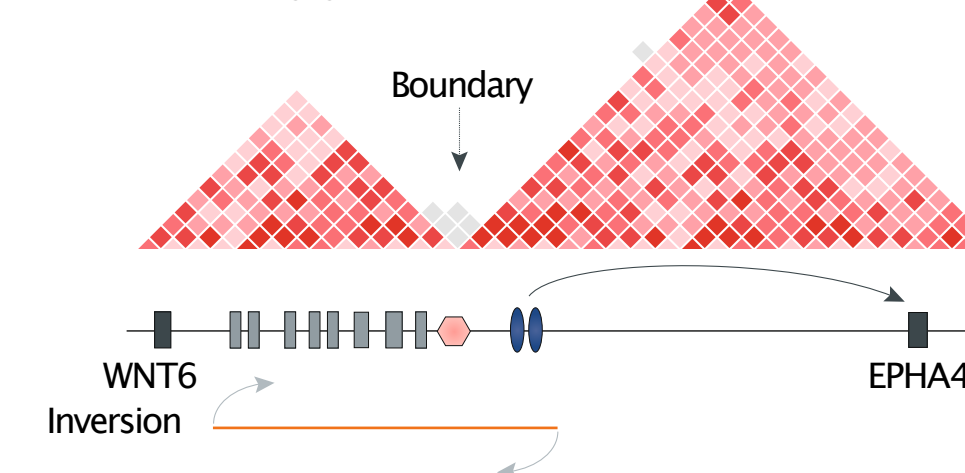
TAD shuffling: loss of function



Hypoplastic corpus callosum via loss of function of MEF2C at 5q14.3

- FOXP1 locus: atypical Rett syndrome
- SOX9 locus: campomelic dysplasia
- DLX5 and DLX6 loci: split hand foot malformation

TAD shuffling: gain of function



F-syndrome: syndactyly

- SHH locus: inversion of enhancer causes short digits (Dsh mouse model)
- SHH locus: inversion of enhancer causes polysyndactyly
- GF11 locus: medulloblastoma
- Translocation at the PITX1 locus: Liebenberg syndrome

■ Genes ● Regulatory elements ⬡ Boundaries

Alexander Barclay
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